

## Review Article

# Role of Phytoconstituents in the *Aloe*-Mediated Nanoparticles

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Currently, green, single-pot biomimetic, and/or biological methods of Nanoparticle (NP) synthesis are preferred over chemical and physical methods because of their rapidity, eco-friendliness, non-pathogenicity, and economic attributes. In addition, these biosynthesis methods exclude the use of high temperature, energy, pressure, and toxic chemicals. Through biosynthetic methods, nanotechnology is related to biotechnology. This has advanced nanobiotechnology which is the development of eco-friendly and biosynthetic nanomaterials/nanoparticles. Therefore, nowadays, biogenic or green synthesis of NPs using plants has emerged as a potential nano-factory, and their applications are based on phytochemicals. The chemical constituents identified in *Aloe* plants include vitamins, minerals, enzymes, polysaccharides, fatty acids, indoles, hydrocarbons, carboxylic acids, aldehydes, ketones, phenolic compounds, phytosterols, pyrimidines, alkaloids, etc. Each of these chemicals has potential biological activities that make them essential for in nanotechnology. *Aloe* phytochemicals play a crucial role in the fabrication of *Aloe*-based NPs by acting as reducing, capping, and stabilizing agents that control the shape and size of the formed NPs. In addition, these phytochemicals play a significant role in the applications of *Aloe*-based NPs owing to their biological properties. Moreover, it is important to understand the role of phytoconstituents from different *Aloe* species and in every part of the plants using nanotechnology.

**Keywords:** *Aloe* species; Phytoconstituents; Nanoparticles; Fabrication; Applications**Introduction**

The development of nanotechnology is a modern multidisciplinary science involving the fields of chemistry, physics, biology, and engineering, the production of Nanoparticles (NPs), both in nature and by humans [1]. The area of nanotechnology is one of the most dynamic views in current-day material science [2]. The word “nanotechnology” refers to the use of matter with dimensions ranging from one to a hundred nanometers at the molecular or atomic level [3]. “Nano” is a Greek word. “Nanos”, means “dwarf, tiny, or very small”. Nowadays, the terms like “creation,” “exploitation,” and “synthesis” are associated with nanotechnology [4]. A nanoparticle is characterized as a little item that acts in the general unit as far as its transport and properties in nanotechnology [5]. There are various chemical and physical methods to synthesize Nanoparticles (NPs). Among them, the sol-gel process, chemical precipitation, chemical vapor deposition, hydrothermal, and microwave methods have been reported mostly [6]. However, these methods are not effective in many aspects. Therefore, currently, green synthesis, single-pot biomimetic, and/or biological methods of synthesis are preferred over chemical and physical methods due to their rapidity, eco-friendliness, non-pathogenic, and economical attributes. Besides, these biosynthesis methods exclude the use of high temperature, energy, pressure, and toxic chemicals [7]. Therefore, nowadays, biogenic or green synthesis of (NPs) using bacteria, fungi, actinomycetes, algae, and higher plants have emerged as potential nano factories [8-10] and their applications are based on the phytoconstituents of these living things. Through

biosynthesis methods, nanotechnology is related to biotechnology. This has been advanced in nanobiotechnology which is the development of eco-friendliness and biosynthetic nanomaterials/nanoparticles [11].

The green synthesis of nanomaterials such as silver [12], zinc oxide [13], magnesium oxide [14], gold [15], cerium oxide [16], copper oxide [17], titanium dioxide [18], activated carbon [19], palladium [20] and tin oxide [21] has been conducted extensively in recent years. The reasons that make green synthesis very important are due to the simple work-up procedure, environmentally benign nature, reusable, low cost, and ease of isolation [22]. Nanoparticles have a novel or superior behavior with defined shape and size. This is because of the high surface area to volume ratio. The physicochemical parameters of Nanoparticles (NPs) are different from that of bulk or large material and single atom and molecule [23,24]. The size of Nanoparticles (NPs) is 1-100 nm with their unique surface, optical, electrical, magnetic, and biological properties [25].

Out of various biomaterials employed for these purposes, plant extracts have attracted much attention due to their effectiveness, availability, and green characteristics [26,27]. Additionally, it has been noticed that the NPs prepared using plant extracts are more stable, cheap, monodispersed, and take less time to reduce [28]. The influence of the added particles like phytocomponents such as polysaccharides, flavanones, terpenoids, etc. attached to the nanoparticle can change its overall properties, especially the antimicrobial property [29]. In addition to plant extracts have an intense array of antioxidants such

**Table 1:** Reports of *Aloe* species phytoconstituents.

<i>Aloe</i> spp.	Part of Part	Phytoconstituents Present	Ref.
<i>A. vera</i>	Leaf (gel)	Tannins, saponins phlobatannins, flavonoids, anthraquinones, terpenoids, steroids, alkaloids, carbohydrates, and glycosides	[44]
<i>A. gilbertii</i>	Root	Alkaloids, anthraquinones, terpenoids, and flavonoids	[45]
<i>A. elegans</i>	Root	anthraquinones, terpenoids, phenols, saponins, tannins, glycosides	[45]
<i>A. tormentorii</i>	Leaf	alkaloids, anthraquinones, terpenes, phenols, flavonoids, coumarins, saponins, and tannins	[46]
<i>A. adigratana</i> Reynolds	Leaf (Gel)	Alkaloids, glycosides, steroids, carbohydrates, fixed oils and fats, amino acids and proteins, flavonoids, tannins, terpenoids, and saponins,	[47]
<i>A. arborescens</i>	Leaf	Alkaloids, terpenoids, steroids, flavonoids, tannins, and reducing sugars	[48]
<i>A. ferox</i>	Leaf	Phenols, flavonoids, flavonols, proanthocyanidins, tannins, alkaloids, and saponins	[49]
<i>A. striata</i>	Leaf	Flavonoids, terpeneoids, and aromatic compounds	[50]
<i>A. vera</i>	Leaf	Alkaloids, glycosides, reducing sugars, phenolic compounds, steroids, terpenoids, flavonoids, tannins, and saponin glycosides	[51]
<i>A. turkanensis</i>	Whole Leaf	Tannins, anthraquinones, terpenoids/steroids, saponins, and alkaloids	[52]
<i>A. perryi</i>	Flower	Glycosides, phytosterols, proteins, and amino acids, flavonoids, phenols, and carbohydrates	[53]
<i>A. pulcherrima</i> Gil. and Seb.	Leaf (latex)	Anthraquinones, Flavonoids, Saponins, Glycosides, Tannins, Phenols, and Alkaloids	[54]

as polyphenols [30,31], reducing sugars [32], nitrogenous bases, and amino acids [33], which can produce nanoparticles of metal and metal oxide from metal ions [34].

*Aloe* species can store water and important chemical constituents in their swollen and succulent leaves because of their ability to survive in conditions such as hot and dry, which makes them a unique source of phytochemicals [35]. *Aloe* plants have been widely known and used for centuries as topical and oral therapeutic agents due to their health, beauty, medicinal, and skin care properties [36]. The range of chemical constituents of the *Aloe* species can be used in preparing beauty and cosmetics, medicinal and pharmaceutical, personal care and toiletry products, and bittering agents in alcoholic drinks, and they are also grown as ornamental plants [37]. The phytoconstituents and bioactivity of *Aloe* spp. have attracted research interest since the trade in 'drug Aloes', prepared from the leaf exudate, expanded rapidly in the 19th century [38]. But nowadays, the applications of *Aloe* plants do not limited to the *Aloe* alone; it is incorporated into different substances to give novel ideas such as chemical synthesis and drug delivery [39]. Currently, many researchers are focused on the incorporation of *Aloe* extracts into substances such as metal/metal oxides at the nanoscale. This is due to the *Aloe* species having a variety of phytoconstituents responsible for the target application. Therefore, these phytochemicals have a great role in *Aloe*-based nanotechnology [11].

However, due to several complexities in the identification of exact chemical components responsible for the synthesis and applications of nanoparticles, the green synthesis of nanoparticles becomes challenging. Moreover, there is a lack of a comprehensive review that presents a general idea about the roles of phytochemicals in both fabrication and applications of *Aloe*-mediated NPs. Herein; the review summarizes the recent update on these ideas somewhat. In addition to that almost all kinds of literature, fabricated NPs from leaves of *Aloe* especially, *A. vera*. But other parts of the plants like flowers and roots are also rich in bioactive compounds. Therefore, it is very important to synthesize NPs from other than leaves of *Aloes* and identify the roles of responsible phytoconstituents in them.

## *Aloe* Phytoconstituents

The chemical constituents that have been identified in *Aloe* plants include vitamins, minerals, enzymes, simple and complex polysaccharides, fatty acids, indoles, hydrocarbons, dicarboxylic acids, aldehydes, ketones, phenolic compounds, phytosterols pyrimidines, and alkaloids with potential biological and toxicological activities [40-42]. The biological properties of *Aloe* such as anti-inflammatory, antimicrobial, antitumoral, and antioxidant are due to various compounds of *Aloe* extracts. These properties and activities are synergistic rather than one single class of compounds [43]. If the same climatic, geographic, harvesting time, solvent system, extraction, etc. conditions are applied to the same *Aloe* species, the part of the plants makes difference in the presence and absence of the phytochemicals. Table 1 shows the different *Aloe* species' phytochemicals from the different parts of the plants.

Several constituents from various classes such as alkaloids, anthrones, chromones, flavonoids, glycoproteins, naphthalenes, and pyrones have been isolated from different *Aloe* spp [55]. Aloin, Aloesin, Aloenin, Aloeresin, *Aloe*-emodin, apigenin, acemannan, and chrysophanol are some examples of such bioactive compounds (Table 2 and Figure 1).

The chemical structure of some *Aloe* species phytochemicals components is represented in (Table 2) is shown in (Figure 1) [36,64-66].

The most commonly used medicinal *Aloe* parts, leaves are heterogeneous and can be divided into three major parts, (i) the outer green epidermis, primarily consisting of structural components, known as skin/peel; (ii) the outer pulp region below the epidermis, consisting of vascular bundles where the yellow bitter latex/sap is derived; and (iii) the inner leaf pulp, consisting of *Aloe* gel and containing parenchyma cells. Regarding the different compositions of these leaf portions, they are also likely to have distinct classes of bioactive compounds, which are believed to contribute to the different biological properties of leaves [58]. Although leaves are the most used part of the plant, recently some studies have reported the

**Table 2:** The main phytoconstituents of *Aloe* spp.

Class of phytoconstituents	Components	Ref.
Carbohydrates	Pure mannan, acetylated mannan, acetylated glucomannan, glucogalactomannan, galactan, galactogalacturan, arabinogalactan, xylan, pectic, cellulose, acemannan, glucose, galactose, manose, fucose, and aldopentose	[56,57]
Anthraquinones	<i>Aloe</i> -emodin, <i>Aloesaponarin</i> , desoxyerythrolaccin, chrysophanol, 1,5-dihydroxy-3-hydroxy methylanthraquinone, helminthosporin, 7-hydroxy <i>Aloe</i> emodin, nat <i>Aloe</i> emodin and its ester nat <i>Aloe</i> emodin-8-methyl ester, <i>Aloechrysone</i> , <i>Aloesaponol</i> , bianthracene O-glycosides: <i>Aloe</i> emodin-11-O-rhamnoside, <i>Aloesaponol</i> -6-O-glucoside, <i>Aloesaponol</i> -8-O-glucoside, <i>Aloesaponol</i> -O-methyl-4-O-glucoside, and elgonicardine	[58,36]
Anthrones	Aloin A and B (collectively called aloin and also often referred to as barbaloin), 5-hydroxyaloin A, 7-hydroxyaloin, 10-hydroxyaloin B, 5-hydroxyaloin A 6'-O-acetate, 7-hydroxyaloin-6'-O-monoacetate, homonataloin, nataloin, aloinoside, barbendol, <i>Aloe</i> -emodinanthrone, chrysophanolanthrone, <i>Aloe</i> emodin-10-C-rhamnoside, 8-O-methyl-7-hydroxyaloin, 6'-O-cinnamoyl-8-O-methyl-7-hydroxyaloin, 6'-O-p-coumaroyl-7-hydroxyaloin, 7-hydroxyaloin-4',6'-O-diacetate, 6'-O-cinnamoyl-5-hydroxyaloin A, microstigmin A, littoraloin, littoraloside, microdantin, homonataloside, and deacetylittoraloin.	[57,59]
Chromones	<i>Aloesin</i> , iso- <i>Aloeresin</i> , 7-O-methyl <i>Aloesin</i> , 8-[C-β-D-[2-O-(E)- cinnamoyl]glucopyranosyl]-2-[(R)- 2-hydroxypropyl]-7-methoxy-5-methylchromone, 8-C-glycosyl-7-O-methyl <i>Aloediol</i> , and 8-C-glycosyl-7-methoxy-(S)- <i>Aloesol</i>	[36,57]
Coumarins	Feralolide and dihydroisocoumarin glycoside	[57]
Flavonoids	Naringenin, dihydroisorhamnetin, apigenin, isovitexin, isorhamnetin, genistein, saponarin, kaempferol, and quercetin	[36,60]
Alkaloids	N-methyltryamine, O,N-dimethyltryamine, γ-coniceine, coniine	[59]
Pyrans and pyrenes	Bisbenzopyran, <i>Aloenin</i> , <i>Aloenin</i> aglycone, <i>Aloenin</i> acetal, <i>Aloenin</i> B, and <i>Aloe</i> -2"-p-O-coumaroyl ester,	[59]
Benzene, naphthalene and furan derivatives	Protocatechuic acid, methyl-p-coumarate, pluridone, isoeuletherol, isoeuletherol-5-O-glucoside, feroxidin, feroxidin A, feroxidin B, plicataloside, 5-OH-3-methylnaphto[2,3-c]furan-4(1H)-one, 3-methylnaphto[2,3-c]furan-4(9H)-one, and 3-methylnaphto[2,3-c] furan-4,9-dione	[57]
Sterols	Cholesterol, campesterol, β-sitosterol, and lupeol	[57,59]
Proteins	Lectins, lectin-like substance	[57]
Enzymes	Alkaline phosphatase, amylase, cyclooxygenase, cyclooxygenase, lipase, oxidase, phosphoenolpyruvate carboxylase, and superoxide dismutase	[60,61]
Vitamins	B1, B2, B6, B12, C, E, β-carotene, choline, folic acid, α-tocopherol	[61,62]
Inorganic compounds	Calcium, chlorine, chromium, copper, iron, magnesium, manganese potassium, phosphorous, sodium, and zinc.	[57,62]
Miscellaneous including lipids	Arachidonic acid, γ-linolenic acid, triglycerides, triterpenoid, gibberillin, lignins, potassium sorbate, salicylic acid, and uric acid	[61,63]

bioactive roots [67] and flowers [68] of the plant. *Aloe* species have become of great interest to researchers who have tried to identify the compounds responsible for these beneficial effects.

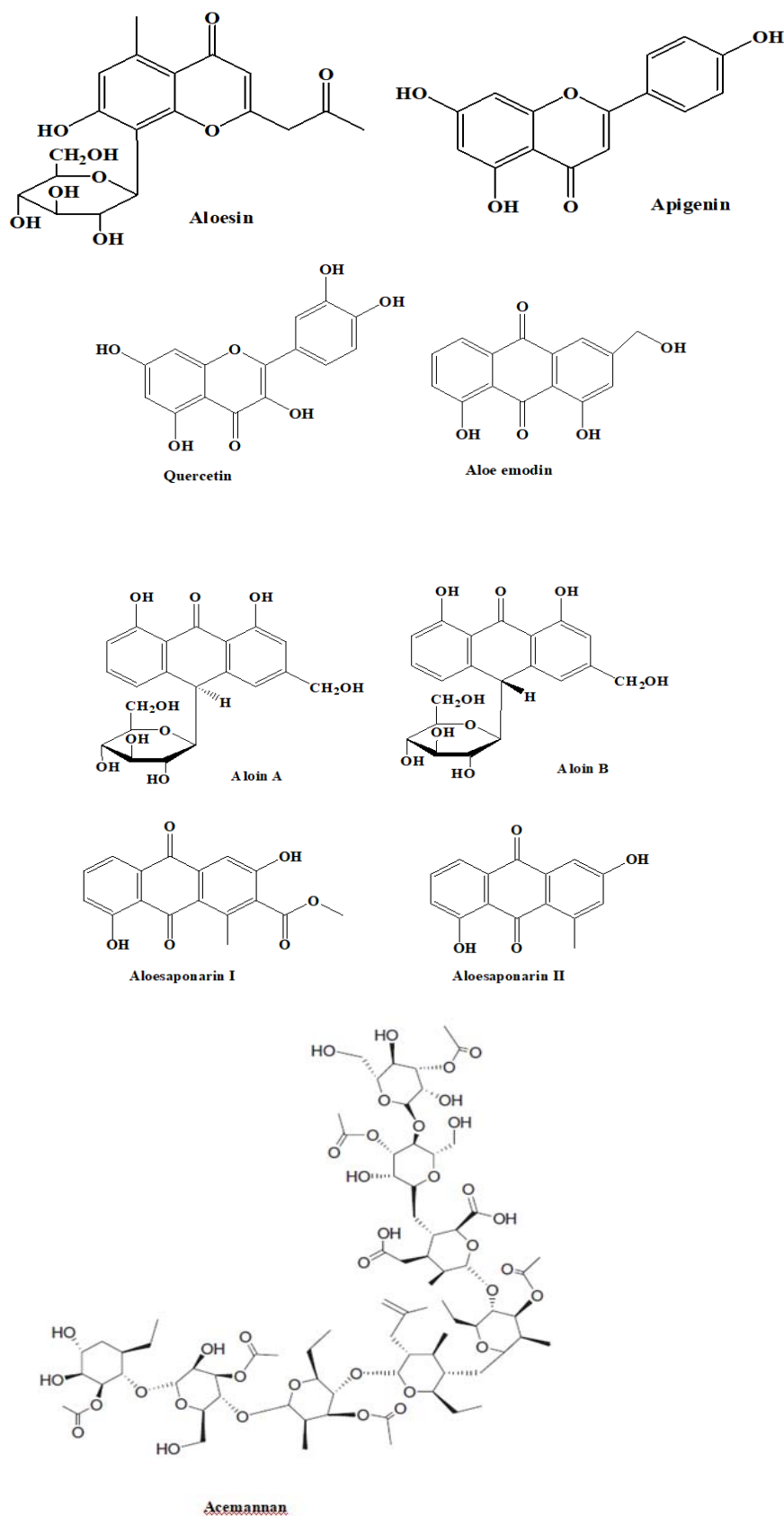
## Aloe-Based Nanoparticles

Recently, *Aloe*-based nanoparticles have been utilized for their wide applications. The fabrication of NPs by using *Aloe* species is due to chemical compounds present in the *Aloe* genus [69]. In works of literature, numerous *Aloe*-mediated NPs have been fabricated along with their various applications. In *Aloe*-based NPs the leaf gel [70], leaf skin (peel) [71], whole leaves [72], and/or flower [73] of *Aloe* species with metal/ metal oxide have been conducted. the metal and metal oxide of *Aloe* based NPs such as silver NPs, gold NPs, selenium NPs, copper NPs, iron NPs, iron oxide NPs, silver oxide NPs, zinc oxide NPs, magnesium oxide NPs, titanium oxide NPs, and indium oxide NPs with various applications such as cytotoxicity, UV protection, antibacterial activity, catalytic activity, antibiofilm potential, photocatalytic activity, antifungal, and antioxidant activities have been described. *Aloe*-based NPs fabrication is affected by various factors such as type of metal, different *Aloe* spp., method of NPs formation, temperature, pH, and type of solvent used [28]. In addition to that, the part of the *Aloe* plant such as leaf skin, leaf gel, leaf latex, whole leaf, root, and flower is also a great factor to make difference in the fabrication of *Aloe*-based NPs due to the phytochemicals present in each part are different.

In the synthesis of *Aloe* mediated NPs, *Aloe* extract is prepared separately before being added to precursors. The extracts can be

prepared from different parts of the plant such as the leaf, flower, and root. The mature, healthy, and fresh *Aloe* parts are used to utilize for this purpose. The selected part of the plant is washed with distilled water to remove any dirt or debris on the surface [70]. If the synthesis of NPs is based on whole leaf, the leaf extract is prepared by cutting it finely, if the skin of the plant the is targeted, the extract is prepared by peeling off the leaf carefully using a sharp knife and if the gel is needed, the leaf is slit longitudinally into half, the skin is discarded, the gel is scraped off by sharp-edged spoon/knife from the inner leaf into a container. If the latex/sap part of the *Aloe* is the target extract, the cut leaf is kept 45° to obtain latex. Flower and root parts of the plant are also used to prepare *Aloe* extract. The identified *Aloe* extracts are ground to be kept for the next steps. In the literature, there are different methods to make extracts to store for further use. Among the different methods, boiling the prepared extract with distilled water for certain minutes is the common method [73-75].

The NPs that are prepared from the addition of plant extracts like *Aloe* extract to precursors such as metals and metal oxides green synthesis. Based on the aim of nanoparticle synthesis, a different form of green synthesis can be followed. During the synthesis of NPs, the formation of NPs is indicated by physicochemical properties. Among the many physicochemical properties reduction, a color change is mandatory to show the completion of NPs [76]. This shows that there is the reduction in the chemical reaction between *Aloe* extracts and precursors because *Aloe* species possess reducing agent phytochemicals.



**Figure 1:** The chemical structures of some phytoconstituent components of *Aloe* species.

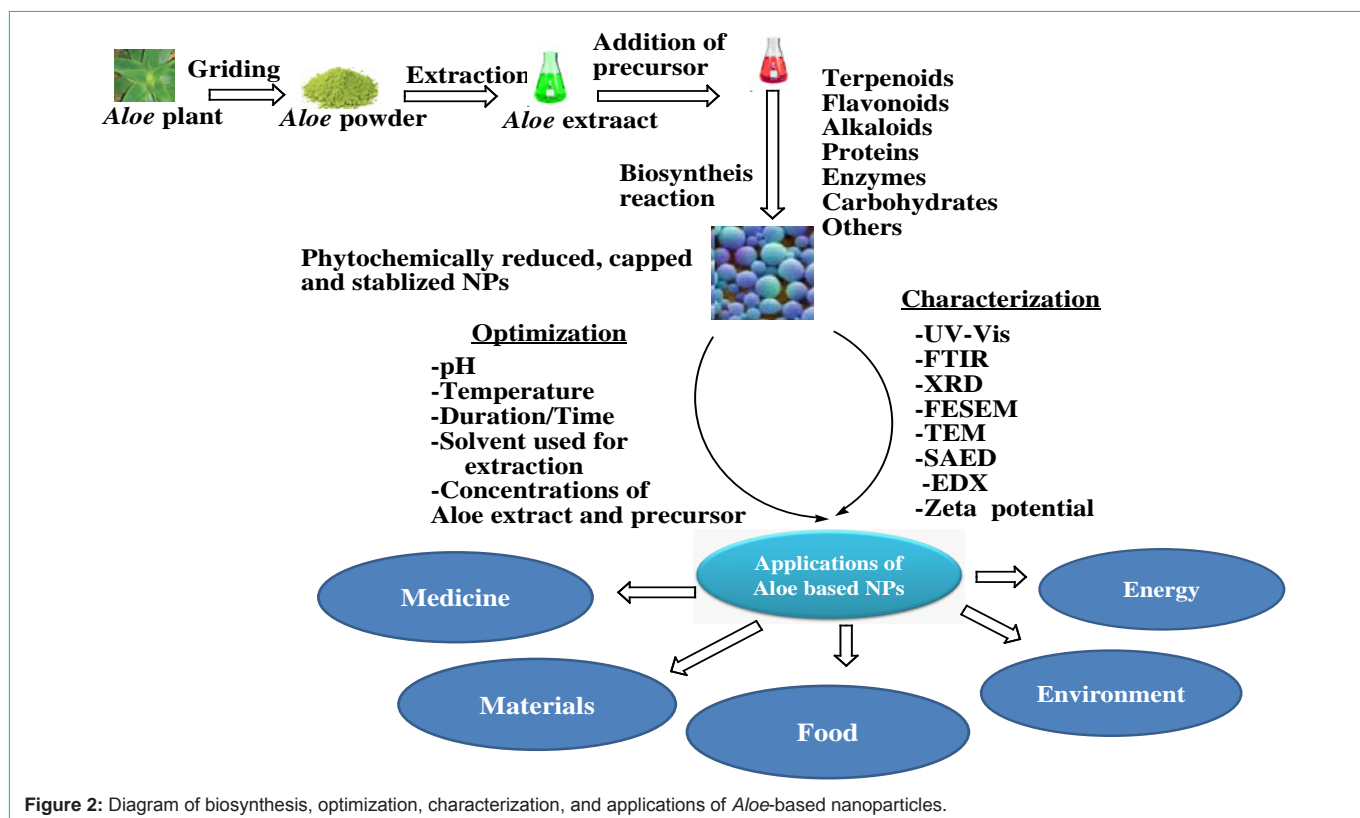
## Role of Phytoconstituents in Aloe Based Nanoparticles

### Role of Phytoconstituents in the Fabrication of Nanoparticles

It has been described that the plant chemical compositions are used for the NPs fabrication because they act as reducing, capping, and/or stabilizing agents [77]. Some of these bioactive molecules act as electron shuttles in metal reduction, while other constituents are responsible for the capping of resulting NPs, which not only controls the aggregation of NPs but also results in post-surface modification of NPs [78]. The phytochemicals that are responsible for these purposes of the *Aloe-based* NPs are anthrones, anthraquinones, chromones, polysaccharides, proteins, alkaloids, flavonoids, and Miscellaneous bioactive compounds, (Table 1, 2, and Figure 1). Hydroxyl and carboxylic groups present may act as reducing agents and stabilizing agents in the synthesis of nanoparticles [79]. The functional groups such as phenolics and alkaloids are responsible for capping and stabilizing nanoparticles reduced [80]. The size and stability of the formed nanostructure are also controlled by the reduction mechanism. The stability of nanoparticles can be attributed to the formation of stable bonding between metallic nanoparticles and phytochemicals present in the *Aloe* extract [81]. There are various roles of *Aloe* phytochemicals in the formation of *Aloe-based* NPs. However, roles such as reducing, capping, and stabilizing agents are very important in the characterizations and applications of *Aloe-based* NPs. These three properties are interrelated to one another. If the formed NPs are reduced or capped to precursor, then it stays stable. The stable NPs can be applied to the target applications.

Role of phytoconstituents in reducing NPs: Reducing agents have the role of driving electrons from the solution to the ions (usually metallic ones) to form atoms. In other words, they reduce the salts into the reduced form, which is usually insoluble [82]. A possible mechanism of the formation of *Aloe-based* NPs from *Aloe* spp. and the phytochemical involved are presented in (Figures 3 and 4). There is the presence of a -OH group in most phytochemicals obtained from *Aloe* spp. and this -OH served as a reducing agent, converting metal ions into metal/metal oxide NPs. Also, carbonyl functional groups are present in the phytochemical of *Aloe* spp. play a significant role in NPs fabrication [28]. In another study, the production of silver nanoparticles is demonstrated by the sharp peak around 400 nm for aloin-mediated silver nanoparticles in the UV-Vis spectrum, which indicates the availability of reducing functional groups in aloin [74]. The role of the functional groups in acemannan molecules in the formation of AgNP has been described as acting as a reducing and stabilizing agent, (Figure 4) [29]. The ZnO-biomolecule complex formation is due to the linkage between the  $Zn^{+2}$  ions and the functional group hydroxyl that present in the biomolecules like aromatic compounds like aloin (polyphenol) present in *A. vera* extract acts as the reducing agent for the ZnO NP synthesis [11].

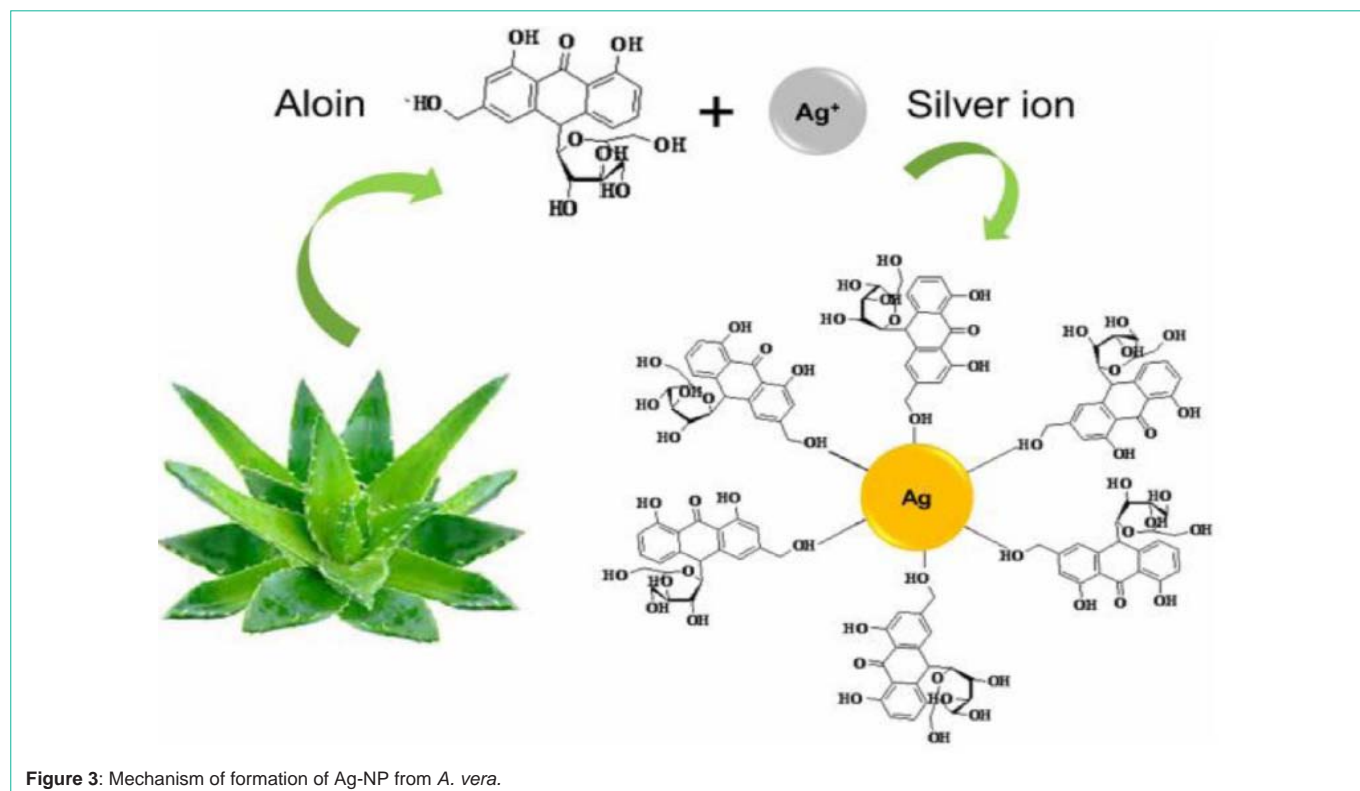
**Role of phytoconstituents in capping NPs:** The environmentally friendly single-step protocols using plant extracts without involving any extrinsic surfactants, capping agents, and/or templates have been explored for metal NPs synthesis [83]. A capping agent is an amphiphilic molecule consisting of a polar head group and a nonpolar hydrocarbon tail and the functionality of the capping agent depends upon both the parts [84]. The surface chemistry and size distribution of nanoparticles get altered after capping with biocompatible



**Figure 2:** Diagram of biosynthesis, optimization, characterization, and applications of *Aloe-based* nanoparticles.

**Table 3:** Applications and responsible phytochemicals of *Aloe* based NPs.

Precursors	<i>Aloe</i> spp.	Part of <i>Aloe</i>	Applications	Responsible phytoconstituents	Ref.
Ag	<i>A. vera</i>	Leaf (Gel)	Antioxidant activity	Tannins, flavonoids, and proteins	[104]
Se	<i>A. vera</i>	Leaf	Antioxidant activity	Hydroxyl groups, methyl, carboxylic acid, and amine	[105]
ZnO	<i>A. vera</i>	Leaf	Antioxidant, antibacterial, and antiproliferative activities	Alkaloids, carbohydrates, flavonoids, phenolic, saponins, tannins, and terpenoids	[106]
Ag	<i>A. vera</i>	Leaf	Antibacterial and Antimalarial	Amine groups and carboxylic acids	[107]
Ag /Ag <sub>2</sub> O	<i>A. vera</i>	Leaf	UV protection and antibacterial	Anthraquinone compounds, alcoholic compounds, proteins, polyphenols, terpenoids alkaloids, flavonoids, and polysaccharide	[108]
Au	<i>A. vera</i>	Leaf (gel)	Antibacterial Activity	Alcohol, phenol, carbonyl compounds, and amine	[109]
ZnO	<i>A. vera</i>	Leaf (skin)	Antibacterial Activity	Phenolic compounds, terpenoids and proteins	[110]
ZnO	<i>A. socotrina</i>	Leaf	Antibacterial Activity	Amines and hydroxyl groups	[39]
CuO	<i>A. vera</i>	Leaf	Antibacterial Activity	Hydroxyl groups, alkenes, and Alkanes	[111]
Se	<i>A. vera</i>	Leaf	Antibacterial and antifungal activities	Hydroxyl groups, carboxylic acid, and amine	[112]
Ag	<i>A. vera</i>	Leaf	Antifungal Activity	Amide and alcoholic hydroxyl groups	[74]
ZnO	<i>A. vera</i>	Leaf (gel)	Antimicrobial Activity	Alcohols, phenols, amines, and carboxylic acids	[113]
Ag	<i>A. vera</i>	Leaf (gel)	Antimicrobial Activity	Flavanones or terpenoids	[114]
Ag	<i>A. vera</i>	Leaf (gel)	Wound healing	Amino acids, organic acids, and mineral elements	[115]
TiO <sub>2</sub>	<i>A. barbadensis</i>	Whole leaf	Antibiofilm Potential	Terpenoids, flavonoids, and proteins	[116]
Fe <sub>3</sub> O <sub>4</sub>	<i>A. vera</i>	Leaf (gel)	Cytotoxicity Assessment	Hydroxyl group, phenolics, and carbohydrate	[117]
TiO <sub>2</sub>	<i>A. vera</i>	Leaf (gel)	Photocatalytic Activity	Amino groups, alcohols, phenols, and amines	[76]
Ag	<i>A. vera</i>	Leaf	Catalytic activity	Phenols, alcohols, carboxylic acids, amines, and minerals	[118]

**Figure 3:** Mechanism of formation of Ag-NP from *A. vera*.

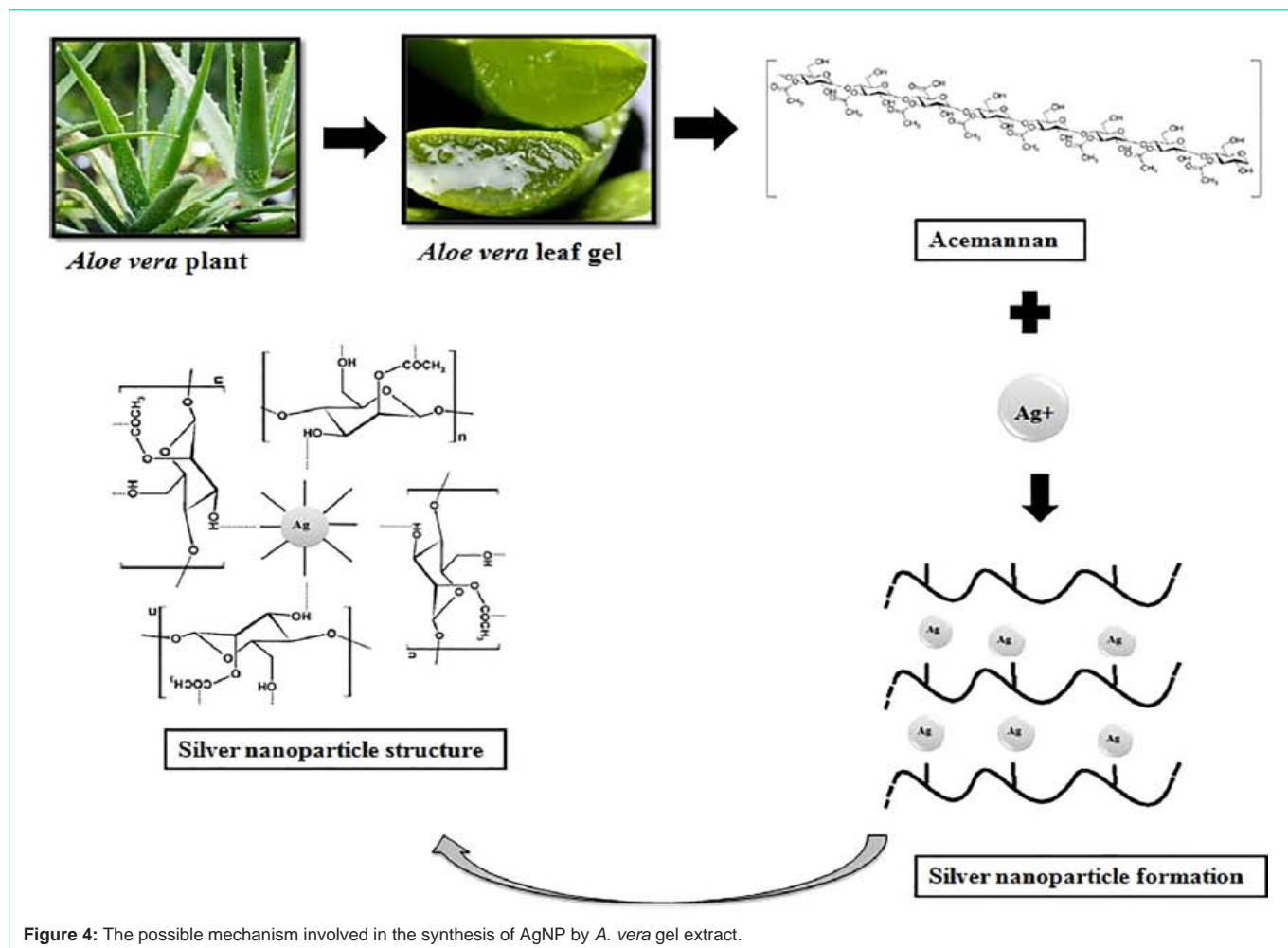


Figure 4: The possible mechanism involved in the synthesis of AgNP by *A. vera* gel extract.

surfactants [85]. The biological activities of nanoparticles are enhanced by surface capping as the role of phytochemicals. The controlled size, morphology, and surface composition achieved by nanoparticles' capping are crucial in determining the vital application of nanoparticles [86]. It has been reported that capping agent molecules prevent nanoparticles from aggregation and oxidation to stabilize the NPs [87]. *Aloe* species have phytochemicals and/or functional groups responsible for capping agents [88,89]. The presence of bioactive molecules in *Aloe* extract was associated with  $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>NPs surface, also illustrated by their stretching/bending vibrations. The appearance of prominent bands indicated the surface association of O–H bearing carbohydrates/alcohols, 1° and 2° amines, amide-I, proteins, and aromatic amines, respectively. Due to these capping agents, the overall, FTIR data suggested the stability of *Aloe* extract bioactive molecules capped/absorbed on the surface of NPs [90]. The authors proposed the capping of reduced silver by acemannan as the possible chemistry involved in the formation of AgNP as represented in (Figure 4). The surrounding acemannan is a surfactant and inhibits the AgNP agglomeration. In another study, the proteins responsible for the synthesis of CuNPs act as surface coating molecules that keep away from the internal agglomeration of the particles [73]. The biomolecules surrounding the NPs decomposed into gaseous products at high temperatures resulting in NPs during

calcination or annealing [91]. By developing the green chemistry of phytochemical coating on metal nanoparticle surfaces, their toxicity could be reduced [92].

**Role of phytoconstituents in stabilizing NPs:** It is well known that nanoparticles in their free form are thermodynamically unstable due to high surface energy. Due to Brownian motion, the high surface energy nanoparticles collide and the final state of nanoparticles is dictated by the type of interaction between the colloidal nanoparticles [93]. The intrinsically green approach involves the phytochemicals for the stabilization of NPs [94]. GC-MS analysis of *A. vera* leaf extract of ZnONPs revealed the predominance of compounds like ethanone, 1-phenyl, guanosine, pentadecanoic acid, and tetraconate in *Aloe* extract, which might be responsible for conferring stability to associated ZnONPs, besides proteins and other auxiliary phytochemicals [75]. The stability of CuO nanoparticles may be due to the free amino and carboxylic groups of *A. barbadensis* that have interacted with the copper surface [95]. The stability of NiO nanorods may be due to the free carboxylic and amino groups that interact with the surface of the nickel has been stated in the same way. In both cases, the proteins present in the medium prevent agglomeration and help the stability of NiO by covering the metal nanoparticles [96]. The formation of NPs complexes is due to the biomolecules present in *Aloe*. Due to these biomolecules having a

strong chelating ability, it is readily attached and adsorbed onto the NP surfaces. As a result stability of NPs is achieved [11].

### Role of Phytoconstituents in the Applications of *Aloe* Based Nanoparticles

In addition to other plant-based NPs [97], the *Aloe* plants have a variety of applications because they are rich in active components [28]. Researchers have demonstrated the possible mechanisms of action of medicinal plants and their active ingredients or active compounds/ phytochemicals, which may exert these mechanisms individually or in combination with other compounds present in the plants [98]. The influence of additional particles of *Aloe* phytochemicals attached to the nanoparticle can change its overall properties, especially in medical applications such as antimicrobial, anticancer, antioxidant, etc. [29]. Antibacterial activities of *Aloe*-based NPs have been described by many authors with the responsible phytochemicals and functional groups [99]. The enhanced antibacterial activity of AgNPs synthesized using *A. vera* extract was described as it is attributed to active components in the extract especially, acemannan as the main reason [28]. In another study, the addition of *A.vera* in the Nanofiber Membranes (NFM) can increase the antibacterial effect of the NFM. This might be due to the presence of substances such as acemannan, anthraquinones, and salicylic acid in *A.vera*, resulting in its better antimicrobial activity [100].

In the literature, the peak separation potential of Lithium Titanate (LTO) is lower than the Green Synthesis of Lithium Titanate (GSLTO); possibly due to the utilization of *A.vera* extract, which contains various phytochemical compounds like tannin, saponin, flavonoids, etc. has been stated [69]. These phytochemical species also take a role in the electrochemical performance of the GSLTO sample [101]. There are many different types of phytochemicals with different structures. It has been suggested that the structures of phytochemicals are associated with their different activities [102]. The synergistic activities of these phytochemicals [63] and a synergistic effect between particles and phytochemicals [29] make nanobiotechnology an interested and applicable scientific research. It is concluded that the insulin-loaded nanoemulsion topical gel with *A. vera* showed a synergetic effect toward efficient healing of the wound in diabetes, and is more effective than the application of topical insulin alone [103]. Although the role of some phytochemicals has been described well, still it needs more clarification on which phytochemicals are more responsible for the *Aloe*-based NPs. (Table 3) indicates the combination of phytochemicals as well as functional groups in the applications of *Aloe*-based NPs. In this case, the idea of responsible phytochemicals shows the role of these phytochemicals in investigated applications.

### Limitations and Future Aspects

Despite being rich in all parts, the works of literature regarding *Aloe*-based NPs were limited to the leaf of *Aloe* species. Also among the leaf parts, the gel was under the consideration. But there was no clear evidence for the selection of leaf and/or gel only. Therefore, the *Aloe* plants' parts such as roots, flowers, etc. are rich in phytoconstituents and they can be used in the biosynthesis of *Aloe*-based NPs. Although many works have been done on *A.vera* and it is available everywhere, it should not be *A.vera* alone to make investigations like biosynthesis of NPs. Other *Aloe* species should be analyzed for their role in the

*Aloe*-mediated NPs.

### Conclusion

Currently, *Aloe*-based nanoparticles are very important in a broad area of study. The first reason for this idea is due to the unique nature of *Aloe* plants. They have a variety of chemical compositions with chemical and biological properties. Therefore, the incorporation of these *Aloe* biochemicals into another substance like metal or metal oxides make the biosynthesis of NPs which are very necessary for all aspects than other means of NPs syntheses. The second idea that brings the importance of *Aloe*-based nanoparticles is the new idea of nanotechnology. The combination of these points forms what is known as *Aloe*-based nanobiotechnology. Although the combination of *Aloe* phytochemicals and precursors is known, *Aloe* phytoconstituents have a great role in the formation and applications of *Aloe*-based NPs. Nowadays, this science has wide applications in medicine, food, environmental protection, material preparations, etc.

### Competing Interests

The author declares that he has no competing interests.

### Authors' Contributions

All parts of the manuscript were prepared by the corresponding author.

### References

- Benelmekki M. An introduction to nanoparticles and nanotechnology. Chapetr. Designing Hybrid Nanoparticles. 2014; 1.
- Reed SM, James EH. Green Chemistry in the organic teaching laboratory: an environmentally benign synthesis of adipic acid. Journal of Chemical Education. 2000; 77: 1627.
- Dhand C, Dwivedi N, Loh XJ, Ying ANJ, Verma NK, et al. Methods and strategies for the synthesis of diverse nanoparticles and their applications: a comprehensive overview. RSC Adv. 2015; 5: 105003–105037.
- Rai M, Yadav A, Gade A. Current trends in phytosynthesis of metal nanoparticles. Crit Rev Biotechnol 2008; 28: 277–284.
- Nagaraj G, Brundha D, Kowsalya V, Chandraleka C, Sangavi S, Jayalakshmi R, et al. Biosynthesis of zinc doped *Aloe Vera* for green nanoparticles. Materials Today: Proceedings. 2020; 43: 3354-3358.
- Logeswari P, Silambarasan S, Abraham J. Ecofriendly synthesis of silver nanoparticles from commercially available plant powders and their antibacterial properties. Scientia Iranica. 2013; 20: 1049-1054.
- Gnanasangeetha D, Thambavani S. One pot synthesis of zinc oxide nanoparticles via chemical and green method. Res J Mater Sci. 2013; 1: 1–8.
- Husen A, Siddiqi KS. Phytosynthesis of nanoparticles: concept, controversy and application. Nanoscale Research Letters. 2014; 9: 229 - 229.
- Sharma S, Kumar K. *Aloe vera* leaf extract as a green agent for the synthesis of CuO nanoparticles inactivating bacterial pathogens and dye. J. Dispersion Sci. Technol. 2020; 1–13.
- Bachheti RK, Abate L, Deepti, Bachheti A, Madhusudhan A, Husen A. Algae-, fungi-, and yeast-mediated biological synthesis of nanoparticles and their various biomedical applications. (journal). 2021;:701-734. doi:10.1016/B978-0-12-821938-6.00022-0
- Thongam, DD, Chaturvedi H. Effect of biochemical compounds on ZnO nanomaterial preparation using *Aloe vera* and lemon extracts. Materials Today: Proceedings. 2021; 44: 4299-4304.
- Ravichandran V, Vasanthi S, Shalini S, Shah SAA, Tripathy M, et al. Green synthesis, characterization, antibacterial, antioxidant and photocatalytic



- activity of *Parkia speciosa* leaves extract mediated silver nanoparticles. *Results Phys.* 2029; 15: 102565.
13. Rajabi HR, Naghiha R, Kheirizadeh M, Sadatfaraji H, Mirzaei A, Alvand ZM. Microwave assisted extraction as an efficient approach for biosynthesis of zinc oxide nanoparticles: Synthesis, characterization, and biological properties. *Materials science & engineering. C, Materials for biological applications.* 2017; 78: 1109-1118.
  14. Ramanujam K, Sundrarajan M. Antibacterial effects of biosynthesized MgO nanoparticles using ethanolic fruit extract of *Emblca officinalis*. *Journal of photochemistry and photobiology. B, Biology.* 2014; 141: 296-300.
  15. Kumar J, Kwon Y, Baek K. Green biosynthesis of gold nanoparticles by onion peel extract: synthesis, characterization and biological activities. *Adv. Powder Technol.* 2016; 27: 2204–2213.
  16. Rajeshkumar S, Naik P. Synthesis and biomedical applications of Cerium oxide nanoparticles – A Review. *Biotechnology Reports.* 2018; 17: 1-5.
  17. Vidovix TB, Quesada HB, Januario EFD, Bergamasco R, Vieira AMS. Green synthesis of copper oxide nanoparticles using *Punica granatum* leaf extract applied to the removal of methylene blue. *Mater Lett.* 2019; 257: 126685.
  18. Dobrucka R. Synthesis of Titanium Dioxide Nanoparticles Using *Echinacea purpurea* Herba. *Iranian Journal of Pharmaceutical Research : IJPR.* 2017; 16: 756-762.
  19. Karnan M, Subramani K, Sudhan N, Ilayaraja N, Sathish M. *Aloe vera* Derived Activated High-Surface-Area Carbon for Flexible and High-Energy Supercapacitors. *ACS applied materials & interfaces.* 2016; 8: 35191-35202.
  20. Nasrollahzadeh M, Sajjadi M, Dadashi J, Ghafuri H. Pd-based nanoparticles: Plant-assisted biosynthesis, characterization, mechanism, stability, catalytic and antimicrobial activities. *Advances in colloid and interface science.* 2020; 276: 102103.
  21. Matussin S, Harunsani MH, Tan AL, Khan MM. Plant-extract-mediated SnO<sub>2</sub> nanoparticles: synthesis and applications. *ACS Sustainable Chem Eng.* 2020; 8: 3040-3054.
  22. Mashrai H, Khanam Shamsuzzaman RN, Aljawfi. Biological synthesis of ZnO nanoparticles using *C. albicans* and studying their catalytic performance in the synthesis of steroidal pyrazolines. *Arab J Chem.* 2017; 10: S1530–S1536.
  23. Jeevanandam J, Barhoum A, Chan YS, Dufresne A, Danquah MK. Review on nanoparticles and nanostructured materials: history, sources, toxicity and regulations. *Beilstein Journal of Nanotechnology.* 2018;9:1050-1074. doi:10.3762/bjnano.9.98
  24. Khan I, Saeed K, Khan I. Nanoparticles: properties, applications and toxicities. *Arabian J Chem.* 2019; 12: 908–931.
  25. Selim YA, Azb MA, Ragab I, El-Azim MHMA. Green Synthesis of Zinc Oxide Nanoparticles Using Aqueous Extract of *Deverra tortuosa* and their Cytotoxic Activities. *Scientific Reports.* 2020; 10.
  26. Njagi EC, Huang H, Stafford L, Genuino H, Galindo HM, Collins JB, et al. Biosynthesis of iron and silver nanoparticles at room temperature using aqueous sorghum bran extracts. *Langmuir : the ACS journal of surfaces and colloids.* 2011; 27: 264-271.
  27. Govindaraju K, Basha SK, Kumar VG, Singaravelu G. Silver, gold and bimetallic nanoparticles production using single-cell protein (*Spirulina platensis*) Geitler. *Journal of Materials Science.* 2008; 43: 5115-5122.
  28. Bachheti A, Bachheti RK, Abate L, Husen A. Current status of *Aloe*-based nanoparticle fabrication, characterization and their application in some cutting-edge areas. *South African Journal of Botany.* 2021.
  29. Anju T, Parvathy S, Veetil MV, Rosemary J, Ansalna T, Shahzabanu M, et al. Green synthesis of silver nanoparticles from *Aloe vera* leaf extract and its antimicrobial activity. *Materials Today: Proceedings.* 2021; 43: 3956-3960.
  30. Shankar SS, Rai A, Ahmad A, Sastry M. Rapid synthesis of Au, Ag, and bimetallic Au core-Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. *Journal of colloid and interface science.* 2004; 275: 496-502.
  31. Ahmad N, Sharma S, Alam MK, Singh VN, Shamsi SF, Mehta BR, et al. Rapid synthesis of silver nanoparticles using dried medicinal plant of basil. *Colloids and surfaces. B, Biointerfaces.* 2010; 81: 81-86.
  32. Kasthuri J, Veerapandian S, Rajendiran N. Biological synthesis of silver and gold nanoparticles using apiin as reducing agent. *Colloids and surfaces. B, Biointerfaces.* 2009; 68: 55-60.
  33. Panigrahi S, Kundu S, Ghosh S, Nath S, Pal T. General method of synthesis for metal nanoparticles. *J Nanoparticle Res.* 2004; 6: 411–4.
  34. Zayed MF, Eisa WH, Shabaka AA. Malva parviflora extract assisted green synthesis of silver nanoparticles. *Spectrochimica acta. Part A, Molecular and biomolecular spectroscopy.* 2012;98:423-428. doi:10.1016/j.saa.2012.08.072
  35. Yadeta AT. Food applications of *Aloe* species: A review. *J Plant Sci Phytopathol.* 2022; 6: 024-032.
  36. Salehi B, Albayrak S, Antolak H, Kręgiel D, Pawlikowska E, et al. *Aloe* Genus Plants: From Farm to Food Applications and Phytopharmacotherapy. *Int J Mol Sci.* 2018; 19: 2843.
  37. Sbhathu DB, Berhe GG, Hndeya AG, Abdu A, Mulugeta A, Abraha HB, et al. Hair Washing Formulations from *Aloe elegans* Todaro Gel: The Potential for Making Hair Shampoo. *Advances in Pharmacological and Pharmaceutical Sciences.* 2020; 2020: 1-9.
  38. Grace OM, Kokubun T, Veitch NC, Simmonds MSJ. Characterisation of a nataloin derivative from *Aloe ellenbeckii*, a maculate species from East Africa. *South African Journal of Botany.* 2008; 74: 761–763.
  39. Fahimmunisha BA, Ishwarya R, AlSalhi MS, Devanesan S, Govindarajan M, et al. Green fabrication, characterization and antibacterial potential of zinc oxide nanoparticles using *Aloe socotrina* leaf extract: a novel drug delivery approach. *J Drug Delivery Sci Technol.* 2020; 55: 101465.
  40. BOUDREAU MD, BELAND FA. An Evaluation of the Biological and Toxicological Properties of *Aloe Barbadensis* (Miller), *Aloe Vera*. *Journal of Environmental Science and Health, Part C.* 2006; 24: 103-154.
  41. Nejjatzadeh-Barandozi F. Antibacterial activities and antioxidant capacity of *Aloe vera*. *Organic and Medicinal Chemistry Letters.* 2013; 3: 5.
  42. Boudreau MD, Mellick PW, Olson GR, Felton RP, Thorn BT, Beland FA. Clear evidence of carcinogenic activity by a whole-leaf extract of *Aloe barbadensis miller (Aloe vera)* in F344/N rats. *Toxicological sciences : an official journal of the Society of Toxicology.* 2013; 131: 26-39.
  43. Andrea B, Dumitrița R, Florina C, Francisc D, Anastasia V, Socaci S, et al. Comparative analysis of some bioactive compounds in leaves of different *Aloe* species. *BMC Chemistry.* 2020; 14.
  44. Jha A, Prakash D, Bisht D. A Phytochemical screening of the ethanolic extract of *A.vera* gel. *International Journal of Science and Research.* 2019; 8(10): 1543-1545.
  45. Mudin J, Etana D, Salah H, Dagne A, Milkyas E. Anthraquinones from the roots of *Aloe gilbertii* and *Aloe elegans*. *J Natur Sci Res.* 2018; 8: 1-7.
  46. Ranghoo-Sanmukhiya M, Govinden-Soulange J, Lavergne C, Khoyratty S, Silva DD, Frederich M, et al. Molecular biology, phytochemistry and bioactivity of three endemic *Aloe* species from Mauritius and Réunion Islands. *Phytochemical analysis : PCA.* 2010; 21: 566-574.
  47. Hadera brhane G, Gopalakrishnan VK, Hagos Z, Hiruy M, Devaki K, et al. Phytochemical screening and in vitro antioxidant activities of ethanolic gel extract of *Aloe adigratana* Reynolds. *Journal of Pharmacy Research.* 2018; 12: 13-19.
  48. Gauniyal P, Teotia UVS. Phytochemical screening and antimicrobial activity of some medicinal plants against oral flora. *Asian Pac J Health Sci.* 2014; 1: 255–263.
  49. Wintola OA, Afolayan AJ. Phytochemical constituents and antioxidant activities of the whole leaf extract of *Aloe ferox* Mill. *Pharmacognosy Magazine.* 2011; 7: 325.
  50. Bisi-Johnson MA, Obi CL, Samuel BB, Eloff JN, Okoh AI. Antibacterial activity of crude extracts of some South African medicinal plants against multidrug resistant etiological agents of diarrhoea. *BMC Complementary and*

- Alternative Medicine. 2017; 17.
51. Kumar S, Yadav A, Yadav M, Yadav JP. Effect of climate change on phytochemical diversity, total phenolic content and in vitro antioxidant activity of *Aloe vera* (L.) Burm.f. BMC Research Notes. 2017; 10.
52. Muthii RZ, Mucunu MJ, Peter MM, Gitahi KS. Phytochemistry and toxicity studies of aqueous and methanol extract of naturally growing and cultivated *Aloe turkanensis*. J Pharmacogn Phytochem. 2015; 3: 144–147.
53. Kedarnath N, Surekha RS, Mahantesh S, Patil C. Phytochemical screening and antimicrobial activity of *A. vera*. World Res J Med Aromat Plants. 2012; 1: 11–13.
54. Amare GG, Meharie BG, Belayneh YM. Evaluation of Antidiabetic Activity of the Leaf Latex of *Aloe pulcherrima* Gilbert and Sebsebe (Aloaceae). Evidence-based Complementary and Alternative Medicine : eCAM. 2020; 2020: 1-9.
55. Amoo SO, Aremu AO, Staden JV. Unraveling the medicinal potential of South African *Aloe* species. Journal of ethnopharmacology. 2014; 153: 19-41.
56. Ni Y, Turner D, Yates KM, Tizard I. Isolation and characterization of structural components of *Aloe vera* L. leaf pulp. International immunopharmacology. 2004; 4: 1745-1755.
57. Akaberi M, Sobhani Z, Javadi B, Sahebkar A, Emami SA. Therapeutic effects of *Aloe* spp. in traditional and modern medicine: A review. Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie. 2016; 84: 759-772.
58. Cock IE. The Genus *Aloe*: Phytochemistry and Therapeutic Uses Including Treatments for Gastrointestinal Conditions and Chronic Inflammation. Progress in drug research. Fortschritte der Arzneimittelforschung. Progres des recherches pharmaceutiques. 2015; 70: 179-235.
59. Dagne E, Bisrat D, Viljoen A, Van Wyk BE. Chemistry of *Aloe* species. Curr Org Chem. 2020; 4: 1055–1078.
60. Kahramanoğlu, Chen C, Chen J, Wan C. Chemical Constituents, Antimicrobial Activity, and Food Preservative Characteristics of *Aloe vera* Gel. Agronomy. 2019; 9: 831.
61. Sharrif Moghaddasi M, Sandeep Kumar V. *Aloe vera* their chemicals composition and applications: A review. Int J Biol Med Res. 2011; 2: 466-71.
62. Mahor G, Ali SA. Recent update on the medicinal properties and use of *Aloe vera* in the treatment of various ailments. Biosci Biotechnol Res Commun. 2016; 9: 277-292.
63. Hamman JH. Composition and applications of *A. vera* leaf gel (review). Molecules. 2008; 13: 1599-1616.
64. Cao X, Huang D, Dong Y, Zhao H, Ito Y. Separation of Aloins A and B from *Aloe Vera* Exudates by High Speed Countercurrent Chromatography. Journal of Liquid Chromatography & Related Technologies. 2007; 30: 1657-1668.
65. Megeressa M, Bisrat D, Mazumder A, Asres K. Structural elucidation of some antimicrobial constituents from the leaf latex of *Aloe trigonantha* L.C. Leach. BMC Complementary and Alternative Medicine. 2015; 15.
66. Teka T, Kassahun H. Characterization and Evaluation of Antioxidant Activity of *Aloe schelpei* Reynolds. Drug Design, Development and Therapy. 2020; 14: 1003-1008.
67. Abdissa N, Gohlke S, Frese M, Sewald N. Cytotoxic Compounds from *Aloe megalacantha*. Molecules : A Journal of Synthetic Chemistry and Natural Product Chemistry. 2017; 22: 1136.
68. Martínez-Sánchez A, López-Cañavate ME, Guirao-Martínez J, Roca MJ, Aguayo E. *Aloe vera* Flowers, a Byproduct with Great Potential and Wide Application, Depending on Maturity Stage. Foods. 2020; 9: 1542.
69. Perumal P, Sivaraj P, Abhilash K, Soundarya G, Balraju P, Selvin PC. Green synthesized spinel lithium titanate nano anode material using *Aloe Vera* extract for potential application to lithium ion batteries. Journal of Science: Advanced Materials and Devices. 2020; 5: 346-353.
70. Sai Priya G, Kanneganti A, Anil Kumar K, Venkateswara Rao K, Bykkam S. Biosynthesis of cerium oxide nanoparticles using *A. barbadensis* Miller gel. International Journal of Scientific and Research Publications. 2014; 4: 1-4.
71. Chaudhary A, Kumar N, Kumar R, Salar RK. Antimicrobial activity of zinc oxide nanoparticles synthesized from *A. vera* peel extract. SN Appl Sci. 2019; 1: 136.
72. Klinkaewnarong J, Swatsitang E, Masingboon C, Seraphin S, Maensiri S. Synthesis and characterization of nanocrystalline HAp powders prepared by using *Aloe vera* plant extracted solution. Current Applied Physics. 2010; 10: 521-525.
73. Karimi J, Mohsenzadeh S. Rapid, Green, and Eco-Friendly Biosynthesis of Copper Nanoparticles Using Flower Extract of *Aloe Vera*. Synthesis and Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry. 2015; 45: 895-898.
74. Medda S, Hajra A, Dey U, Bose P, Mondal NK. Biosynthesis of silver nanoparticles from *Aloe vera* leaf extract and antifungal activity against *Rhizopus* sp. and *Aspergillus* sp. Applied Nanoscience. 2014; 5: 875-880.
75. Ali K, Dwivedi S, Azam A, Saquib Q, Al-Said MS, Alkhedairy AA, et al. *Aloe vera* extract functionalized zinc oxide nanoparticles as nanoantibiotics against multi-drug resistant clinical bacterial isolates. Journal of colloid and interface science. 2016; 472: 145-156.
76. Nithya A, Rakesh K, Jothivenkatachalam K. Biosynthesis, characterization and application of titanium dioxide nanoparticles. Nano Vision. 2013; 3: 169–174.
77. Albeladi SSR, Malik MA, Al-thabaiti SA. Facile biofabrication of silver nanoparticles using *Salvia officinalis* leaf extract and its catalytic activity towards Congo red dye degradation. Journal of materials research and technology. 2020; 9: 10031-10044.
78. Okafor F, Janen A, Kukhtareva T, Edwards V, Curley M. Green Synthesis of Silver Nanoparticles, Their Characterization, Application and Antibacterial Activity †. International Journal of Environmental Research and Public Health. 2013; 10: 5221-5238.
79. Vilchis-Nestor AR, Sanchez-Mendieta V, Camacho-López MA, Gómez-Espinosa RM, Camacho-López MA, et al. Solventless synthesis and optical properties of Au and Ag nanoparticles using *Camellia sinensis* extract. Materials Letters. 2008; 62: 3103-3105.
80. Amin M, Anwar F, Janjua MRSa, Iqbal MA, Rashid U. Green Synthesis of Silver Nanoparticles through Reduction with *Solanum xanthocarpum* L. Berry Extract: Characterization, Antimicrobial and Urease Inhibitory Activities against *Helicobacter pylori*. International Journal of Molecular Sciences. 2012; 13: 9923-9941.
81. Kanchana A, Devarajan S, Rathakrishnan Ayyappan S. Green synthesis and characterization of palladium nanoparticles and its conjugates from *Solanum trilobatum* leaf extract, NanoMicro Letters. 2010; 2: 169-176.
82. Villaverde-Cantizano G, Laurenti M, Rubio-Retama J, Contreras-Cáceres R. CHAPTER 1. Reducing Agents in Colloidal Nanoparticle Synthesis – an Introduction. Royal Society of Chemistry. 2021; 1-27.
83. Zhang Y, Yang D, Kong Y, Wang X, Pandoli O, Gao G. Synergetic Antibacterial Effects of Silver Nanoparticles@*Aloe Vera* Prepared via a Green Method. Nano Biomedicine and Engineering. 2010; 2: 252-257.
84. Gulati S, Sachdeva M, Bhasin KK. Capping agents in nanoparticle synthesis: Surfactant and solvent system. In AIP Conference Proceedings. 2018; 1953: 030214.
85. Aisida SO, Alnasir MH, Botha S, Bashir AKH, Bucher R, Ahmad I, et al. The role of polyethylene glycol on the microstructural, magnetic and specific absorption rate in thermoablation properties of Mn-Zn ferrite nanoparticles by sol-gel protocol. Eur Polymer J. 2020; 132: 109739.
86. Javed R, Zia M, Naz S, Aisida SO, Ain NU, Ao Q. Role of capping agents in the application of nanoparticles in biomedicine and environmental remediation: recent trends and future prospects. Journal of Nanobiotechnology. 2020; 18.
87. Ajitha B, Ashok Kumar Reddy Y, Sreedhara Reddy P, Jeon Hwan-Jin, Won Ahn Chi. Role of capping agents in controlling silver nanoparticles size, antibacterial activity and potential application as optical hydrogen peroxide

- sensor. RSC Adv. 2016; 6: 36171-36179.
88. Fahimmunisha BA, Ishwarya R, Al Salhi MS, Devanesan S, Govindarajan M, Vaseeharan B. Green fabrication, characterization and antibacterial potential of zinc oxide nanoparticles using *Aloe socotrina* leaf extract: A novel drug delivery approach. Journal of Drug Delivery Science and Technology. 2020; 55: 101465.
89. Arshad H, Saleem M, Pasha U, Sadaf S. Synthesis of *A. vera*-conjugated silver nanoparticles for use against multidrug-resistant microorganisms, Electronic Journal of Biotechnology. 2022; 55: 55–64.
90. Ali K, Saquib Q, Siddiqui MA, Ahmad J, Al-Khedhairi AA, Musarrat J. Anti-cancer efficacy of *Aloe vera* capped hematite nanoparticles in human breast cancer (MCF-7) cells. Journal of Drug Delivery Science and Technology. 2020; 60: 102052.
91. Sangeetha G, Rajeshwari S, Venckatesh R. Green synthesis of zinc oxide nanoparticles by *Aloe barbadensis miller* leaf extract: Structure and optical properties. Materials Research Bulletin. 2011; 46: 2560-2566.
92. Amini SM. Preparation of antimicrobial metallic nanoparticles with bioactive compounds. Materials science & engineering. C, Materials for biological applications. 2019; 103: 109809.
93. Zewde B, Ambaye A, Stubbs III J, Raghavan D. A Review of stabilized silver nanoparticles – synthesis, biological properties, characterization, and potential areas of applications. JSM Nanotechnol Nanomed. 2016; 4: 1043.
94. Jha AK, Orasad K, Prasad K, Kulakarni AR. Plant system: natures nanofactory, Colloids Surf. B: Biointerfaces. 2009; 73: 219–223.
95. Gunalan S, Sivaraj R, Venckatesh R. *Aloe barbadensis* Miller mediated green synthesis of mono-disperse copper oxide nanoparticles: optical properties. Spectrochimica acta. Part A, Molecular and biomolecular spectroscopy. 2012; 97: 1140-1144.
96. Juibari NM, Eslami A. Synthesis of nickel oxide nanorods by *Aloe vera* leaf extract. Journal of Thermal Analysis and Calorimetry. 2018; 136: 913-923.
97. Ovais M, Khalil AT, Islam NU, Ahmad I, Ayaz M, Saravanan M, et al. Role of plant phytochemicals and microbial enzymes in biosynthesis of metallic nanoparticles. Applied Microbiology and Biotechnology. 2018; 102: 6799-6814.
98. Rao PV, Nallappan D, Madhavi K, Rahman S, Wei LJ, Gan SH. Phytochemicals and Biogenic Metallic Nanoparticles as Anticancer Agents. Oxidative Medicine and Cellular Longevity. 2016; 2016: 1-15.
99. Durán N, Durán M, Jesus MBD, Seabra AB, Fávoro WJ, Nakazato G. Silver nanoparticles: A new view on mechanistic aspects on antimicrobial activity. Nanomedicine : nanotechnology, biology, and medicine. 2016; 12: 789-799.
100. Yin J, Xu L. Batch preparation of electrospun polycaprolactone/chitosan/*Aloe vera* blended nanofiber membranes for novel wound dressing. International journal of biological macromolecules. 2020; 160: 352-363.
101. Gao S, Yang S, Shu J, Zhang S, Li Z, Jiang K. Green fabrication of hierarchical CuO hollow micro/nanostructures and enhanced performance as electrode materials for lithium-ion batteries. The Journal of Physical Chemistry C. 2008; 112: 19324-19328.
102. Cao Y, Xie Y, Liu L, Xiao A, Li Y, et al. Influence of phytochemicals on the biocompatibility of inorganic nanoparticles: a state-of-the-art review. Phytochemistry Reviews. 2017; 16: 555-63.
103. Chakraborty T, Gupta S, Nair A, Chauhan S, Saini V. Wound healing potential of insulin-loaded nanoemulsion with *Aloe vera* gel in diabetic rats. Journal of Drug Delivery Science and Technology. 2021; 64: 102601.
104. Sohal JK, Saraf A, Shukla K, Shrivastava M. Determination of antioxidant potential of biochemically synthesized silver nanoparticles using *Aloe vera* gel extract. Plant Science Today. 2019; 6: 208-217.
105. Vyas J, Rana S. Antioxidant Activity and Biogenic Synthesis of Selenium Nanoparticles Using the Leaf Extract of *Aloe Vera*. International Journal of Current Pharmaceutical Research. 2017; 9: 147.
106. Mahendiran D, Subash G, Selvan DA, Rehana D, Kumar RS, Rahiman AK. Biosynthesis of Zinc Oxide Nanoparticles Using Plant Extracts of *Aloe vera* and *Hibiscus sabdariffa*: Phytochemical, Antibacterial, Antioxidant and Anti-proliferative Studies. Bio Nano Science. 2017; 7: 530-545.
107. Dinesh D, Murugan K, Madhiyazhagan P, Panneerselvam C, Kumar PM, Nicoletti M, et al. Mosquitocidal and antibacterial activity of green-synthesized silver nanoparticles from *Aloe vera* extracts: towards an effective tool against the malaria vector *Anopheles stephensi*?. Parasitology Research. 2015; 114: 1519-1529.
108. Zhou Q, Lv J, Ren Y, Chen J, Gao D, Lu Z, et al. A green in situ synthesis of silver nanoparticles on cotton fabrics using *Aloe vera* leaf extraction for durable ultraviolet protection and antibacterial activity. Textile Research Journal. 2017; 87: 2407-2419.
109. SP KN, K V. Green Synthesis of Silver and Gold Nanoparticles using *Aloe Vera* Gel and Determining its Antimicrobial Properties on Nanoparticle Impregnated Cotton Fabric. Journal of Nanotechnology Research. 2020; 02: 42-50.
110. Ali K, Dwivedi S, Azam A, Saquib Q, Al-Said MS, Alkhedhairi AA, et al. *Aloe vera* extract functionalized zinc oxide nanoparticles as nanoantibiotics against multi-drug resistant clinical bacterial isolates. Journal of colloid and interface science. 2016; 472: 145-156.
111. Kumar PPNV, Shameem U, Kollu P, Kalyani RL, Pammi SVN. Green Synthesis of Copper Oxide Nanoparticles Using *Aloe vera* Leaf Extract and Its Antibacterial Activity Against Fish Bacterial Pathogens. BioNanoScience. 2015; 5: 135-139.
112. Fardsadegh B, Jafarizadeh-Malmiri, H. *Aloe vera* leaf extract mediated green synthesis of selenium nanoparticles and assessment of their in vitro antimicrobial activity against spoilage fungi and pathogenic bacteria strains. Green Process. Synth. 2019; 8: 399–407.
113. O. S, J. S. *Aloe Vera* Mediated Green Synthesis of ZnO Nanostructure under Sol-gel Method: Effect of Antimicrobial Activity. Journal of Nano-and electronic Physics. 2020; 12: 02041-1-02041-5.
114. Logaranjan K, Raiza AJ, Gopinath SC, Chen Y, Pandian K. Shape-and sizecontrolled synthesis of silver nanoparticles using *A. vera* plant extract and their antimicrobial activity. Nanoscale Res. Lett. 2016; 11: 1–9.
115. Yousefpoor Y, Bolouri B, Bayati M, Shakeri A, Torbaghan YE. The combined effects of *Aloe vera* gel and silver nanoparticles on wound healing in rats. Nanomedicine Journal. 2016; 3: 57-64.
116. Rajkumari J, Magdalane CM, Siddhardha B, Madhavan J, Ramalingam G, Al-Dhabi NA, et al. Synthesis of titanium oxide nanoparticles using *Aloe barbadensis* mill and evaluation of its antibiofilm potential against *Pseudomonas aeruginosa* PAO1. Journal of photochemistry and photobiology. B, Biology. 2019; 201: 111667.
117. Reyhaneh Rahmani, Mohsen Gharanfoli, Mehran Gholamin, Majid Darroudi, Jamshidkha Chamani, et al. Plant-mediated synthesis of superparamagnetic iron oxide nanoparticles (SPIONs) using *Aloe vera* and flaxseed extracts and evaluation of their cellular toxicities. Ceramics International. 2020; 46: 3051-3058.
118. Gupta S, Tejavath KK. Catalytic Reduction of Organic Dyes with Green Synthesized Silver Nanoparticles using *Aloe vera* Leaf Extract. Journal of Nanoscience Nanoengineering and Applications. 2019: 9-21.