

Review Article

Synthesis Strategies and Potential Applications of Gold Nanoparticles in Cancer Theranostics: Labelling and Visualizing, Targeted Drug Delivery, Photoablation Therapy and Sensing

Adena SKR, Upadhyay M, Vardhan H and Mishra B*

Department of Pharmaceutical Engineering & Technology, Indian Institute of Technology (BHU), India

*Corresponding authors: Mishra B, Department of Pharmaceutical Engineering & Technology, Indian Institute of Technology (BHU), Varanasi-221 005, UP, India

Received: October 05, 2018; Accepted: November 03, 2018; Published: November 10, 2018

Abstract

Nanotechnology has transformed into a champion amongst the furthestmost exciting and cutting-edge areas of research in the field of biomedicine. Gold Nanoparticles (GNPs) exhibit exceptional advantages when compared to other nanoparticles, mainly because of its unique properties. GNPs have widely used in quite a lot of cancer theranostic applications due to their very small size, inert nature, biocompatibility, high dispersity, noncytotoxicity, stability, larger surface area, tunable physical, chemical, optical and electronic properties. Biocompatible polymers are being utilized for the surface modification of GNPs to increase the therapeutic payload and also the stability of the GNPs, leading to enhanced systemic circulation and efficient cellular uptake of the GNPs in the cancer theranostics. The main purpose of the present review is to illustrate different synthesis methods viz. chemical, physical, green methods and also the theranostic applications of GNPs. The fundamental ideas and the mechanism involved in each of these applications, the main features of the GNPs required for each of these applications and additionally, a few examples are portrayed.

Keywords: Cancer; Gold nanoparticles; Theranostic; Targeted drug delivery; Polymers

Introduction

Over the past half century, chemotherapy has extensively enhanced the cancer treatment. But sadly, lack of selectivity for the conventional chemotherapeutic agents, less than one percent of the drugs were taken up by the tumor cells and its microenvironment, the remaining drug was going to the surrounding healthy tissues [1]. As chemotherapeutic agents are usually intended for a specific site in the body, this conventional method of delivering the drug is devoid of efficiency and need a significant amount of drug thereby leading to adverse side effects to the surrounding healthy systems. Multi Drug Resistance (MDR) is another major problem that is associated with the failure of chemotherapy. MDR results in low intracellular drug concentration in the tumor cells due to restraintment of efficient drug accumulation thereby resulting in a low response. Another major problem associated with the failure of chemotherapy is due to the differences in the tumor cells' chemo sensitivity, resulting in the difference in the tumor cell's percentage of response. Hence developing drug delivery systems that are efficacious selectively is one of the ultimate challenges confronting chemotherapy at present and that can be possible by Gold Nanoparticles (GNPs) [2].

GNPs are available in different shapes and sizes, and their unique properties make them suitable for various applications as shown in (Figure 1). As GNPs have a large surface area, their surfaces can be accessible to further alteration with hydrophilic, hydrophobic, anionic, cationic and neutral moieties, so that their applications can be drawn out to a further extent. GNP's electronic properties facilitate their

use as sensitizers in radiotherapy with 3-6 folds improved potential when compared to the substances usually used in clinical trials (e.g., gadolinium complexes) [3]. GNP's optical properties facilitate them to induce hyperthermia at the tumor site upon irradiation by NIR light and show enhanced potential in photothermal therapy [4].

GNPs turned out to be excellent drug nanocarriers for anti-cancer drugs as they can effectively carry a high therapeutic payload and selectively release the chemotherapeutic drug at the tumor microenvironment [5]. They are selectively delivered at the tumor micro environment by active or passive targeting methods. The former is accomplished by conjugation with ligands having an affinity for the receptors expressed on malignant cells or its microenvironment. Various ligands used for this purpose include antibodies, oligosaccharides, organic molecules, and peptides. GNPs adopt dual way approach to killing cancer cells by delivering the anti-cancer drugs effectively at the tumor site and exert photoablation therapy. The latter is accomplished by EPR (enhanced permeation and retention) effect in which GNPs reaches the interstitial space of the tumor through the leaky vasculatures and the impaired lymphatic drainage constrains the clearance of the GNPs [6]. GNPs tend to accumulate and interact with the cancer cells due to their distinctive enhanced permeability and retention property [7].

GNPs have a variety of applicability over conventional contrast agents as gold has a higher atomic number and high absorption coefficient because of its electron density, so it increases computed tomography contrast further and is used as molecular probes in

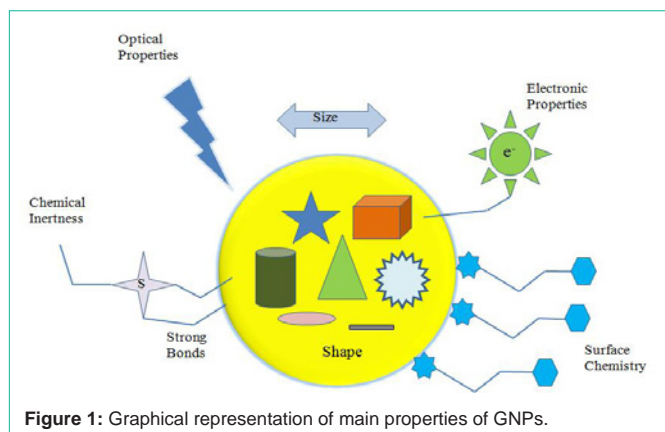


Figure 1: Graphical representation of main properties of GNPs.

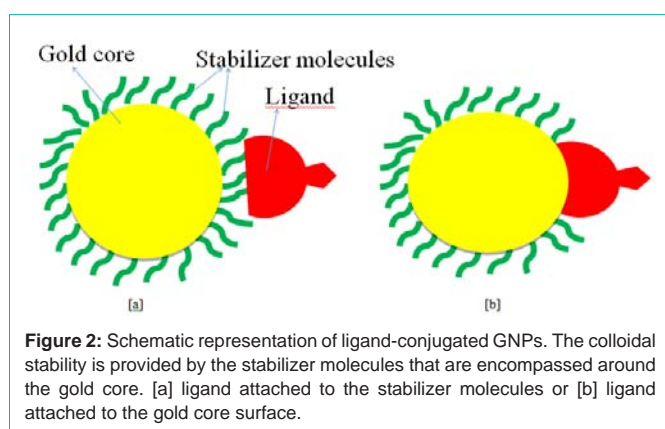


Figure 2: Schematic representation of ligand-conjugated GNPs. The colloidal stability is provided by the stabilizer molecules that are encompassed around the gold core. [a] ligand attached to the stabilizer molecules or [b] ligand attached to the gold core surface.

X-ray computed tomography imaging [8]. GNPs surfaces are readily accessible for adaptation with specific biomarkers or targeting molecules and are utilized in biomedical prospects [9,10]. They are widely used as contrast agents in molecular imaging such as MRI, CT, PET, ultrasound and optical imaging [11,12]. GNPs can get tunable absorption due to their unique anisotropic geometry and can be potentially used in the fields of photothermal therapy and biosensing [13]. Due to its Surface Plasmon Resonance (SPR), GNPs are widely used in tumor imaging, photoablation therapy, drug delivery and immune chromatographic detection of pathogens [14]. GNPs have a wide variety of applications in in-vivo imaging due to its SPR and scattering properties [15]. Surface modification of GNPs with a thermolabile polymer is utilized as a drug carrier which upon interaction with near infrared radiation releases their effectors. Because of their distinctive electronic and optical properties, GNPs have been extensively used in color indicating probes for the sensing of various analytes [16].

The purpose of the present review is to layout various applied properties of GNPs, their synthesis methods and to outline the applications and their utilization in distinctive areas of biomedicine. The theranostic applications of GNPs are catalogued into labelling and visualizing, targeted drug delivery, photoablation therapy, and sensing.

Synthesis Strategies of GnpS

GNPs of different nanoshapes are reported based on the synthesis method employed and the experimental conditions

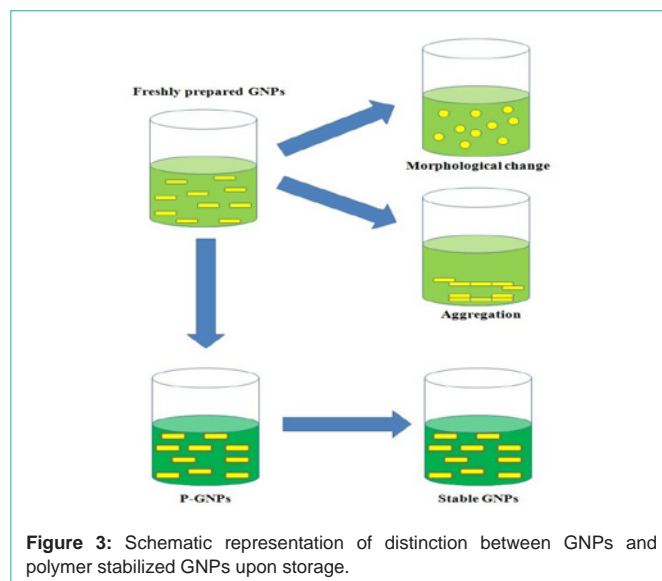


Figure 3: Schematic representation of distinction between GNPs and polymer stabilized GNPs upon storage.

involved, including gold nanospheres, nanoshells, nanocages, nanorods, nanoclusters, nanoboxes, nanocrystals, nanocubes, and nanostars [17,18]. However, when compared to other nanoshapes gold nanospheres, nanoshells, nanocages, and nanorods are widely investigated for various theranostic applications. Various synthesis methods, including chemical, physical and green methods have so far been introduced.

Chemical Methods

In chemical method, the synthesis of GNPs involves the reduction by using reducing agents like citrate acid, borohydrides, oxalic acid, formaldehyde, hydrogen peroxide, acetylene, and stabilization by using stabilizing agents like oxygen, phosphorus and nitrogen based ligands, polymers and surfactants. Turkevich method is the well-known, and widely used method for the synthesis of GNPs in which tetrachloroauric acid in water is boiled and the reducing agent trisodium citrate dehydrate is rapidly added into it with vigorous stirring. The color of the resulting solution changes from light yellow to wine red after a few minutes. In this method, there is no need of adding a stabilizing agent as citrate ions play both the role of reducing and stabilizing agents [19]. Brust-Schiffrin method is an easy and simple method for the stable GNPs synthesis with controlled size and good colloidal dispersity. In this method, tetraoctyl ammonium bromide is used as a phase transfer agent which transfers the aqueous AuCl_4 solution to a toluene phase. The resultant is reduced by using sodium borohydride in the presence of dodecanethiol. The thiol stabilized GNPs results from a color change of orange to deep brown [20]. Schematic representation of ligand conjugated GNPs is shown in (Figure 2). Other nanoshapes can be synthesized besides growing of GNPs of spherical shape.

In general, non-spherical GNPs have the tendency to aggregate. High curvature of GNPs and the nature of ligands on their surface are the main factors that influence the flocculation. Smaller GNPs due to a less number of ligands capping while larger GNPs due to interparticular interaction *via* weak hydrogen bonds shows a low affinity towards flocculation [21,22]. Flocculation is the main limitation that poses a challenge for the synthesis of a stable GNPs

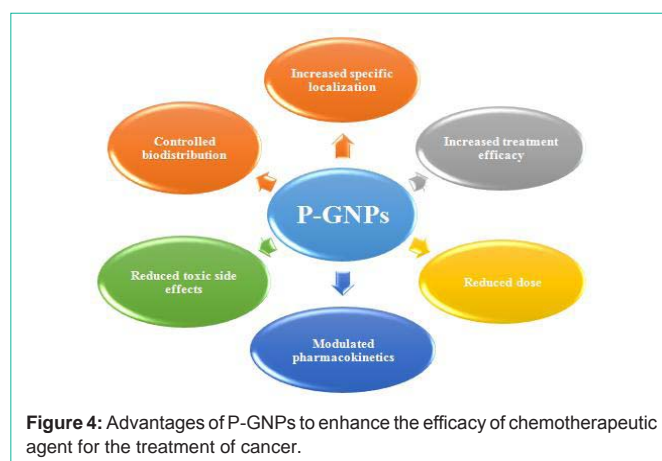
Table 1: Polymers used in the synthesis of P-GNPs.

Polymer	Nature	Reference
Cellulose derivatives	β linked D-glucose units	[24]
Chitosan	Linear polysaccharide, hydrophilic, cationic	[25]
Dextran	Branched polysaccharide, hydrophilic, cationic	[26]
Gelatin	From porcine skin, thermostable	[27]
Guar gum	Galactomannan, cationic	[28]
Heparin	Sulfated glycosaminoglycan, UV sensitive, anionic	[29]
Hyaluronic acid	Nonsulfated glycosaminoglycan, water soluble	[30]
Maltose	Disaccharide	[31]
Polycaprolactone	Film formation by thermal pressing	[32]
Polydiallyl dimethyl ammonium	Polyelectrolyte, Layer by layer assembly forming, Cationic	[33]
Polyethylene glycol	Biocompatible, non-toxic	[34]
Polyethylene imine	Cationic	[35]
Polystyrene sulfonate	Ion exchange resin, anionic	[36]
Polyvinyl caprolactam	Thermoresponsive	[37]
Polyvinyl pyrrolidone	pH- responsive	[38]
Pullulan	Amphiphilic, maltotriose units, hydrophilic, cationic	[39]
Xanthan gum	Repeated pentasaccharide, hydrophilic, anionic	[40]

colloidal solution. So, to avoid this limitation and to synthesize a stable GNPs colloidal solution, polymers are employed in the formulation. With the advancement in nanotechnology, Polymeric Gold Nanoparticles (P-GNPs) are extensively employed as drug nanocarriers for the targeted drug delivery. Direct, grafting to and grafting from methods are mainly followed in the synthesis of P-GNPs [23]. Polymers generally used in the synthesis of P-GNPs are cellulose derivatives, chitosan, dextran, gelatin, guar gum, heparin, hyaluronic acid, maltose, polycaprolactone, polydiallyl dimethyl ammonium, polyethylene glycol, polyethylene imine, polystyrene sulfonate, polyvinyl caprolactam, polyvinyl pyrrolidone, pullulan, and xanthan gum are illustrated in (Table 1) [24-40]. Graphical illustration of the distinction between GNPs and polymer stabilized GNPs are shown in (Figure 3).

These polymers show a variety of advantages in cancer therapy viz. improving biocompatibility, imparting non-immunogenicity, improving the stability of GNPs, tuning of surface density, tuning of solubility and increasing the hydrophilicity of the outer surface [41]. In the direct synthesis method, tetrachloroauric acid is subjected to reduction in the presence of sulphur-terminated polymers to form P-GNPs in a single step [42]. In grafting to method, functionalized polymers are attached to the surface of GNPs by polymers containing thiol or amine group. In grafting from method, GNP's surface is subjected to polymerization in the presence of a chain transfer agent like thiocarbonyl compounds [43,44].

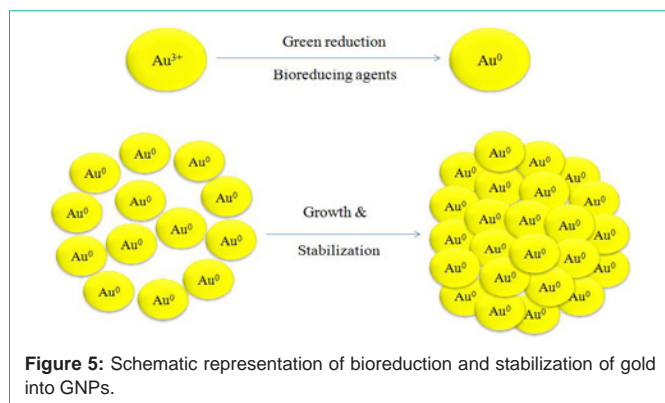
Synthesis of controllable size P-GNPs is the prerequisite for its potential theranostic applications in the treatment of cancer, which could be possible by controlling the proportions of the polymers used in the formulation. Some polymers have both reducing and stabilizing properties, e.g., xanthan gum, polystyrene and polyethylene imine, therefore, using these polymers as capping agents possibly will circumvent the use of reducing agents [45]. The advantages of

**Figure 4:** Advantages of P-GNPs to enhance the efficacy of chemotherapeutic agent for the treatment of cancer.

P-GNPs to enhance the efficacy of the chemotherapeutic agent for the treatment of cancer are illustrated in (Figure 4).

Physical Methods

Gamma irradiation method is one of the best method used for the GNPs synthesis with high purity and controlled size, where alginate solution which is a natural polysaccharide is used as a stabilizing agent [46]. Bovine serum albumin protein is employed as a stabilizing agent for the synthesis of very small GNPs in gamma irradiation method [47]. By adopting heating or photochemical reduction method, GNPs are synthesized in which malate, citrate and tartrate ligands are used to reduce tetrachloroauric acid [48]. In the photochemical method, redox and polymerization reactions are used to synthesize gold-polyethylene glycol nanoparticles. In this method, glycine and tetrachloroauric acid solution were exposed to UV radiation wherein amino acid functionalized with glycine is used as photochemical initiator [49]. In microwave method, *Cissus quadrangularis* aqueous



extract is used as a reducing agent [50]. In seed approach method, synthesis of stable GNPs is done by Co-60 irradiation using chitosan as a reducing agent. Synthesis of porous GNPs is done by using alloys of gold and silver where firstly micro emulsion of nanoparticles are prepared by using tetrachloroauric acid and silver nitrate and then reduced with sodium borohydride followed by de-alloying with nitric acid [51].

Green Methods

Chemical and physical synthesis strategies were proved to be harmful and may be injurious to humans as well as the environment due to the use of toxic chemical and elevated temperature in the synthesis process [52]. The green synthesis of GNPs is getting

Table 2: Green synthesis of GNPs using different bioreducing agents.

Bioreducing agent	Size	Shape	Reference
Azadirachta indica	11 nm	Spherical	[56]
Bacillus cereus	10-30 nm	Spherical	[57]
Butea monosperma	70-75 nm	Spherical	[58]
Micrococcus luteus	6-50 nm	Spherical	[59]
Streptomyces sp.	40 nm	Spherical	[60]
Brevibacterium casei	10-50 nm	Spherical	[61]
Mentha piperita	90 nm	Spherical	[62]
Bacillus stearothermophilus	5-30 nm	Triangular, spherical	[63]
Trichoderma viride	20-30 nm	Spherical	[64]
Zingiber officinale	5-15 nm	Spherical	[65]
Volvariella volvacea	20-150 nm	Triangular, spherical, hexagonal	[67]
Natural honey	15 nm	Spherical	[68]
Anacardium occidentale	36 nm	Hexagonal	[69]
Olive	50-100 nm	Triangular	[70]
Hibiscus cannabinus	13 nm	Spherical	[71]
Mango peel	6-18 nm	Spherical	[72]
Coriander	6-57 nm	Spherical	[73]
Nerium oleander	2-10 nm	Spherical	[74]
Solanum nigrum	50 nm	Spherical	[75]
Botrytis cinerea	1-100 nm	Triangular, spherical, hexagonal, decahedral and pyramidal	[76]

prevailing due to its non-toxicity and natural reduction at room temperature. This method emerges to be safe for clinical research as it is biocompatible, rapid, simple, suitable for large scale production, cost-efficient and eco-friendly [53]. There is a growing need to develop eco-friendly technologies in the field of nanotechnology to synthesize GNPs using micro-organisms and plants. A schematic illustration of the mechanism of bioreduction and stabilization of GNPs is shown in (Figure 5).

The biocompatibility is vital for GNPs for their theranostic applications. In GNPs green synthesis the addition of external stabilizing agents is not necessary as the biogenic components of micro-organisms and plants act themselves as stabilizing as well as capping agents.

An easy biosynthesis method for the preparation of GNPs is by using eggshell membrane. In this method eggshell membrane is immersed in tetrachloroauric acid solution, to form stable GNPs. In another method, chitosan is used as a natural reducing and stabilizing agent to synthesize GNPs in aqueous sodium chloride solution [54]. GNPs are also synthesized by using sunlight irradiation method in the reduction of gold salt is done by using solar energy. By using citrus fruits juice extracts which are obtained from Citrus reticulata, Citrus sinensis and Citrus limon [55] tetrachloroauric acid is reduced to produce GNPs. Green synthesis of GNPs using different bioreducing agents are illustrated in (Table 2) [56-75].

Applications of Gnp in Cancer Theranostics

Labelling and visualizing

GNPs are mainly used for labelling applications, where they provide contrast for the examination and visualization by directing and enriching them at the region of interest. GNPs can be visualized with a wide variety of techniques, which makes them exceptionally appealing contrast agent. GNPs substantially tend to absorb and scatter visible light. A surface plasmon is observed in which the free electrons present in the GNPs get excited by the light energy and upon light absorption, the electrons show collective oscillation in the GNPs [76]. Transfer of energy to the gold lattice results in relaxation of the excited electrons and the light absorption leads to heating of GNPs.

The visualization of the particles in numerous ways is possible mainly due to the interaction of GNPs with the light. GNPs of diameter more than 25nm are directly imaged by using Differential Interference Contrast (DIC) or optical microscopy in phase contrast mode [77]. GNPs can be used for labelling with different colors, which depends on their particle's size and shape [78]. In the case of small GNPs, the cross section for absorption decreases slowly whereas for scattering it decreases rapidly. Absorption of the light results in heating of the GNPs which subsequently leads to heating of the GNP's environment. This is observed by photothermal imaging and photoacoustic imaging. The density fluctuations of the GNPs are recorded by DIC microscopy in the former and expansion of the liquid environment due to the absorption of heat is recorded in the latter [79].

Due to light absorption, a local heat pulse is generated, resulting in the expansion of the GNP's environment which ultimately leads to the generation of a sound wave that can be detected with the help of a microphone. Small GNPs tends to emit fluorescence

Table 3: Summary of GNPs imaging applications.

Imaging	Property Required	Shape suitable	Reference
X-ray	X-ray absorption, no morphology/shape relationship	Spherical	[86-88]
Optical	Strong light scattering	Spherical	[89,90]
Fluorescence	Strong inherent fluorescence	Spherical, rod, shell	[91-92]
Photoacoustic	Strong absorption in the NIR window	Spherical, rod, cage	[93-94]
SERS	Strong electromagnetic field	Spherical, star	[95-96]

NIR- Near-infrared; SERS-Surface-enhanced raman spectroscopy.

upon photoexcitation which can be visualized by using fluorescence microscopy [80]. Apart from the visible light interaction, the electron waves and X-rays interaction is used as a valuable tool for the visualization of GNPs. Due to high atomic weight of GNPs, they give high contrast in Transmission Electron Microscopy (TEM). GNPs are used as X-ray contrast agents as they can efficiently scatter X-rays. With the help of neutron activation, GNPs can be radioactively labelled and can be detected by gamma radiation [81].

Immunostaining

From a long back GNPs have been used in biology for immunostaining wherein by using antibodies the specific targeted components of outside and inside of the cells can be labelled. They can't be visualized without labelling as they lack contrast. GNPs conjugated with antibodies that are specific to the components of interest are added to the fixed and permeabilized cells. The GNPs provide brilliant contrast for TEM imaging and with optical microscopy [82], larger structures can also be imaged. Even without fixing and permeabilizing the cells immunostaining is possible, but only labelling of the structures on the cell's surface can be done. In this case, photoacoustic imaging provides an additional feature in contrasting besides the abovementioned imaging techniques. When GNPs approach together, the plasmon resonance frequency tends to shift to higher wavelengths. When compared to freely scattered single GNPs, small aggregates of GNPs absorb light at wavelengths above the plasmon resonance. GNPs generate a photoacoustic signal when GNPs conjugated with antibodies binds to the receptors on the cell surface, unlike the GNPs which arbitrarily distributed on the surface of the cells [83].

X-rays contrast agents

The concept used in immunostaining for visualizing the components of cells is also used for providing in-vivo contrast to the organs in animals and human beings. When GNPs conjugated with antibodies or ligands are administered into the circulation system, they will bind to the specific organ through ligand-receptor

Table 4: Shapes of various GNPs and their applications in the field of medicine.

Shape	Applications
Branched particle	Substrates for surface enhanced raman spectroscopy which could be utilized for imaging at the single molecule level [135].
Hollow particle	Photothermal cancer therapy, drug delivery, optical imaging, and catalysis [136].
Faceted particle	Catalytic activity, effective and reproducible substrates for SERS [137].
Nanocage	Contrast agent for optical coherence tomography [138] and endomicroscopy imaging [139].
Nanocube	Catalysis and field enhancement applications [140].
Nanorod	Selective biomarkers in diagnostics and selective targeting in photothermal therapeutics [141].
Triangular particle	Optical sensor application and field enhancement [142].

interaction. They eventually provide contrast and resolve the structure of the desired organ through imaging. The main problem associated with GNPs as contrast agents is that their short circulation time in the body, only a small amount of GNPs get the opportunity to bind to the specific organ while the significant amount of GNPs is eliminated through the eliminating organs of the body.

By using CT, GNPs are imaged with a significant signal to noise ratio [84] so that the exposure times can be reduced leading to a reduction in the radiation harm to the surrounding healthy tissues (Table 3) [86-96]. The organs of the body are imaged for the treatment by the penetration of X-rays through the skin. Moreover, X-ray computed tomography setups are readily available in many hospitals and diagnostic centres. GNPs causes less damage when compared to the quantum dots that are used as fluorescent semiconductor detecting and imaging agents [85].

Targeted Drug Delivery

GNPs are used as drug nanocarriers for the delivery of therapeutic agents into the tumor cells from quite a long time. Therapeutic agents are adsorbed on the GNP's surface and the entire conjugate by means gene guns or particle ingestion is introduced into the tumor cells. Therapeutic agents will detach from the GNPs inside the tumor cells eventually after its introduction. DNA is adsorbed on the GNP's surface and these nanobullets are introduced into the tumor cells for the ballistic influx of DNA [97,98]. GNPs ingestion into the tumor cells is either nonspecific or specific through receptor-ligand interaction and the main objective involved in this is to transfer and deliver the therapeutic agents adsorbed on the GNP's surface into the cells [99]. To deliver the therapeutic agents from the GNPs into the cytosol, GNP's surface is encapsulated with membrane disruptive peptides [100,101].

The delivery of therapeutic agents into tumor cells through particle ingestion is mainly useful in gene therapy and drug targeting. In gene therapy, for the expression of corresponding proteins, DNA adsorbed on the GNP's surface is introduced into the tumor cells [102]. In drug targeting, for the delivery of anticancer drug specifically to the target tumor cells or its microenvironment, GNPs are conjugated with specific ligands. To facilitate these applications, there is no need to exploit any of the unique properties of GNPs and their characteristics like inertness, stable, small and relatively easy to conjugate with ligands make them be used as a biocompatible and reliable means for targeted drug delivery [103,104].

Photoablation Therapy (PAT)

PAT is mainly grouped into Photothermal Therapy (PTT) and

Photodynamic Therapy (PDT) [105]. In order to accomplish the penetration in the blood and the tissue, NIR light is usually used. In PTT the absorbed light is converted into local heat i.e., hyperthermia which shows a significant cytotoxic effect on the tumor cells and its microenvironment [106]. In PDT the photon energy is converted to generate ROS (reactive oxygen species) by the stimulation of photosensitizers to kill tumor cells and its microenvironment. The absorption of light by the GNPs results in the excitation of the free electrons and this excitation at the SPR eventually leads to the cumulative oscillation of the energized free electrons. The GNP's crystal lattice and the electrons interaction lead to the relaxation of the electrons and the heat energy is exchanged to the lattice and subsequently scattered to the surrounding environment [107]. Apart from its various imaging methods, heating of GNPs in a controlled manner leads to manipulation of the surrounding tissues in various ways [108].

Intracellular delivery and the molecular uptake which mainly depends on phagocytosis and endocytosis can be potentiated by inducing hyperthermia at the tumor site [109]. Due to extreme sensitivity, even a small increase in the temperature by a few degrees leads to the cells death. Temperature above 38°C results in fever and above 42°C results in cell damage in humans and by making use of hyperthermia in a controlled way cancer can be treated. This is possible by coordinating the GNPs whose surface is conjugated with ligands that are specific to the receptors that are over-expressed on tumor cells. When the locally enriched GNPs in the tumor cells are heated with the help of external stimuli, the tumor cells which are in the vicinity of the GNPs can be killed selectively [110]. In this way, we can enrich malignant tissues with GNPs and illuminate the tissue. Tumor cells can be destroyed locally by GNPs mediated heat [111] without exposing the whole organism to higher temperatures.

PDT is a light sensitizer stimulation therapy which comprises of a particular wavelength of drug activating light, a photosensitizer drug and oxygen [112]. The stimulation of photosensitizer leads to energy transfer and eventually yields highly reactive oxygen species which imparts apoptosis and necrosis by inducing microvasculature damage [113]. Unlike radiotherapy, PDT does not have any harmful impact on cells and tissues that are devoid of photosensitizer drug as the light used for the stimulation is nonionizing. So it can be used safely without causing any harm to the neighbouring healthy tissue after taking a multi-dosage regimen [114,115]. By using fluorescence resonance energy transfer mechanism, the self-quenching capacity of GNPs plays a major role in image guided PDT in annihilating the tumor.

Sensing

Besides using GNPs in labelling, visualizing and targeted drug delivery, they are also used in active sensor applications. GNPs acts as sensors and explicitly register the existence of analyte molecules to give a readout which specifies the amount of analyte. GNP based sensors have a significant impact in diagnostics because of its small size.

Quenching

When the fluorophores are in close vicinity to the gold surface, their fluorescence can be quenched and this can be used for competitive

displacement sensor strategy [116]. For quantitative identification of an individual analyte molecule, ligands are conjugated with the GNPs that distinctively bind to the analyte molecule. By saturating with the analyte molecules, the binding sites of the ligands get jammed and then these analyte molecules are customized with fluorophores. Their fluorescence can be quenched as they are firmly connected to the GNPs. The free analyte molecules dislocate the analyte molecules that are bound to the ligands in a constant dynamic equilibrium. When the concentration of the free analyte molecules is more, the fluorophore labelled molecules discharges from the GNP's surface and there will be no quenching. This strategy can be changed by using GNPs as quenchers for quantum dots which are substituted by analyte molecules [117].

Surface Plasmon Resonance

GNPs are used in sensing mainly because of its most reliable intrinsic feature, plasmon resonance frequency [118] that can be altered directly by coupling the molecules to the GNP's surface [119]. When the GNPs are closely arranged, the plasmon resonance frequency drastically changes and they form minute aggregates which can be used for colorimetric identification of the analyte molecules. This technique was developed by Mirkin and his associates and is considered as one of the most distinguished illustrations of the gold based sensor [120,121]. To find the DNA, this innovative assay was developed. Oligonucleotides correlated to the target sequence that was supposed to be identified was conjugated with GNPs. When the target sequence does not exist then the GNPs freely scatters and the colloidal solution seems red. When it exists, the GNPs bind to it by hybridization of DNA complimentary strands. The hybridization leads to the development of GNPs aggregates, resulting in the adjustment in the plasmon resonance and the solution shows up a blue/ violet color. At the point when the sample is subjected to heat, mismatch in even a single sequence result in an altered melting temperature which causes a change in the color. The quantitative identification of DNA sequences is feasible with this idea, even at a very low concentration [122].

Apart from DNA assays, the same idea is also used for the quantitative identification of the metals or proteins [123]. The enzymatic action can also be checked with the same idea [124]. Other than the analytes detection, such color changes can furthermore be utilized to measure the lengths. Discrete areas of analytes can be linked to GNPs and the conformation changes in the analytes can be examined [125].

Surface-Enhanced Raman Scattering

With the help of raman scattering, many analyte molecules are distinguished, owing to their characteristic spectra [126]. In raman scattering, the scattering is inelastic and the scattered light show lower or higher energy than the incident light. When the scattered light has lower energy, the energy is stored in the analyte molecules and when it has higher energy, the energy is attained from the analyte molecules. This shift in the energy is characteristic for the entities in the analyte molecules and they possess a characteristic raman spectrum which is used for their identification and detection. The scattering efficiency mainly depends on the incident light's wavelength. Characteristically raman signals are relatively weak and for that reason, an adequate

concentration of the analyte molecules is required to provide signals that are relatively enough. In surface enhanced raman spectroscopy, the raman scattering can be significantly enhanced by guiding the analyte molecules close to the surface of GNP's having a high curvature [127,128]. Because of the GNP's plasmon resonance, the electric field near the GNPs will be stronger than the field strength of the incident light.

Binding of the ligands to the GNP's surface significantly enhances the raman signal of the analyte molecules and helps in its identification [129,130]. Recent advancements comprise of GNPs customized with raman active analyte molecules for the identification of proteins [131,132] or DNA [133] and two photon excitation [134]. Various shapes of GNPs and their respective applications in the field of medicine are illustrated in (Table 4) [135-142].

Conclusion

GNPs have brought a change in the field of biomedicine by virtue of its applications in labelling and visualization, targeted drug delivery, photoablation therapy and sensing due to their exceptional stability, inert nature, biocompatibility, non-cytotoxicity, very small size, large surface area, and tunable physical, chemical, optical and electronic properties. The remarkable properties of the GNPs make them as a most valuable means in numerous applications. Almost all the comparative applications can be performed by using diverse material nanoparticles like quantum dots, the numerous properties and features possessed by GNPs make them unique. GNPs are inert and biocompatible and till now there is no evidence of gold corrosion. The synthesis of GNPs and its conjugation with the biomolecules can be done in a simple way. The unique optical properties of GNPs make them be visualized with distinctive strategies specifically because of its SPR and they are used as sensors in view of any changes in SPR.

GNPs enhance the quality life of patients due to the fact that the side effects of conventional drugs are reduced by conjugation with GNPs. By using GNPs, targeted drug delivery of therapeutic drugs can be achieved as the drug gets released at a specific site and can interact with the tumorous cell. GNPs can be used in various *in vitro* assays and standard kits and there is a lot of scope for them in new research to widespread its usage by using its unique properties. They can also be used in a number of commercial sensor assays for the identification of analyte molecules.

The use of GNPs in the cancer treatment for the development of stable systems with consistent reproducibility is a future challenge GNPs are facing today. To accomplish an effective cancer response, the development of GNPs of 10nm size for their delivery to the cytosol is a prerequisite to be fulfilled by GNPs.

References

- van der Veldt AA, Hendrikse NH, Smit EF, Mooijer MP, Rijnders AY, Gerritsen WR, et al. Biodistribution and radiation dosimetry of ¹¹C-labelled docetaxel in cancer patients. *European journal of nuclear medicine and molecular imaging*. 2010; 37: 1950-1958.
- Bayazitoglu Y, Kheradmand S, Tullius TK. An overview of nanoparticle assisted laser therapy. *International Journal of Heat and Mass Transfer*. 2013; 67: 469-486.
- Hainfeld JF, Smilowitz HM, O'Connor MJ, Dilmanian FA, Slatkin DN. Gold nanoparticle imaging and radiotherapy of brain tumors in mice. *Nanomedicine*. 2013; 8: 1601-1609.
- Kodiha M, Hutter E, Boridy S, Juhas M, Maysinger D, Stochaj U. Gold nanoparticles induce nuclear damage in breast cancer cells, which is further amplified by hyperthermia. *Cell Mol Life Sci*. 2014; 71: 4259-4273.
- Kim CS, Tonga GY, Solfiell D, Rotello VM. Inorganic nanosystems for therapeutic delivery: Status and prospects. *Advanced Drug Delivery Reviews*. 2013; 65: 93-99.
- Nichols JW, Bae YH. EPR: Evidence and fallacy. *Journal of Controlled Release*. 2014; 190: 451-464.
- Choi Y, Lee K, Gupta KC, Park S-Y, Kang I-K. The role of ligand-receptor interactions in visual detection of HepG2 cells using a liquid crystal microdroplet-based biosensor. *Journal of Materials Chemistry B*. 2015; 3: 8659-8669.
- Amjadi M, Farzampour L. Fluorescence quenching of fluoroquinolones by gold nanoparticles with different sizes and its analytical application. *Journal of Luminescence*. 2014; 145: 263-268.
- Guo Q, Guo Q, Yuan J, Zeng J. Biosynthesis of gold nanoparticles using a kind of flavonol: Dihydromyricetin. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2014; 441: 127-132.
- Lan M-Y, Hsu Y-B, Hsu C-H, Ho C-Y, Lin J-C, Lee S-W. Induction of apoptosis by high-dose gold nanoparticles in nasopharyngeal carcinoma cells. *Auris Nasus Larynx*. 2013; 40: 563-568.
- Qie F, Astolfo A, Wickramaratna M, Behe M, Evans MD, Hughes TC, et al. Self-assembled gold coating enhances X-ray imaging of alginate microcapsules. *Nanoscale*. 2015; 7: 2480-2488.
- Tiedemann D, Taylor U, Rehbock C, Jakobi J, Klein S, Kues WA, et al. Reprotoxicity of gold, silver, and gold-silver alloy nanoparticles on mammalian gametes. *Analyst*. 2014; 139: 931-942.
- Pal R, Panigrahi S, Bhattacharyya D, Chakraborti AS. Characterization of citrate capped gold nanoparticle-quercetin complex: Experimental and quantum chemical approach. *Journal of Molecular Structure*. 2013; 1046: 153-163.
- Yang C, Neshatian M, van Prooijen M, Chithrani DB. Cancer nanotechnology: enhanced therapeutic response using peptide-modified gold nanoparticles. *Journal of nanoscience and nanotechnology*. 2014; 14: 4813-4819.
- Rossi M, Della Pina C, Falletta E. Gold nanomaterials: From preparation to pharmaceutical design and application. *Current pharmaceutical design*. 2016; 22: 1485-1493.
- Murawala P, Tirmale A, Shiras A, Prasad B. In situ synthesized BSA capped gold nanoparticles: effective carrier of anticancer drug methotrexate to MCF-7 breast cancer cells. *Materials Science and Engineering: C*. 2014; 34: 158-167.
- Khan MS, Vishakante GD, Siddaramaiah H. Gold nanoparticles: A paradigm shift in biomedical applications. *Advances in Colloid and Interface Science*. 2013; 199-200: 44-58.
- Minati L, Benetti F, Chiappini A, Speranza G. One-step synthesis of star-shaped gold nanoparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2014; 441: 623-628.
- Turkevich J, Stevenson PC, Hillier J. A study of the nucleation and growth processes in the synthesis of colloidal gold. *Discussions of the Faraday Society*. 1951; 11: 55-75.
- Brust M, Walker M, Bethell D, Schiffrin DJ, Whyman R. Synthesis of thiol-derivatised gold nanoparticles in a two-phase liquid-liquid system. *Journal of the Chemical Society, Chemical Communications*. 1994: 801-802.
- Dreaden EC, Austin LA, Mackey MA, El-Sayed MA. Size matters: gold nanoparticles in targeted cancer drug delivery. *Therapeutic delivery*. 2012; 3: 457-478.
- Afroz ARMN, Sivalapalan ST, Murphy CJ, Hussain SM, Schlager JJ, Saleh NB. Spheres vs. rods: The shape of gold nanoparticles influences aggregation and deposition behavior. *Chemosphere*. 2013; 91: 93-98.
- Li D, He Q, Li J. Smart core/shell nanocomposites: Intelligent polymers modified gold nanoparticles. *Advances in Colloid and Interface Science*.

- 2009; 149: 28-38.
24. Wu X, Lu C, Zhou Z, Yuan G, Xiong R, Zhang X. Green synthesis and formation mechanism of cellulose nanocrystal-supported gold nanoparticles with enhanced catalytic performance. *Environmental Science: Nano*. 2014; 1: 71-79.
 25. Wu Y, Zuo F, Lin Y, Zhou Y, Zheng Z, Ding X. Green and facile synthesis of gold nanoparticles stabilized by chitosan. *Journal of Macromolecular Science, Part A*. 2014; 51: 441-446.
 26. Correard F, Maximova K, Estève M-A, Villard C, Roy M, Al-Kattan A, et al. Gold nanoparticles prepared by laser ablation in aqueous biocompatible solutions: assessment of safety and biological identity for nanomedicine applications. *International journal of nanomedicine*. 2014; 9: 5415.
 27. Canpean V, Gabudean A, Astilean S. Enhanced thermal stability of gelatin coated gold nanorods in water solution. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2013; 433: 9-13.
 28. Pandey S, Goswami GK, Nanda KK. Green synthesis of polysaccharide/gold nanoparticle nanocomposite: An efficient ammonia sensor. *Carbohydrate polymers*. 2013; 94: 229-234.
 29. Rodríguez-Torres MdP, Díaz-Torres LA, Salas P, Rodríguez-González C, Olmos-López M. UV photochemical synthesis of heparin-coated gold nanoparticles. *Gold Bulletin*. 2014; 47: 21-31.
 30. Cheng D, Han W, Yang K, Song Y, Jiang M, Song E. One-step facile synthesis of hyaluronic acid functionalized fluorescent gold nanoprobe sensitive to hyaluronidase in urine specimen from bladder cancer patients. *Talanta*. 2014; 130: 408-414.
 31. Fent GM, Casteel SW, Kim DY, Kannan R, Katti K, Chanda N, et al. Biodistribution of maltose and gum arabic hybrid gold nanoparticles after intravenous injection in juvenile swine. *Nanomedicine: Nanotechnology, Biology and Medicine*. 2009; 5: 128-135.
 32. Mao Z, Wang B, Ma L, Gao C, Shen J. The influence of polycaprolactone coating on the internalization and cytotoxicity of gold nanoparticles. *Nanomedicine: Nanotechnology, Biology and Medicine*. 2007; 3: 215-223.
 33. Zhao E, Zhao Z, Wang J, Yang C, Chen C, Gao L, et al. Surface engineering of gold nanoparticles for in vitro siRNA delivery. *Nanoscale*. 2012; 4: 5102-5109.
 34. Asadishad B, Vossoughi M, Alemzadeh I. Folate-receptor-targeted delivery of doxorubicin using polyethylene glycol-functionalized gold nanoparticles. *Industrial & engineering chemistry research*. 2010; 49: 1958-1963.
 35. Kim J, Park J, Kim H, Singha K, Kim WJ. Transfection and intracellular trafficking properties of carbon dot-gold nanoparticle molecular assembly conjugated with PEI-pDNA. *Biomaterials*. 2013; 34: 7168-7180.
 36. Jahan S, Mansoor F, Kanwal S. Polymers effects on synthesis of AuNPs, and Au/Ag nanoalloys: indirectly generated AuNPs and versatile sensing applications including anti-leukemic agent. *Biosensors and Bioelectronics*. 2014; 53: 51-57.
 37. Beija M, Marty J-D, Destarac M. Thermoresponsive poly (N-vinyl caprolactam)-coated gold nanoparticles: sharp reversible response and easy tunability. *Chemical Communications*. 2011; 47: 2826-2828.
 38. Dhumale VA, Gangwar RK, Datar SS, Sharma RB. Reversible aggregation control of polyvinylpyrrolidone capped gold nanoparticles as a function of pH. *Materials Express*. 2012; 2: 311-318.
 39. Ganeshkumar M, Ponrasu T, Raja MD, Subamekala MK, Suguna L. Green synthesis of pullulan stabilized gold nanoparticles for cancer targeted drug delivery. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2014; 130: 64-71.
 40. Pooja D, Panyaram S, Kulhari H, Rachamalla SS, Sistla R. Xanthan gum stabilized gold nanoparticles: characterization, biocompatibility, stability and cytotoxicity. *Carbohydrate polymers*. 2014; 110: 1-9.
 41. Danila D, Johnson E, Kee P. CT imaging of myocardial scars with collagen-targeting gold nanoparticles. *Nanomedicine: Nanotechnology, Biology and Medicine*. 2013; 9: 1067-1076.
 42. Scaravelli RCB, Dazzi RL, Giacomelli FC, Machado G, Giacomelli C, Schmidt V. Direct synthesis of coated gold nanoparticles mediated by polymers with amino groups. *Journal of Colloid and Interface Science*. 2013; 397: 114-121.
 43. Takara M, Toyoshima M, Seto H, Hoshino Y, Miura Y. Polymer-modified gold nanoparticles via RAFT polymerization: a detailed study for a biosensing application. *Polymer Chemistry*. 2014; 5: 931-939.
 44. Navarro JR, Lerouge F, Cepraga C, Micouin G, Favier A, Chateau D, et al. Nanocarriers with ultrahigh chromophore loading for fluorescence bio-imaging and photodynamic therapy. *Biomaterials*. 2013; 34: 8344-8351.
 45. Kirtane AR, Kalscheuer SM, Panyam J. Exploiting nanotechnology to overcome tumor drug resistance: Challenges and opportunities. *Advanced Drug Delivery Reviews*. 2013; 65: 1731-1747.
 46. Guo Q, Zou S, Li J, Li D, Jiao H, Shi J. Facile synthesis of morphology- and size-controllable polythiophene/gold composites in aqueous medium. *Journal of Nanoparticle Research*. 2014; 16: 1-11.
 47. Deb S, Patra HK, Lahiri P, Dasgupta AK, Chakrabarti K, Chaudhuri U. Multistability in platelets and their response to gold nanoparticles. *Nanomedicine: Nanotechnology, Biology and Medicine*. 2011; 7: 376-384.
 48. Guo W, Pi Y, Song H, Tang W, Sun J. Layer-by-layer assembled gold nanoparticles modified anode and its application in microbial fuel cells. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2012; 415: 105-111.
 49. Ganeshkumar M, Sastry TP, Kumar MS, Dinesh MG, Kannappan S, Suguna L. Sun light mediated synthesis of gold nanoparticles as carrier for 6-mercaptopurine: Preparation, characterization and toxicity studies in zebrafish embryo model. *Materials Research Bulletin*. 2012; 47: 2113-2119.
 50. Pérez Y, Mann E, Herradón B. Preparation and characterization of gold nanoparticles capped by peptide-biphenyl hybrids. *Journal of colloid and interface science*. 2011; 359: 443-453.
 51. Kojima C, Kawabata H, Harada A, Horinaka H, Kono K. Design of a Novel Drug Carrier with Photoresponsive Properties: Drug-encapsulated and Alkanethiol-modified Gold-nanoparticle-loaded PEGylated Dendrimer. *Chemistry Letters*. 2013; 42: 612-614.
 52. Birla S, Tiwari V, Gade A, Ingle A, Yadav A, Rai M. Fabrication of silver nanoparticles by Phoma glomerata and its combined effect against *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Letters in Applied Microbiology*. 2009; 48: 173-179.
 53. Rai M, Yadav A, Gade A. CRC 675-current trends in phytosynthesis of metal nanoparticles. *Critical reviews in biotechnology*. 2008; 28: 277-284.
 54. Krishnaswamy K, Orsat V. Insight into the nanodielectric properties of gold nanoparticles synthesized from maple leaf and pine needle extracts. *Industrial Crops and Products*. 2015; 66: 131-136.
 55. Tarnawski R, Ulbricht M. Amphiphilic gold nanoparticles: Synthesis, characterization and adsorption to PEGylated polymer surfaces. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2011; 374: 13-21.
 56. Nazeruddin G, Prasad N, Waghmare S, Garadkar K, Mulla I. Extracellular biosynthesis of silver nanoparticle using *Azadirachta indica* leaf extract and its anti-microbial activity. *Journal of Alloys and Compounds*. 2014; 583: 272-277.
 57. Gupta VK, Atar N, Yola ML, Darcan C, İdil Ö, Üstündağ Z. Biosynthesis of silver nanoparticles using chitosan immobilized *Bacillus cereus*: nanocatalytic studies. *Journal of Molecular Liquids*. 2013; 188: 81-88.
 58. Patra S, Mukherjee S, Barui AK, Ganguly A, Sreedhar B, Patra CR. Green synthesis, characterization of gold and silver nanoparticles and their potential application for cancer therapeutics. *Materials Science and Engineering: C*. 2015; 53: 298-309.
 59. Arunkumar P, Thanalakshmi M, Kumar P, Premkumar K. *Micrococcus luteus* mediated dual mode synthesis of gold nanoparticles: Involvement of extracellular α -amylase and cell wall teichuronic acid. *Colloids and Surfaces B: Biointerfaces*. 2013; 103: 517-22.

60. Manivasagan P, Venkatesan J, Kang K-H, Sivakumar K, Park S-J, Kim S-K. Production of α -amylase for the biosynthesis of gold nanoparticles using *Streptomyces* sp. MBRC-82. *International journal of biological macromolecules*. 2015; 72: 71-78.
61. Kalishwaralal K, Deepak V, Pandian SRK, Kottaisamy M, BarathManiKanth S, Kartikeyan B, et al. Biosynthesis of silver and gold nanoparticles using *Brevibacterium casei*. *Colloids and Surfaces B: Biointerfaces*. 2010; 77: 257-262.
62. MubarakAli D, Thajuddin N, Jeganathan K, Gunasekaran M. Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. *Colloids and Surfaces B: Biointerfaces*. 2011; 46: 360-365.
63. Luo P, Liu Y, Xia Y, Xu H, Xie G. Aptamer biosensor for sensitive detection of toxin A of *Clostridium difficile* using gold nanoparticles synthesized by *Bacillus stearothermophilus*. *Biosensors and Bioelectronics*. 2014; 54: 217-221.
64. Mishra A, Kumari M, Pandey S, Chaudhry V, Gupta K, Nautiyal C. Biocatalytic and antimicrobial activities of gold nanoparticles synthesized by *Trichoderma* sp. *Bioresource technology*. 2014; 166: 235-242.
65. Kumar KP, Paul W, Sharma CP. Green synthesis of gold nanoparticles with *Zingiber officinale* extract: characterization and blood compatibility. *Process Biochemistry*. 2011; 46: 2007-2013.
66. Philip D. Biosynthesis of Au, Ag and Au-Ag nanoparticles using edible mushroom extract. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2009; 73: 374-381.
67. Philip D. Honey mediated green synthesis of gold nanoparticles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2009; 73: 650-653.
68. Sheny D, Mathew J, Philip D. Synthesis characterization and catalytic action of hexagonal gold nanoparticles using essential oils extracted from *Anacardium occidentale*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2012; 97: 306-310.
69. Khalil MM, Ismail EH, El-Magdoub F. Biosynthesis of Au nanoparticles using olive leaf extract: 1st nano updates. *Arabian Journal of Chemistry*. 2012; 5: 431-437.
70. Bindhu M, Rekha PV, Umamaheswari T, Umadevi M. Antibacterial activities of *Hibiscus cannabinus* stem-assisted silver and gold nanoparticles. *Materials Letters*. 2014; 131: 194-197.
71. Yang N, WeiHong L, Hao L. Biosynthesis of Au nanoparticles using agricultural waste mango peel extract and its in vitro cytotoxic effect on two normal cells. *Materials Letters*. 2014; 134: 67-70.
72. Narayanan KB, Sakthivel N. Coriander leaf mediated biosynthesis of gold nanoparticles. *Materials Letters*. 2008; 62: 4588-4590.
73. Tahir K, Nazir S, Li B, Khan AU, Khan ZUH, Gong PY, et al. Nerium oleander leaves extract mediated synthesis of gold nanoparticles and its antioxidant activity. *Materials Letters*. 2015; 156: 198-201.
74. Muthuvel A, Advallan K, Balamurugan K, Krishnakumar N. Biosynthesis of gold nanoparticles using *Solanum nigrum* leaf extract and screening their free radical scavenging and antibacterial properties. *Biomedicine & Preventive Nutrition*. 2014; 4: 325-332.
75. Castro M, Cottet L, Castillo A. Biosynthesis of gold nanoparticles by extracellular molecules produced by the phytopathogenic fungus *Botrytis cinerea*. *Materials Letters*. 2014; 115: 42-44.
76. Clavero C. Plasmon-induced hot-electron generation at nanoparticle/metal-oxide interfaces for photovoltaic and photocatalytic devices. *Nature Photonics*. 2014; 8: 95-103.
77. Aioub M, Kang B, Mackey MA, El-Sayed MA. Biological Targeting of Plasmonic Nanoparticles Improves Cellular Imaging via the Enhanced Scattering in the Aggregates Formed. *The journal of physical chemistry letters*. 2014; 5: 2555-2561.
78. Tang B, Sun L, Li J, Zhang M, Wang X. Sunlight-driven synthesis of anisotropic silver nanoparticles. *Chemical engineering journal*. 2015; 260: 99-106.
79. Leduc C, Si S, Gautier J, Soto-Ribeiro M, Wehrle-Haller B, Gautreau A, et al. A highly specific gold nanoprobe for live-cell single-molecule imaging. *Nano letters*. 2013; 13: 1489-1494.
80. Yahia-Ammar A, Sierra D, Mérola F, Hildebrandt N, Le Guével X. Self-Assembled Gold Nanoclusters for Bright Fluorescence Imaging and Enhanced Drug Delivery. *ACS nano*. 2016; 10: 2591-2599.
81. Comenge J, Fragueiro O, Sharkey J, Taylor A, Held M, Burton NC, et al. Preventing Plasmon Coupling between Gold Nanorods Improves the Sensitivity of Photoacoustic Detection of Labeled Stem Cells *in Vivo*. *ACS nano*. 2016; 10: 7106-7116.
82. Marasini C, Jacchetti E, Moretti M, Canale C, Moran O, Vassalli M. Visualization of single proteins from stripped native cell membranes: A protocol for high-resolution atomic force microscopy. *Microscopy research and technique*. 2013; 76: 723-732.
83. Mallidi S, Kim S, Karpouk A, Joshi PP, Sokolov K, Emelianov S. Visualization of molecular composition and functionality of cancer cells using nanoparticle-augmented ultrasound-guided photoacoustics. *Photoacoustics*. 2015; 3: 26-34.
84. Cole LE, Ross RD, Tilley JM, Vargo-Gogola T, Roeder RK. Gold nanoparticles as contrast agents in x-ray imaging and computed tomography. *Nanomedicine*. 2015; 10: 321-341.
85. Hoshino K, Joshi PP, Bhavne G, Sokolov KV, Zhang X. Use of colloidal quantum dots as a digitally switched swept light source for gold nanoparticle based hyperspectral microscopy. *Biomedical optics express*. 2014; 5: 1610-1615.
86. Wang H, Zheng L, Guo R, Peng C, Shen M, Shi X, et al. Dendrimer-entrapped gold nanoparticles as potential CT contrast agents for blood pool imaging. *Nanoscale research letters*. 2012; 7: 190.
87. Peng C, Zheng L, Chen Q, Shen M, Guo R, Wang H, et al. PEGylated dendrimer-entrapped gold nanoparticles for in vivo blood pool and tumor imaging by computed tomography. *Biomaterials*. 2012; 33: 1107-1119.
88. Schirra CO, Senpan A, Roessler E, Thran A, Stacy AJ, Wu L, et al. Second generation gold nanobeacons for robust K-edge imaging with multi-energy CT. *Journal of materials chemistry*. 2012; 22: 23071-23077.
89. Craig GA, Allen PJ, Mason MD. Synthesis, characterization, and functionalization of gold nanoparticles for cancer imaging. *Cancer Nanotechnology: Methods and Protocols*. 2010: 177-193.
90. Qian W, Huang X, Kang B, El-Sayed MA. Dark-field light scattering imaging of living cancer cell component from birth through division using bioconjugated gold nanoprobe. *Journal of biomedical optics*. 2010; 15: 046025.
91. van Schooneveld MM, Cormode DP, Koole R, van Wijngaarden JT, Calcagno C, Skajaa T, et al. A fluorescent, paramagnetic and PEGylated gold/silica nanoparticle for MRI, CT and fluorescence imaging. *Contrast media & molecular imaging*. 2010; 5: 231-236.
92. Zheng J, Zhou C, Yu M, Liu J. Different sized luminescent gold nanoparticles. *Nanoscale*. 2012; 4: 4073-4083.
93. Nam SY, Ricles LM, Suggs LJ, Emelianov SY. In vivo ultrasound and photoacoustic monitoring of mesenchymal stem cells labeled with gold nanotracers. *PLoS One*. 2012; 7: e37267.
94. Chen Y-S, Frey W, Kim S, Kruizinga P, Homan K, Emelianov S. Silica-coated gold nanorods as photoacoustic signal nanoamplifiers. *Nano letters*. 2011; 11: 348-354.
95. Mecker LC, Tyner KM, Kauffman JF, Arzhantsev S, Mans DJ, Gryniwicz-Ruzicka CM. Selective melamine detection in multiple sample matrices with a portable Raman instrument using surface enhanced Raman spectroscopy-active gold nanoparticles. *Analytica chimica acta*. 2012; 733: 48-55.
96. Nolan JP, Duggan E, Liu E, Condello D, Dave I, Stoner SA. Single cell analysis using Surface Enhanced Raman Scattering (SERS) tags. *Methods*. 2012; 57: 272-279.

97. Ibraheem D, Elaissari A, Fessi H. Gene therapy and DNA delivery systems. *International journal of pharmaceutics*. 2014; 459: 70-83.
98. Gotesman M, Williams SA. Using a Handheld Gene Gun for Genetic Transformation of *Tetrahymena thermophila*. *Cytoskeleton Methods and Protocols: Methods and Protocols*. 2016: 373-383.
99. Bazak R, Houri M, El Achy S, Kamel S, Refaat T. Cancer active targeting by nanoparticles: a comprehensive review of literature. *Journal of cancer research and clinical oncology*. 2015; 141: 769-784.
100. Kulkarni SA, Feng S-S. Effects of particle size and surface modification on cellular uptake and biodistribution of polymeric nanoparticles for drug delivery. *Pharmaceutical research*. 2013; 30: 2512-2522.
101. Ding Y, Jiang Z, Saha K, Kim CS, Kim ST, Landis RF, et al. Gold nanoparticles for nucleic acid delivery. *Molecular Therapy*. 2014; 22: 1075.
102. Vial S, Reis RL, Oliveira JM. Recent advances using gold nanoparticles as a promising multimodal tool for tissue engineering and regenerative medicine. *Current Opinion in Solid State and Materials Science*. 2016.
103. Rana S, Bajaj A, Mout R, Rotello VM. Monolayer coated gold nanoparticles for delivery applications. *Advanced drug delivery reviews*. 2012; 64: 200-216.
104. Li S, Xu L, Ma W, Wu X, Sun M, Kuang H, et al. Dual-Mode Ultrasensitive Quantification of MicroRNA in Living Cells by Chiroplasmonic Nanopyramids Self-Assembled from Gold and Upconversion Nanoparticles. *Journal of the American Chemical Society*. 2015; 138: 306-312.
105. Kim J-W, Jung Y-W, Nam J-W. Study on the Development of Road Icing Forecast and Snow Detection System Using State Evaluation Algorithm of Multi Sensing Method. *Journal of the Korea institute for structural maintenance and inspection*. 2013; 17: 113-121.
106. Wang S, Dai Z, Ke H, Qu E, Qi X, Zhang K, et al. Contrast ultrasound-guided photothermal therapy using gold nanoshelled microcapsules in breast cancer. *European journal of radiology*. 2014; 83: 117-122.
107. Hogan NJ, Urban AS, Ayala-Orozco C, Pimpinelli A, Nordlander P, Halas NJ. Nanoparticles heat through light localization. *Nano letters*. 2014; 14: 4640-4645.
108. Jiang K, Smith DA, Pinchuk A. Size-dependent Photothermal conversion efficiencies of plasmonically heated gold nanoparticles. *The Journal of Physical Chemistry C*. 2013; 117: 27073-27080.
109. Gormley AJ, Larson N, Banisadr A, Robinson R, Frazier N, Ray A, et al. Plasmonic photothermal therapy increases the tumor mass penetration of HPMA copolymers. *Journal of Controlled Release*. 2013; 166: 130-138.
110. Choi J, Kim H-Y, Ju E-J, Jung J, Park J, Chung H-K, et al. Use of macrophages to deliver therapeutic and imaging contrast agents to tumors. *Biomaterials*. 2012; 33: 4195-4203.
111. Lee J, Chatterjee DK, Lee MH, Krishnan S. Gold nanoparticles in breast cancer treatment: promise and potential pitfalls. *Cancer letters*. 2014; 347: 46-53.
112. Gupta A, Avci P, Sadasivam M, Chandran R, Parizotto N, Vecchio D, et al. Shining light on nanotechnology to help repair and regeneration. *Biotechnology Advances*. 2013; 31: 607-631.
113. Chinen AB, Guan CM, Ferrer JR, Barnaby SN, Merkel TJ, Mirkin CA. Nanoparticle probes for the detection of cancer biomarkers, cells, and tissues by fluorescence. *Chemical reviews*. 2015; 115: 10530-10574.
114. Seo S-H, Kim B-M, Joe A, Han H-W, Chen X, Cheng Z, et al. NIR-light-induced surface-enhanced Raman scattering for detection and photothermal/photodynamic therapy of cancer cells using methylene blue-embedded gold nanorod@ SiO₂ nanocomposites. *Biomaterials*. 2014; 35: 3309-3318.
115. Master A, Livingston M, Sen Gupta A. Photodynamic nanomedicine in the treatment of solid tumors: Perspectives and challenges. *Journal of Controlled Release*. 2013; 168: 88-102.
116. Liu D, Huang X, Wang Z, Jin A, Sun X, Zhu L, et al. Gold nanoparticle-based activatable probe for sensing ultralow levels of prostate-specific antigen. *ACS nano*. 2013; 7: 5568-5576.
117. Zhu J, Chang H, Li J-J, Li X, Zhao J-W. Dual-mode melamine detection based on gold nanoparticles aggregation-induced fluorescence "turn-on" and "turn-off" of CdTe Quantum Dots. *Sensors and Actuators B: Chemical*. 2016.
118. Monreal RC, Antosiewicz TJ, Apell SP. Diffuse surface scattering in the plasmonic resonances of ultralow electron density nanospheres. *The journal of physical chemistry letters*. 2015; 6: 1847-1853.
119. Zheng T, Bott S, Huo Q. Techniques for accurate sizing of gold nanoparticles using dynamic light scattering with particular application to chemical and biological sensing based on aggregate formation. *ACS Applied Materials & Interfaces*. 2016.
120. Kim Y, Macfarlane RJ, Jones MR, Mirkin CA. Transmutable nanoparticles with reconfigurable surface ligands. *Science*. 2016; 351: 579-582.
121. Ma C, Wang W, Mulchandani A, Shi C. A simple colorimetric DNA detection by target-induced hybridization chain reaction for isothermal signal amplification. *Analytical biochemistry*. 2014; 457: 19-23.
122. Yin H-q, Jia M-x, Shi L-j, Liu J, Wang R, Lv M-m, et al. Evaluation of a novel ultra-sensitive nanoparticle probe-based assay for ricin detection. *Journal of immunotoxicology*. 2014; 11: 291-295.
123. Ilkhani H, Sarparast M, Noori A, Bathaie SZ, Mousavi MF. Electrochemical aptamer/antibody based sandwich immunosensor for the detection of EGFR, a cancer biomarker, using gold nanoparticles as a signaling probe. *Biosensors and Bioelectronics*. 2015; 74: 491-497.
124. Hutter E, Maysinger D. Gold-nanoparticle-based biosensors for detection of enzyme activity. *Trends in pharmacological sciences*. 2013; 34: 497-507.
125. Zijlstra P, Paulo PM, Orrit M. Optical detection of single non-absorbing molecules using the surface plasmon resonance of a gold nanorod. *Nature nanotechnology*. 2012; 7: 379-382.
126. Szymanski HA. *Raman spectroscopy: theory and practice*: Springer Science & Business Media. 2012.
127. Yamamoto YS, Ishikawa M, Ozaki Y, Itoh T. Fundamental studies on enhancement and blinking mechanism of Surface-Enhanced Raman Scattering (SERS) and basic applications of SERS biological sensing. *Frontiers of Physics*. 2014; 9: 31-46.
128. Vendrell M, Maiti KK, Dhaliwal K, Chang Y-T. Surface-enhanced Raman scattering in cancer detection and imaging. *Trends in biotechnology*. 2013; 31: 249-257.
129. Patra PP, Chikkaraddy R, Tripathi RP, Dasgupta A, Kumar GP. Plasmo-fluidic single-molecule surface-enhanced Raman scattering from dynamic assembly of plasmonic nanoparticles. *Nature communications*. 2014; 5.
130. Conde J, Bao C, Cui D, Baptista PV, Tian F. Antibody-drug gold nanoantennas with Raman spectroscopic fingerprints for in vivo tumour theranostics. *Journal of Controlled Release*. 2014; 183: 87-93.
131. Fu C, Gu Y, Wu Z, Wang Y, Xu S, Xu W. Surface-enhanced Raman scattering (SERS) biosensing based on nanoporous dielectric waveguide resonance. *Sensors and Actuators B: Chemical*. 2014; 201: 173-76.
132. Zhan L, Zhen SJ, Wan XY, Gao PF, Huang CZ. A sensitive surface-enhanced Raman scattering enzyme-catalyzed immunoassay of respiratory syncytial virus. *Talanta*. 2016; 148: 308-312.
133. Xu L-J, Lei Z-C, Li J, Zong C, Yang C-J, Ren B. Label-free surface-enhanced Raman spectroscopy detection of DNA with single-base sensitivity. *Journal of the American Chemical Society*. 2015; 137: 5149-5154.
134. Xiao L, Harihar S, Welch DR, Zhou A. Imaging of epidermal growth factor receptor on single breast cancer cells using surface-enhanced Raman spectroscopy. *Analytica chimica acta*. 2014; 843: 73-82.
135. Luo Z, Li W, Lu D, Chen K, He Q, Han H, et al. A SERS-based immunoassay for porcine circovirus type 2 using multi-branched gold nanoparticles. *Microchimica Acta*. 2013; 180: 1501-1507.
136. Wang W, Pang Y, Yan J, Wang G, Suo H, Zhao C, et al. Facile synthesis of hollow urchin-like gold nanoparticles and their catalytic activity. *Gold Bulletin*. 2012; 45: 91-98.

137. Gopinath V, Priyadarshini S, MubarakAli D, Loke MF, Thajuddin N, Alharbi NS, et al. Anti-Helicobacter pylori, cytotoxicity and catalytic activity of biosynthesized gold nanoparticles: Multifaceted application. *Arabian Journal of Chemistry*. 2016.
138. Wang Y, Liu Y, Luehmann H, Xia X, Wan D, Cutler C, et al. Radioluminescent gold nanocages with controlled radioactivity for real-time in vivo imaging. *Nano letters*. 2013; 13: 581-585.
139. Bisker G, Yeheskely-Hayon D, Minai L, Yelin D. Controlled release of Rituximab from gold nanoparticles for phototherapy of malignant cells. *Journal of Controlled Release*. 2012; 162: 303-309.
140. Biju V. Chemical modifications and bioconjugate reactions of nanomaterials for sensing, imaging, drug delivery and therapy. *Chemical Society Reviews*. 2014; 43: 744-764.
141. Robinson R, Gerlach W, Ghandehari H. Comparative effect of gold nanorods and nanocages for prostate tumor hyperthermia. *Journal of Controlled Release*. 2015; 220: 245-252.
142. Dmitriev V, dos Santos TL, da Costa KQ. Resonant Properties of Gold Bowtie Nanoantennas of Modified Triangular Geometries for Optical Sensors Applications. *Journal of Microwaves, Optoelectronics & Electromagnetic Applications*. 2013; 12.