

Review Article

Management of Acute Ischemic Hypoxic Encephalopathy in Newborns with Hyperbaric Oxygen: A Review

Sánchez EC*

Department of Hyperbaric Medicine, Universidad Nacional Autónoma de México, Mexico

*Corresponding author: E. Cuauhtémoc Sánchez, Department of Hyperbaric Medicine, Hospital Agustín O´Horan and Centro de Especialidades Médicas del Sureste, Universidad Nacional Autónoma de México, Calle 38 # 101c x 27 y 29 Col. Buenavista, Mérida 97127, Yucatán, Mexico

Received: September 20, 2016; Accepted: November 21, 2016; Published: November 23, 2016

Abstract

Objectives: Describe the use of HBO₂ in the management of acute ischemic hypoxic encephalopathy.

Materials and Methods: The therapeutic mechanisms of hyperbaric oxygen in acute ischemic hypoxic encephalopathy are outlined, based on information obtained from peer-reviewed medical literature.

Results: HBO₂ promotes survival of marginal tissue (penumbra), reduces edema, improves microcirculation, breaks the edema-hypoxia-edema vicious cycle, enhances healing, promotes growth factors up-regulation and improves neovascularization. At cellular level, it maintains the levels of ATP, restores mitochondrial dysfunction, reduces oxidative stress and apoptosis, and promotes anti-oxidant effects. HBO₂ did not show significant differences in fatality rates at six months (RR 0.97, 95%CI 0.34 to 2.75, p=0.96). It showed improvement in the disability and functional performance scales. Trouillas Disability Scale was lower with HBO₂ (MD 2.2 point reduction with 95%CI 0.15 to 4.3, p=0.04), and the mean Orgogozo Scale was higher (MD 27.9 points, 95%CI 4.0 to 51.8, p=0.02).

Conclusion: When used promptly, HBO₂ can modify cerebral inflammation, ischemia, hypoxia, and ischemia-reperfusion injury. It helps preserve marginal tissue and recover the ischemic and metabolic penumbra. A prospective, randomized, and controlled study within the window of opportunity (< 5h) in a stroke center is needed, to determine the real use of HBO₂ in these cases.

Keywords: Acute ischemic hypoxic encephalopathy, Hyperbaric oxygenation, Ischemia-reperfusion-injury, Mitochondrial dysfunction

Introduction

Acute ischemic-hypoxic encephalopathy (stroke) is the most important preventable cause of disability for Americans and the second cause of death worldwide. Every year, more than 795,000 people in the US have a stroke and about 610,000 of these, are first or new strokes. About 25% of strokes (185,000) happen in people who had a previous stroke. The prevalence of stroke (2009-2012) for people aged 20-39, was 0.2 for men and 0.7 for women; for ages 40-59, 1.9 for men and 2.2 for women; for ages 60-79, 6.1 for men and 5.2 for women; and for ages 80+, 15.8 for men and 14.0 for women. About 87% of all strokes are ischemic strokes [1-3].

The risk of having a stroke varies with race and ethnicity. The risk of having a first stroke is nearly twice as high for African Americans compared to whites, and African Americans are more likely to die following a stroke than are whites. Hispanics' risk of stroke falls between that of whites and African Americans. American Indians, Alaska Natives, and African Americans are more likely to have had a stroke than all other groups [1-3].

The incidence of mortality and morbidity due to neonatal acute ischemic-hypoxic encephalopathy (AIHE) has not changed substantially in the last 40 years. It is estimated that close to 25% of the neonatal deaths and 8% of all deaths at 5 years of age throughout the world, annually, are associated with signs of asphyxia at birth

[4,5]. Death or moderate to severe disability can occur in 50-60% of infants diagnosed as having moderate to severe AIHE [6,7].

The average length of stay in non-federal short-stay hospitals was 4.8 days, although in some hospitals; it extended to up to 10.9 days [3]. The annual cost of stroke in the US is estimated in \$34 billion. It includes the cost of health care services, medications and workdays lost [1-3]. Stroke has high direct costs and very high indirect costs. It is estimated that indirect costs account for 80% of the total cost. According to the World Health Organization (WHO) stroke will become the fourth cause of the Global Burden of Disease (GBD) in 2030 [8,9].

In some clinical studies, the 5-year survival rate is 50%. Specialized stroke units have reduced morbidity and mortality (OR 2.2, 95%CI 1.6-2.8, p<0.001), but less than 20% of all stroke cases arrive at the units within the window of treatment (3 to 5 hours) [10]. The phrase "time is brain", emphasizes that human tissue is rapidly lost as stroke progresses and emergent evaluation and therapy are required [11].

AIHE is such a devastating pathology that any gain, no matter how small, can make a big difference in these children's and their families' quality of life (QoL). Prompt treatment should restore adequate perfusion and correct metabolic or cellular alterations. Maintaining adequate perfusion and the cellular metabolic needs may be the cornerstone to reduce CNS damage and promote early

neuro protection [3,12,13]. The extent of the damage depends on the duration, extension, localization, comorbidities and metabolic changes of the lesion [3,14].

Pathophysiology of AHIE (Ischemia-Reperfusion Injury)

When there is an interruption of the cerebral blood flow or oxygen supply to the Central Nervous System (CNS), several changes occur depending on the degree of hypoxia; these could be reversible or irreversible [15]. Reversibility depends on the mitochondrial ability to maintain ATP production. Once it stops, there is a dysfunction of the ion pumps (Na-K and K-Ca) that eventually will create cytotoxic edema. When the mitochondrial dysfunction is severe, calcium is released into the cytoplasm and becomes the first inflammatory mediator [16,17].

The main player in the pathophysiology of acute ischemic-hypoxic encephalopathy (AIHE) is the ischemia-reperfusion injury (IRI) to the Central Nervous System (CNS). The injury is the result of a series of ischemic and metabolic events that present shortly after the interruption of flow, oxygen, and nutrients to the affected regions of the brain [3].

The center of the lesion is the area of necrosis surrounded by the area of ischemia, hypoxia, and edema, i.e. the recoverable area of the brain (penumbra). The area of penumbra constitutes close to 80% of the damaged brain in IRI. If the ischemic and metabolic penumbras are not resolved in a timely and effective fashion, the damage will extend and deepen due to IRI and apoptosis [3].

Mitochondria in AIHE: There are seven stages of cellular shock in (AIHE). The first four stages are reversible. The reversibility depends on cells' capability to maintain adenosine tri-phosphate (ATP) production by the mitochondria [8]. Once ATP is reduced beyond a critical level (< 1 mol/kg), there is an energy crisis within the cell [9]. During the CNS energy crisis, hypoxia upsets the passage of protons (H⁺) across the mitochondrial complexes, reducing production of ATP at the ATPase-synthase level. This produces oxidative stress and the increase of reactive oxygen species (ROS) and its production inside the mitochondria; also, the promotion of neural (nNOS) and inducible nitric oxide synthase (iNOS) production [18,19]. If the oxidative stress is not controlled early, it will progress to oxidative damage. This generates an increase of the intrinsic apoptotic pathway. The energy crisis also promotes glutamate production, the most important excitatory transmitter in the brain [20-22].

With mitochondrial dysfunction, there is a loss of calcium-potassium and sodium-potassium cellular pumps, which creates ionic misbalance and cytotoxic edema. The release of cytochrome C occurs immediately before total mitochondrial dysfunction, due to the rise of nitric oxide. The opening of the mitochondrial transition pore follows. Once the pore is open, it releases its calcium content into the cytosol, increasing cellular edema and affecting cellular homeostasis [3,23-26].

Inflammation in AIHE: Calcium is the first modulator of inflammatory cascades [27,28]. It stimulates the activation of calcium protease, which enhances xanthine dehydrogenase conversion to xanthine oxidase, promoting the production of reactive oxygen

species (ROS) [29]. It also stimulates the arachidonic acid cascade with the subsequent elevation in the levels of cyclooxygenase (COX), lipoxygenase, leukotriene, thromboxane and prostaglandins [30]. It also promotes the expression of the most important transcription factor for inflammation, nuclear transition factor kappa b (NF κ b) and enhances the production of endothelins and cytokines (IL-1, IL-6, IL-8, TNF- α) [31-34]. There is also an elevation of interferon- γ , glutamate, Caspases (3,8, and 9), hypoxia induced factor-1 α (HIF-1 α), and nitric oxide from inducible nitric oxide synthase (iNOS) [35-38].

At endothelial level, there is an increased expression of adhesion molecules (selectins, vascular adhesion molecule, intercellular adhesion molecule) and leukocyte (integrin beta 2). In the late phase of IRI, the adhesion molecules are the predominant mediators of damage, promoting leukocytes production of ROS, perivenular arteriolar vasoconstriction, apoptosis, and creating the no-flow state [39-42].

Few of the early neuro protectors tested have made a real difference. Stenting, angioplasty, hypothermia, and thrombolysis have shown benefits, especially when applied in the first 3 to 5 hours of the onset of AHIE. Only the Food and Drug Administration (FDA) has accepted the plasminogen activator (rtPA) [43-48]. Nevertheless, none of these have proven to resolve the metabolic penumbra. Medication has not shown the same neuroprotective effects of other therapies [49-51].

Growing evidence suggests that a spectrum of epigenetic processes play an important role in the pathophysiology of cerebral ischemia. DNA methylation, histone deacetylase, histone methylation and micro RNAs (miRNAs) regulate vascular and neuronal regeneration after cerebral ischemia. MiRNAs are supposed to be potential biomarkers for stroke and other related pathologies. Epigenetic strategies for ischemic stroke treatments may modulate neural cell regeneration and promote brain repair and functional recovery [52,53].

Hyperbaric Oxygen in Acute Hypoxic Ischemic Encephalopathies

Hyperbaric oxygen (HBO₂) is a treatment option in which a patient breathes 100% oxygen while inside a treatment chamber at a pressure higher than 1.4 atmospheres absolute (atm abs). The treatment chamber could be mono- or multi place [3,54].

HBO₂ is based on gas laws. Increasing the pressure inside the chamber will promote the amount of oxygen that will dissolve in all body fluids, especially in plasma. The normal treatment pressure varies from 1.5 to 3.0 atm abs and lasts from 45 to 120 minutes, except for diving accident management that could take longer. The primary effect of breathing oxygen at pressure is hyper oxygenation. Partial pressures in plasma at 2.0 atm abs are close to 1500 millimeters of mercury (mmHg) and at 3.0 atm abs are around 2200 mm Hg [3,54].

HBO₂ produces several secondary effects. Hyper oxygenation increases the endovascular oxygen partial pressure. Restoring tissue oxygen tension to normal or supernormal levels, will break the edema-hypoxia-edema vicious cycle. The vasoconstriction effect of HBO₂, without affecting tissue oxygenation due to the increased partial pressure of oxygen, reduces tissue edema and

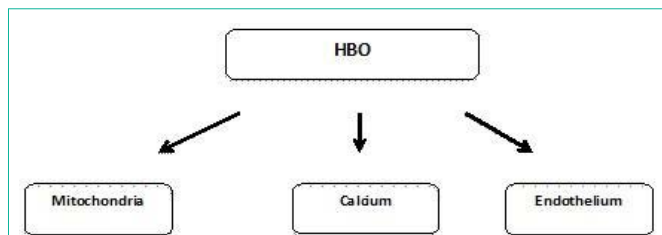


Figure 1: Effects of hyperbaric oxygen on IHE (part 1).

Restores O₂ levels Chelates calcium from cytosol <Expression of Maintains ATP levels Stabilizes Na/K ATPase ICAM-1 <Intrinsic ROS production Stabilizes Ca-dependent enzymes Integrin β2> Bcl-2 < IRI < Damage <Bax <Production of glutamate <Endothelins <Caspases 3 and 9 <Cytotoxic edema <Aquaporin < Apoptosis damage.

improves microcirculation [55,56]. HBO₂ also restores damage to the blood brain barrier (BBB) [57,58] and modulates the aquaporin response in the choroid plexus, neurons; and astrocytes; aquaporin 4 in astrocytes; and aquaporin 9 in astrocytes and catecholaminergic neurons [59,60].

HBO₂ promotes wound healing, angiogenesis, lymphogenesis, and increases growth factor production [3,12,54,55,61-67]. HBO₂ has direct and indirect antimicrobial effects [12,68,69]. It promotes bone remodeling [70-72] and has rheological effects that are synergistic with pentoxifylline [73]. It also reduces hyper coagulation induced by zymosan and in multiple organ failure [74,75]. Finally HBO₂ increases the mobilization of stem cells from bone marrow and has been used for traumatic brain injury and spinal cord injury [76-78].

By reducing cerebral loss of energy, hyperbaric oxygenation restores cellular ion homeostasis, reduces acidosis, and stabilizes cellular calcium. It also limits excitatory mediators, ROS toxicity, apoptosis, and ischemia reperfusion injury. This is accomplished through several mechanisms. First, it increases oxygen tension and restores oxygen content at tissue and cellular level. If it is applied in a timely fashion, it salvages the cerebral ischemic and metabolic penumbrae, and restores mitochondrial oxidative phosphorylation (Figure 1) [3,12,79,80].

The restoration of cellular oxygen tension and mitochondrial function reduces the expression of NFκB, phospholipase A₂, release of calcium, production of COX-2 miRNA, cytokines (IL-1, IL-6, IL-8, and TNFα), iNOS, and nNOS. Paradoxically, it increases the level of IL-10, an anti-inflammatory cytokine, also mediated by NFκB [3,12,81-85].

Hyperbaric oxygen restores the flux of protons (H⁺) in the mitochondrial complexes, thereby restoring the production of ATP. This limits mitochondrial DNA (mtDNA) damage, Caspases expression, oxidative damage (by ROS), the expression of the Nogo-A, Ng-R, and RhoA transcription factors; and it has an anti-apoptotic effect by increasing BCL-2, heme oxygenase-1 (HO-1), and heat shock proteins (HSP-70 and 72) [86-89].

Hyperbaric oxygen reduces the expression of intercellular adhesion molecule-1 (ICAM-1) and endothelin-1, at the endothelium, and neutrophils β2 integrin; with the concomitant reduction of the late phase of ischemic reperfusion injury (IRI) [90-93]. In addition to HBO₂ protective effects through the restoration and maintenance of oxygenation, it also provides antioxidant protection by inducing

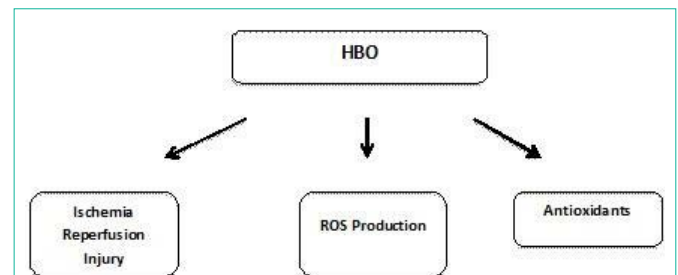


Figure 2: Effects of hyperbaric oxygen on IHE (part 2).

< XO expression (< ROS) <Mitochondrial production > SOD <nNOS and iNOS <XO production > Catalase < Phospholipase A2 <Phospholipase production > GSH-P Lipoxygenase <nNOS and iNOS production > HO-1 Cyclooxygenase > Nrf-2 Modifies NFKb expression > Glutathione < IL-1, IL-6, IL-8> HSP < TNF-α, IFN-γ > IL-10 Modifies Nogo-A/ Rho-A > HSP Modifies miRNA Modifies MMP Modifies HIF-1α.

selective gene and chaperone production [94-96]. The antioxidant effects are also provided by the increased enzymatic (superoxide dismutase, glutathione peroxidase, Nrf2 and catalase) and non-enzymatic (glutathione and cysteine) antioxidant protection (Figure 2) [97-99].

Hyperbaric oxygen has been used for acute hypoxic ischemia encephalopathies (cerebral ischemia and stroke) for more than forty years. Despite the quality and number of articles published, they have not been able to produce enough sound evidence to place HBO₂ as an accepted adjunct treatment option for acute stroke [3].

There are reports involving more than 2000 cases and series of cases. A 2014 Cochrane review by Bennett, et al included eleven randomized, controlled studies involving 705 participants. No significant differences were noted in the case fatality rate at six months in the HBO₂ group compared to the control group (RR 0.97, 95%CI 0.34 to 2.75, p=0.96). Four of 14 scale measures of disability and functional performance indicated improvement following HBO₂; the Trouillas Disability Scale was lower with HBO₂ (MD 2.2 point reduction with 95%CI 0.15 to 4.3, p=0.04), and the mean Orgogozo Scale was higher (MD 27.9 points, 95% CI 4.0 to 51.8, p=0.02). They concluded that further research is required to better define the role of HBO₂ in acute stroke [100].

Use of Hyperbaric Oxygenation (HBO₂) in AHIE in Neonates

HBO₂ use in neonates was almost completely discontinued after Hutchinson's and the USSR experiences. Presently, the largest experience was published by China. The justification for the use of HBO₂ in neonates was the minor modification in morbidity and mortality rates in the last four decades, and the increasing published experience of its possible effects as an early neuro protector. The major controversy was the possible side effects of HBO₂, especially neonatal retinopathy [101].

It is recommended that neonates treated with HBO₂ should be older than 34.5 weeks of gestation, and weight above 1.2 kg. Younger neonates with a lower weight, have a higher possibility of developing complications due to prematurity itself. The most important one would be a pulmonary complication, and retinopathy [102,103]. In the USSR they only treated term neonates reducing the possibility of HBO₂ side effects related to prematurity.

Normally, AHIE is more frequent in neonates with Apgar scores below 3 at one minute and below 5 at five minutes of life, when the resuscitation efforts last longer than 8 minutes, and when the pH is lower than 7.2. Neonates presenting such conditions will require management in a neonatal intensive care unit (NICU). Normally they will present cerebral edema at 4 hours and seizures at 6 hours, thus aggravating their already critical cerebral condition [104-106].

The evaluation of neonates with HIE should include: electroencephalogram (EEG), visual and auditory evoked potentials (EP), transfontanelar Doppler imaging, fundoscopic evaluation, and laboratory tests. The test should be repeated 4 hours after the first HBO₂ treatment and after 24 hours. The HBO₂ treatment should start as early as possible, the sooner the better. It should be within the first 6 hours to guarantee better results. Nevertheless, if there is a good coordination between the neonatologist and the hyperbaric oxygen department, the treatment can start once the patient has been stabilized (first hour). The standard of care inside the hyperbaric chamber requires an inside attendant (neonatologist) and should be the same as in the NICU [101].

There are several special considerations for the treatment of a neonate patient inside a hyperbaric chamber. One of the main problems Hutchinson encountered was the lack of appropriate neonatal medical equipment for hyperbaric use. This has not changed in the last 40 years. There is no neonatal hyperbaric ventilator, although there is a manufacturer that has a hyperbaric ventilator and a neonatal one, but does not see a market for a neonatal hyperbaric ventilator. Also, there are no hyperbaric IV pumps available that can deliver the reduced flows needed by neonatal patients (½ ml per hour) [101,107,108].

To cope with these technical deficiencies, a trained neonatologist should go inside the hyperbaric chamber and ventilate the patient with an Ambu bag. To deal with the low IV volumes required we had to turn the IV pump on and off to be able to deliver the required IV flow. The patient was monitored with ECG and with a transcutaneous oxygen monitor (TCOM). TCOM is used routinely for our ICU patients treated inside the hyperbaric chamber, because it is very sensitive for changes in patients' ventilation. A Bispectral index monitor (BIS) can be used to monitor frontal EEG during HBO₂ treatment; it also helps in evaluating patients' sedation and can be a useful tool to detect early abnormal EEG activity related to oxygen toxicity to the CNS [101].

A transfer protocol in coordination with the NICU and the Hyperbaric Unit should be developed to refer the patient as soon as possible (<4 h). The treatment profile is 1.8-2.0 atmospheres absolute for 45 minutes (20 minutes oxygen, 5 minutes air brake, 20 minutes oxygen), QD or BID. The air brake was included to reduce the pulmonary oxygen toxicity of HBO₂. To avoid hypothermia, the chamber linen was warmed to 40°C in water vapor. The recommended pressurizing and depressurizing rate was 1 psi per minute. Myringotomies should be performed before starting the treatment. The patient should be monitored during the entire procedure. Normally, the patients will be ventilated and dependent on cardiotoxic medication. The range of delay to treatment after resuscitation should be less than 4 hours. [101].

Acute hypoxic ischemic encephalopathy in neonates is a devastating lesion, its morbidity and mortality has not been modified significantly in the last forty years [109-111]. Any beneficial effects in salvaged tissue will make a big difference in the quality of life of patients, their families and society. HBO₂ has shown beneficial effects on IRI, including the CNS [112-118]. It could be an effective treatment for neonatal AHIE

HBO has been used in neonatal rats for hypoxic ischemic brain damage (HIBD) [119,120], and for necrotizing enterocolitis (NEC) [121]. In the model of unilateral carotid artery ligation followed by a 2.5 hours hypoxia (8% O₂ at 37°C) in 7 days old rat pups, Calvert, et al. showed a reduction in atrophy and apoptosis in the HBO treated animals. The sensorimotor function was also improved by HBO (p<0.05) [119]. Wang et al., observed that HBO promoted cell proliferation of neural stem cells, improving some of the neurologic performances, and alleviating the brain damage (p<0.05). The therapeutic window for effective HBO treatment could be delayed up to 12 h after HBO, while the effect decreased 24 h after HIBD [120]. Guven, et al. reported that HBO had an ameliorating effect on oxidative stress, nitrosamine stress, and antioxidant enzymes activities in the intestine of pups subjected to NEC [121].

Hyperbaric Oxygen Side Effects

Pediatric patients are no different than adults, nevertheless neonates are. The fear of oxygen toxicity in neonates has hindered its use in them. In general, term neonates have a good anti-oxidant defense; but premature babies and with low weight, may be at risk due to their immature antioxidant defense system. Although neonatologists are very concerned about premature retinopathy, several publications refer to it as an IRI (it occurs after long exposures to oxygen at FiO₂ of 0.45 or higher, and only after oxygen is discontinued). HBO₂ seems to protect them, instead of causing side effects [101,122-125].

Special care should be taken during treatment to avoid hypothermia and oxygen toxicity, especially pulmonary, in neonates with risk factors for bronchopulmonary dysplasia and/or hyaline membrane. Pulmonary surfactant should be readily available in the event the patient presents pulmonary oxygen toxicity. CNS oxygen toxicity is also a possibility that should be taken into account when treating neonates. The use of bi spectral index monitoring (BIS) could show early signs of it and could be promptly and adequately treated. No other types of hyperbaric oxygen toxicity have been reported in neonates so far [101].

Hypothermia

Lately there have been several reports on hypothermia and its beneficial effects [112]. In a meta-analysis on cooling newborns with hypoxic ischaemic encephalopathy published in 2013, it was reported that in 11 randomized controlled trials comprising 1505 term and late preterm infants with moderate/severe encephalopathy and evidence of intrapartum asphyxia; therapeutic hypothermia resulted in a statistically significant and clinical reduction in the combined outcome of mortality or major neurodevelopmental disability to 18 months of age (RR 0.75 [95% CI 0.68 to 0.83, NNT - 7 (95% CI 5 to 10)]. Cooling also resulted in statically significant reductions in mortality (RR 0.75 [95% CI 0.64 to 0.88, NNT - 11 (95% CI 5 to 14)].

Some adverse effects of hypothermia included and increase sinus bradycardia, increased blood pressure, increase oxygen requirements, and a significant increase in thrombocytopenia. They were transient and reversible after rewarming [126].

Hypothermia may modify apoptosis, reduces cerebral metabolic rate, attenuate the release of excitatory amino acids (glutamate, dopamine), ameliorating the ischemia-impaired uptake of glutamate and lowering production of nitric oxide and free radicals. Some aspects of cooling therapy that remain controversial include: adequate selection of infants, how soon after the insult or birth does cooling need to be started, what level of hypothermia is required, what method (selective head cooling versus whole body cooling) should be used, and what is the duration of cooling required. Time is another critical factor in instituting hypothermia therapy for the newborns. The referral to experienced centers might take away the therapeutic window (< 6 h).

It appears that HBO and hypothermia might have a synergistic effect if they were combined. There is only one report that deals with both therapies used for a patient with hydrogen sulfide intoxication [127-130]. HBO was used before cooling, not simultaneously. Further research is needed in this area to determine the beneficial effects when both therapies are used simultaneously.

Conclusion

In order to define the role of hyperbaric oxygen in the management of acute stroke there is a need to establish a national and international task force to standardize a treatment protocol to be used with the gold standard treatment, in a timely fashion (< 5h), to clarify its real benefit in the management of AHIE.

The timely restoration of cerebral flow in the acute stroke will resolve the ischemic penumbra but does not deal with the metabolic penumbra. To adequately resolve this, it is mandatory to use a treatment option that can restore cellular and tissue oxygen tensions. HBO₂ has proven to be beneficial in the ischemia-reperfusion injury to other parts of the body. There is increasing evidence on the beneficial effects of hyperbaric oxygen in AHIE [124]. Nevertheless, hyperbaric physicians need to produce evidence following the accepted clinical guidelines of the “gold standard treatment” during the same window of opportunity. This can only be accomplished when there are critical care hyperbaric chambers inside the stroke units.

HBO₂ appears to be a safe and very cost-effective treatment for the AHIE of the neonate [125]. It promotes survival of marginal tissue (penumbra), reduces cerebral edema, restores mitochondrial dysfunction, breaks the edema-hypoxia-edema vicious cycle, and improves microcirculation. It also enhances healing, promotes up-regulation of growth factors and reduces, inhibits or prevents IRI. It reduces iNOS-mediated sepsis and shock. Beside the beneficial effects caused by cellular and tissue oxygenation, it has also a very important antioxidant and anti-apoptotic effect [101]. HBO₂ significantly reduces neurologic sequelae and mortality [125].

If we can overcome the fear of treating neonates inside a hyperbaric chamber, develop better and adequate equipment for the management of these patients during HBO₂, and produce a multicenter, multinational, prospective, randomized, and controlled

study; hyperbaric oxygenation will undoubtedly show its beneficial effects in acute hypoxic ischemic encephalopathy of the newborn. This would dramatically change newborn morbidity and mortality and could reduce the global burden of this disease and become a very cost-effective treatment.

References

- Center of Disease Control and Prevention/National Center of Health Statistics. Summary health statistics for U.S. Adults: National Health Interview Survey 2011. *Vital Health Stat* 2012; 10 256: 1-219.
- American Heart Association/American Stroke Association. Health Disease and Stroke Statistics: 2016 Update. *Circulation*.
- Sánchez EC. Mechanisms of action of hyperbaric oxygenation in stroke: a review. *Crit Care Nurs Q*. 2013; 36: 290-298.
- Gonzalez de Dios J, Moya M. Perinatal asphyxia, hypoxic-ischemic encephalopathy and neurological sequelae in full term newborns: an epidemiological study. *Rev Neurol* 1996; 24: 812-819.
- Lawn JE, Cousens S, Zupan J. Lancet Neonatal Survival Steering Team. 4 million neonatal deaths: when? Where? Why?. *Lancet*. 2005; 365: 891-900.
- Lawn J, Shibuya K, Stein C. No cry at birth: global estimates of intrapartum stillbirths and intrapartum-related neonatal deaths. *Bull World Health Organ*. 2005; 83: 409-417.
- Bryce J, Boschi-Pinto C, Shibuya K, Black RE. WHO Child Health Epidemiology Reference Group: WHO estimates of the causes of death in children. *Lancet*. 2005; 365: 1147-1152.
- World Health Organization. *Global Health Estimates: Deaths by Cause, Age, Sex, Country, 2000-2012*. Geneva, WHO, 2014.
- Hong KS, Saver JL, Kang DW, Bae HJ, Yu KH, Koo J, et al. Years of optimum health lost due to complications after acute ischemic stroke: Disability-adjusted life years analysis. *Stroke*. 2010; 41: 1758-1765.
- Xian Y, Holloway RG, Chan PS, Noyes K, Shah MN, Ting HH, et al. Association between stroke center hospitalization for acute ischemic stroke and mortality. *JAMA*. 2011; 305: 373-380.
- Saver JL. Time is brain—quantified. *Stroke*. 2006; 37: 263-266.
- Sanchez EC. Hyperbaric oxygenation in peripheral nerve repair and regeneration. *Neurol Res*. 2007; 29: 184-198.
- Herrmann AG, Deighton RF, Le Bihan T, McCulloch MC, Searcy JL, Kerr LE, et al. Adaptive changes in the neuronal proteome: mitochondrial energy production, endoplasmic reticulum stress, and ribosomal dysfunction in the cellular response to metabolic stress. *J Cereb Blood Flow Metab*. 2013; 33: 673-683.
- Shankaran S, Lupton AR, Ehrenkranz RA, Tyson JE, McDonald SA, Donovan EF, et al. Whole-body hypothermia for neonates with hypoxic-ischemic encephalopathy. *N Engl J Med*. 2005; 353: 1574-1584.
- Penitillia A, Trump BF. The role of cellular membrane systems in shock. *Science*. 1974; 185:277
- Carbonell T, Rama R. Iron, oxidative stress and early neurological deterioration in ischemic stroke. *Curr Med Chem*. 2007; 14: 857-874.
- Davidson SM, Yellon DM, Murphy MP, Duchon MR. Slow calcium waves and redox changes precede mitochondrial permeability transition pore opening in the intact heart during hypoxia and reoxygenation. *Cardiovasc Res*. 2012; 19: 445-453.
- Li G, Wang X, Huang LH, Wang Y, Hao JJ, Ge X, et al. Cytotoxic function of CD8+ T lymphocytes isolated from patients with acute severe cerebral infarction: an assessment of stroke-induced immunosuppression. *BMC Immunol*. 2013.
- Pan JA, Fan Y, Gandhirajan RK, Madesh M, Zong WX. Hyperactivation of the mammalian degenetin MDEG promotes caspase-8 activation and apoptosis. *J Biol Chem*. 2013; 288: 2952-2963.

20. Pilchova I, Klacanova K, Chomova M, Tatkova Z, Dobrota D, Racay P. Possible contribution of proteins of Bcl-2 family in neuronal death following transient global brain ischemia. *Cell MolNeurobiol*. 2015; 35: 23-31.
21. Somarajan BI, Khanday MA, Mallick BN. Rapid eye movement sleep deprivation induces neuronal apoptosis by noradrenaline acting on alpha 1 adrenoceptor and by triggering Mitochondrial intrinsic pathway. *Front Neurol*. 2016; 7:7-25.
22. Brassai A, Suvanjeviev RG, Bán EG, Lakatos M. Role of synaptic and nonsynaptic glutamate receptors in ischaemia induced neurotoxicity. *Brain Res Bull*. 2015; 112: 1-6.
23. Muralikrishna Adibhatla R, Hatcher JF. Phospholipase A2, reactive oxygen species, and lipid peroxidation in cerebral ischemia. *Free Radic Biol Med*. 2006; 40: 376-387.
24. Adibhatla RM, Hatcher JF, Dempsev RJ. Phospholipase A2. hydroxyl radicals, and lipid peroxidation in transient cerebral ischemia. *Antioxid Redox Signal*. 2003; 5: 647-654.
25. Balderas E, Zhang J, Stefani E, Toro L. Mitochondrial BKCa channel. *Front Physiol*. 2015; 6: 104.
26. Li H, Xie Y, Zhang N, Yu Y, Zhang Q, Ding S. Disruption of IP3R2-Mediated Ca2+ signaling pathway in astrocytes ameliorates neuronal death and brain damage while reducing behavioral deficits after focal ischemic Stroke. *Cell Calcium*. 2015; 58: 565-576.
27. Ahlgren H, Bas-Orth C, Freitag HE, Hellwig A, Ottersen OP, Bading H. The nuclear calcium signaling target, activating transcription factor 3 (ATF3), protects against dendrotoxicity and facilitates the recovery of synaptic transmission after an excitotoxic insult. *J BiolChem* 2014; 289: 9970-9982.
28. Kumar VS, Gopalakrishnan A, Naziroğlu M, Rajanikant GK. Calcium ion--the key player in cerebral ischemia. *Curr Med Chem*. 2014; 21: 2065-2075.
29. Sakuma S, Kitamura T, Chihiro K, Kanami T, Sayaka N, Hamashima T, et al. All-trans arachidonic acid generates reactive oxygen species via xanthine dehydrogenase/ oxidase interconversion in the rat liver cytosol. *J Clin Biochem*. 2012; 51: 55-60.
30. Saito Y, Watanabe K, Fujioka D. Disruption of group IVA cytosolic phospholipase A(2) attenuates myocardial ischemia-reperfusion injury partly through inhibition of TNF α mediated pathway. *Am J Physiol Heart Circ Physiol*. 2012; 302: H2018-H2030.
31. Sun XF, Zhang H. NFKB and NFKBI polymorphisms in relation to susceptibility of tumour and other diseases. *Histol Histopathol*. 2007; 22: 1387-1398.
32. Milovanovic M, Volarevic V, Radosavljevic G, Jovanovic I, Pejnovic N, Arsenjevic N, et al. IL-33/ST2 axis in inflammation and immunopathology. *Immunol Res*. 2012; 52: 89-99.
33. Lambertsen KL, Biber K, Finsen B. Inflammatory cytokines in experimental and human stroke. *J Cereb Blood Flow Metab*. 2012; 32: 1677-1698.
34. Coucha M, Li W, Ergui A. The effect of endothelin receptor A antagonism on basilar artery endothelium-dependent relaxation after ischemic stroke. *Life Sci*. 2012; 91: 676-680.
35. Zeng L, Liu J, Wang Y, Wang L, Weng S, Chen S, et al. Cocktail blood biomarkers: prediction of clinical outcomes in patients with acute ischemic stroke. *Eur Neurol*. 2013; 69: 68-75.
36. Li Y, Zhou C, Calvert JW, Colohan AR, Zhang JH. Multiple effects of hyperbaric oxygen on the expression of HIF-1 alpha and apoptotic genes in a global ischemia-hypotension rat model. *Exp Neurol*. 2005; 191: 198-210.
37. Kunz AB, Kraus J, Young P, Reuss R, Wipfler P, Oschmann P, et al. Biomarkers of inflammation and endothelial dysfunction in stroke with and without sleep apnea. *Cerebrovasc Dis*. 2012; 33: 453-460.
38. Supanc V, Biloglav Z, Kes VB, Demarin V. Role of cell adhesion molecules in acute ischemic stroke. *Ann Saudi Med*. 2011; 31: 365-370.
39. Caimi G, Canino B, Ferrara F, Montana M, Musso M, Porreto F, et al. Granulocyte integrins before and after activation in acute ischaemic stroke. *J NeurolSci*. 2001; 186: 23-26.
40. Chiba T, Itoh T, Tabuchi M, Nakazawa T, Satou T. Interleukin-1 accelerates the onset of stroke in stroke-prone spontaneously hypertensive rats. *Mediators Inflamm*. 2012; 2012: 701976.
41. Higashimori A, Morioka N, Shiotani S, Fujihara M, Fukuda K, Yokoi Y. Long-term results of primary stenting for subclavian artery disease. *Catheter Cardiovascular Interv*. 2013.
42. Liu S, Levine SR, Winn HR. Targeting ischemic penumbra: part I - from pathophysiology to therapeutic strategy. *J Exp Stroke Transl Med*. 2010; 3: 47-55.
43. Georgiadis D, Schwarz S, Kollmar R, Schwab S. Endo vascular cooling for moderate hypothermia in patients with acute stroke: first results of a novel approach. *Stroke*. 2001; 32: 2550-2553.
44. Powers WJ, Derdeyn CP, Biller J, Coffey CS, Hoh BL, Jauch EC, et al. 2015 AHA/ASA focused update of the 2013 guidelines for the early management of patients with acute ischemic stroke regarding endovascular treatment. *Stroke*. 2015.
45. The National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group. Tissue plasminogen activator for acute ischemic stroke. *N Engl J Med*. 1995; 333: 1581-1587.
46. Wardlaw JM, del Zoppo G, Yamaguchi T. Thrombolysis for acute ischaemic stroke (Cochrane Review). *The Cochrane Library*. 2002.
47. Jeffers BW, Robbins J, Bhamri R. Efficacy of Calcium Channel Blockers Versus Other Classes of Antihypertensive Medication in the Treatment of Hypertensive Patients With Previous Stroke and/or Coronary Artery Disease: A Systematic Review and Meta-Analysis. *Am J Ther*. 2015.
48. Al Alshaik S, Quinn T, Dunn W, Walters M, Dawson J. Multimodal interventions to enhance adherence to secondary preventive medication after stroke: A systematic review and meta-analyses. *Cardiovasc Ther*. 2016; 34: 85-93.
49. Mukete BN, Cassidy M, Ferdinand KC, Le Jemtel TH. Long-Term Anti-Hypertensive Therapy and Stroke Prevention: A Meta-Analysis. *Am J Cardiovasc Drugs*. 2015; 15: 243-257.
50. Hu Z, Zhong B, Tan J, Chen C, Lei Q, Zeng L. The Emerging Role of Epigenetics in Cerebral Ischemia. *Mol Neurobiol*. 2016.
51. Hu Z, Yu D, Almeida-Suhett C, Tu K, Marini AM, Eiden L, et al. Expression of miRNAs and their cooperative regulation of the pathophysiology in traumatic brain injury. *PLoS One*. 2012; 7: 39357.
52. Gessell LB. *Hyperbaric Oxygen Therapy Indications*. Durham, NC. Undersea and Hyperbaric Medical Society, 2008.
53. Calzia E, Asfar P, Hauser B, Matejovic M, Ballestra C, Radermacher P, et al. Hyperoxia may be beneficial. *Crit Care Med*. 2010; 38: S559-568.
54. Sheikh AY, Rollins MD, Hopf HW, Hunt TK. Hyperoxia improves microvascular perfusion in a murine wound model. *Wound Repair Regen*. 2005; 13: 303-308.
55. Avtan SM, Kaya M, Orhan N, Arslan A, Arican N, Toklu AS, et al. The effects of hyperbaric oxygen therapy on blood-brain barrier permeability in septic rats. *Brain Res*. 2011; 1412: 63-72.
56. Sun L, Zhou W, Mueller C, Sommer C, Heiland S, Bauer AT, et al. Oxygen therapy reduces secondary hemorrhage after thrombolysis in the cerebral ischemia. *J Cereb Blood Flow Metab* 2010; 30: 1651-1660.
57. Fukuda AM, Badaut J Aquaporin 4: a player in cerebral edema and neuroinflammation. *J Neuroinflammation*. 2012; 9: 279.
58. Badaut J, Fukuda AM, Jullienne A, Petry KG. Aquaporin and brain diseases. *Biochim Biophys Acta*. 2014; 1840: 1554-1565.
59. Sirin Y, Olgac V, Dogru-Abbasoglu S, Tapul L, Aktas S, Soley S. The influence of hyperbaric oxygen treatment on the healing of experimental defects filled with different bone graft substitutes. *Int J Med Sci*. 2011; 8:114-125.
60. Cross KJ, Mustoe TA. Growth factors in wound healing. *Surg Clin North Am*. 2003; 83: 531-545, vi.

61. Duan S, Shao G, Yu L, Ren C. Angiogenesis contributes to the neuroprotection induced by hyperbaric oxygen preconditioning against focal cerebral ischemia in rats. *Int J Neurosci*. 2015;125: 625-634.
62. Johnston BR, Ha AY, Brea B, Liu PY. The Mechanism of Hyperbaric Oxygen Therapy in the Treatment of Chronic Wounds and Diabetic Foot Ulcers. *R I Med J* (2013). 2016; 99: 26-29.
63. Gothard L, Stanton A, MacLaren J, Lawrence D, Hall E, Mortimer P, et al. Non-randomized phase II trial of hyperbaric oxygen therapy in patients with chronic arm lymphedema and tissue fibrosis after radiotherapy for early breast cancer. *RadiotherOncol*. 2004; 70: 217-224.
64. Tal S, Hadanny A, Berkovitz N, Sasson E, Ben-Jacob E, Efrati S. hyperbaric oxygen may induce angiogenesis in patients suffering from prolonged post-concussion syndrome due to traumatic brain injury. *RestorNeurolNeurosci*. 2015; 33: 943-951.
65. Huang ET, Mansouri J, Murad MH, Joseph WS, Strauss MB, Tettelbach W, et al. A clinical practice guideline for the use of hyperbaric oxygen therapy in the treatment of diabetic foot ulcers. *Undersea Hyperb Med*. 2015; 42: 205-247.
66. Lima FL, Joazeiro PP, Lancellotti M, de Hollanda LM, de Araújo Lima B, Linares E, et al. Effects of hyperbaric oxygen on *Pseudomonas aeruginosa* susceptibility to imipenem and macrophages. *Future Microbiol*. 2015; 10: 179-189.
67. CimÄYit M, Uzun G, Yildiz S. Hyperbaric oxygen therapy as an anti-infective agent. *Expert Rev Anti Infect Ther*. 2009; 7: 1015-1026.
68. Jan A, Sándor GK, Brkovic BB, Peel S, Kim YD, Xiao WZ, et al. Effect of hyperbaric oxygen on demineralized bone matrix and biphasic calcium phosphate bone substitutes. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2010; 109: 59-66.
69. Grassmann JP, Schnependahl J, Halimi AR, Herten M, Betsch M, Lögters TT, et al. Hyperbaric oxygen therapy improves angiogenesis and bone formation in critical sized diaphyseal defects. *J Othop Res*. 2015; 33: 513-520.
70. Grassmann JP, Schnependahl J, Sager M, Hakimi AR, Herten M, Loegters TT, et al. The effect of bone marrow concentrate and hyperbaric oxygen oxygen therapy on bone repair. *J Mater Sci Mater Med*. 2015; 26: 5331.
71. Verrazzo G, Coppola L, Luongo C, Sammartino A, Giunta R, Grassia A, Ragone R, et al. Hyperbaric oxygen, oxygen-ozone therapy and rheologic parameters of blood in patients with peripheral occlusive arterial disease. *Undersea Hyperb Med*. 1995; 22: 17-22.
72. Imperatore F, Cuzzocrea S, Luongo C, Liguori G, Scafuro A, De Angelis A, et al. Hyperbaric oxygen therapy prevents vascular defragment during zymosan-induced multiple-organ-failure syndrome. *Intensive Care Med*. 2004; 30: 1175-1181.
73. Imperatore F, Cuzzocrea S, De Lucia D, Sessa M Rinaldi B, Capuano A, Liguori G, et al. Hyperbaric oxygen therapy prevents coagulation disorders in an experimental model of multiple organ failure syndrome. *Intensive Care Med*. 2006; 32: 1881-1888.
74. Gomez CR, Knutson GJ, Clifton KB, Schreiber CA, Vuk-PavloviÄž S. Age-dependent response of murine female bone marrow cells to hyperbaric oxygen. *Biogerontology*. 2012; 13: 287-297.
75. Zhou HX, Liu ZG, Liu XJ, Chen QX. Umbilical cord-derived mesenchymal stem cell transplantation combined with hyperbaric oxygen treatment for repair of traumatic brain injury. *Neural Regen Res*. 2016; 11: 107-113.
76. Geng CK, Cao HH, Ying X, Yu HL. Effect of mesenchymal stem cells transplantation combining with hyperbaric oxygen therapy on rehabilitation of rat spinal cord injury. *Asian Pac J Trop Med*. 2015; 8: 468-473.
77. Sun L, Strelow H, Mies G, Veltkamp R. Oxygen therapy improves energy metabolism in focal cerebral ischemia. *Brain Res*. 2011; 1415: 103-108.
78. Holbach KH, Caroli A, Wassmann H. Cerebral energy metabolism in patients with brain lesions of normo- and hyperbaric oxygen pressures. *J Neurol*. 1977; 217: 17-30.
79. Cheng O, Ostrowski RP, Wu B, Liu W, Cheng C, Zhang JH. Cyclooxygenase-2 mediates hyperbaric oxygen preconditioning in the rat model of transient global cerebral ischemia. *Stroke* 2011; 42: 484-490.
80. Zeng Y, Xie K, Dong H, Zhang H, Wang F, Li Y, Xiong L. Hyperbaric oxygen preconditioning protects cortical neurons against oxygen-glucose deprivation injury: role of peroxisome proliferator-activated receptor-gamma. *Brain Res*. 2012; 1452: 140-150.
81. Gois E Jr, Daniel RA, Parra RS, Almeida AL, Rocha JJ, Garcia SB, et al. Hyperbaric oxygen therapy reduces COS-2 expression in a dimethylhydrazine-induced rat model of colorectal carcinogenesis. *Undersea Hyperb med*. 2012; 39: 693-698.
82. Pedoto A, Nandi J, Yang ZJ, Wang J, Bosco G, Oler A, et al. Beneficial effect of hyperbaric oxygen pretreatment on lipopolysaccharide-induced shock in rats. *Clin Exp Pharmacol Physiol*. 2003; 30: 482-488.
83. Weisz G, Lavy A, Adir Y, Melamed Y, Rubin D, Eidelman S, et al. Modification of *in vivo* and *in vitro* TNF-alpha, IL-1, and IL-6 secretion by circulating monocytes during hyperbaric oxygen treatment in patients with perianal Crohn's disease. *J Clin Immunol*. 1997; 17: 154-159.
84. Shyu WC, Lin SZ, Saeki K, Kubosaki A, Matsumoto Y, Onodera T, et al. Hyperbaric oxygen enhances the expression of prion protein and heat shock protein 70 in a mouse neuroblastoma cell line. *Cell MolNeurobiol*. 2004; 24: 257-268.
85. Rosenthal RE, Silbergleit R, Hof PR, Haywood Y, Fiskum G. Hyperbaric oxygen reduces neuronal death and improves neurological outcome after canine cardiac arrest. *Stroke*. 2003; 34: 1311-1316.
86. Wada K, Miyazawa T, Nomura N, Tsuzuki N, Nawashiro H, Shima K. Preferential conditions for and possible mechanisms of induction of ischemic tolerance by repeated hyperbaric oxygenation in gerbil hippocampus. *Neurosurgery*. 2001; 49: 160-166.
87. Rothfuss A, Speit G. Overexpression of heme oxygenase-1 (HO-1) in V79 cells results in increased resistance to hyperbaric oxygen (HBO)-induced DNA damage. *Environ Mol Mutagen*. 2002; 40: 258-265.
88. Buras JA, Stahl GL, Svoboda KK, Reenstra WR. Hyperbaric oxygen downregulates ICAM-1 expression induced by hypoxia and hypoglycemia: the role of NOS. *Am J Physiol Cell Physiol*. 2000; 278: C292-302.
89. Thom SR. Functional inhibition of leukocyte B2 integrins by hyperbaric oxygen in carbon monoxide-mediated brain injury in rats. *Toxicol Appl Pharmacol*. 1993; 123: 248-256.
90. Huang ZX, Kang ZM, Gu GJ, Peng GN, Yun L, Tao HY, et al. Therapeutic effects of hyperbaric oxygen in a rat model of endothelin-1- induced focal cerebral ischemia. *Brain Res*. 2007; 1153: 204-213.
91. Jones SR, Carpin KM, Woodward SM, Khiabani KT, Stephenson LL, Wang WZ, et al. Hyperbaric oxygen inhibits ischemia-reperfusion-induced neutrophil CD18 polarization by a nitric oxide mechanism. *Plast Reconstr Surg*. 2010; 126: 403-411.
92. Godman CA, Chheda KP, Hightower LE, Perdrizet G, Shin DG, Giardina C. Hyperbaric oxygen induces a cytoprotective and angiogenic response in human microvascular endothelial cells. *Cell Stress Chaperones*. 2010; 15: 431-442.
93. Godman CA, Joshi R, Giardina C, Perdrizet G, Hightower LE. Hyperbaric oxygen treatment induces antioxidant gene expression. *Ann N Y Acad Sci*. 2010; 1197: 178-183.
94. Rothfuss A, Speit G. Investigations on the mechanisms of hyperbaric oxygen (HBO)-induced adaptive protection against oxidative stress. *Mutat Res*. 2002; 508: 157-165.
95. Haapaniemi T, Sirsjo A, Nylander G, Larsson J. Hyperbaric oxygen treatment attenuates glutathione depletion and improves metabolic restitution in posts ischemic skeletal muscle. *Free Rad Res*. 1995; 23: 91-101.
96. Bitar MS. The GSK-3 β /Fyn/Nrf2 pathway in fibroblasts and wounds of type 2 diabetes: On the road to an evidence-based therapy of non-healing wounds. *Adipocyte*. 2012; 1: 161-163.

97. Bennett MH, Weibel S, Wasiak J, Schnabel A, French C, Kranke P. Hyperbaric oxygen therapy for acute ischaemic stroke. *Cochrane Database Syst Rev*. 2014.
98. Sánchez EC. Use of hyperbaric oxygenation in neonatal patients: a pilot study of 8 patients. *Crit Care Nurs Q*. 2013; 36: 280-289.
99. Bhaskaran M, Xi D, Wang Y, Huang C, Narasaraju T, Shu W, et al. Identification of microRNAs changed in the neonatal lungs in response to hyperoxia exposure. *Physiol Genomics*. 2012; 44: 970-980.
100. Speer CP. Neonatal respiratory distress syndrome: an inflammatory disease? *Neonatology*. 2011; 99: 316-319.
101. Ferrari DC, Nestic O, Perez-Polo JR. Perspectives on neonatal hypoxia/ischemia-induced edema formation. *Neurochem Res*. 2010; 35: 1957-1965.
102. Thornton JS, Ordidge RJ, Penrice J, Cady EB, Amess PN, Punwani S, et al. Temporal and anatomical variations of brain water apparent diffusion coefficient in perinatal cerebral hypoxic-ischemic injury: relationships to cerebral energy metabolism. *Magn Reson Med*. 1998; 39: 920-927.
103. Zhang N, LH, Lu L. Pathogenic factors of convulsion in neonates with hypoxic-ischemic encephalopathy. *Di Yi Jun Yi Da Xue Xue Bao*. 2002; 22: 1039-1041.
104. Hutchison JH, Kerr MM, Williams KG, Hopkinson WI. Hyperbaric Oxygen in the resuscitation of the newborn. *Lancet*. 1963; 2: 1019-1022.
105. Lawn JE, Cousens S, Zupan J. Lancet Neonatal Survival Steering Team 4 million neonatal deaths: when? Where? Why? *Lancet*. 2005; 365: 891-900.
106. Lawn J, Shibuya K, Stein C. No cry at birth: global estimates of intrapartum stillbirths and intrapartum-related neonatal deaths. *Bull World Health Organ*. 2005; 83: 409-417.
107. Bryce J, Boschi-Pinto C, Shibuya K, Black RE. WHO Child Health Epidemiology Reference Group WHO estimates of the causes of death in children. *Lancet*. 2005; 365: 1147-1152.
108. Bozok S, İlhan G, Yılmaz Y, Dökümcü Z, Tumkaya L, Karamustafa H, et al. Protective effects of hyperbaric oxygen and iloprost on ischemia/reperfusion-induced lung injury in a rabbit model. *Eur J Med Res*. 2012; 17: 14.
109. Caldeira DE, Souza ME, Gomes MC, Picinato MA, Fina CF, Feres O, et al. Effects of hyperbaric oxygen (HBO), as pre-conditioning in liver of rats submitted to periodic liver ischemia/reperfusion. *Acta Cir Bras*. 2013; 1: 66-71.
110. Daniel RA, Cardoso VK, Góis Jr E, Parra RS, Garcia SB, Rocha JJ, et al. Effect of hyperbaric oxygen therapy on the intestinal ischemia reperfusion injury. *Acta Cir Bras*. 2011; 26: 463-469.
111. Ramalho Rj, de Oliveira PS, Cavaglieri RC, Medeiros PR, Filho DM, Poli-de-Figueredo LF, et al. Hyperbaric oxygen therapy induces kidney protection in an ischemia/reperfusion model in rats. *Transpl Proc*. 2012; 44: 233-236.
112. Thom SR. Hyperbaric oxygen: its mechanisms and efficacy. *Plast Reconstr Surg*. 2011; 127: 131S-141S.
113. Zhao H, Zhang Q, Xue Y, Chen X, Haun RS. Effects of hyperbaric oxygen on the expression of claudins after cerebral ischemia-reperfusion in rats. *Exp Brain Res*. 2011; 212: 109-117.
114. Calvert JW, Yin W, Patel M, Badr A, Mychaskiw G, Parent A, et al. Hyperbaric oxygenation prevented brain injury induced by hypoxia-ischemia in a neonatal rat model. *Brain Res*. 2002; 951: 1-8.
115. Wang XL, Zhao YS, Yang YJ, Xie M, Yu XH. Therapeutic window of hyperbaric oxygen therapy for hypoxic-ischemic brain damage in newborn rats. *Brain Res*. 2008; 1222: 87-94.
116. Guven A, Gundogdu G, Uysal B, Cermik H, Kul M, Demirbag S, et al. Hyperbaric oxygen therapy reduces the severity of necrotizing enterocolitis in a neonatal rat model. *J Pediatr Surg*. 2009; 44: 534-540.
117. Calvert JW, Zhou C, Zhang JH. Transient exposure of rat pups to hyperoxia at normobaric and hyperbaric pressures does not cause retinopathy of prematurity. *Exp Neurol*. 2004; 189: 150-161.
118. Ricci B, Calogero G. Oxygen induced retinopathy in newborn rats: effects of prolonged normobaric and hyperbaric oxygen supplementation. *Pediatrics*. 1988; 82:193-198.
119. Ricci B, Calogero G, Lepore D. Variations in the severity of retinopathy seen in newborn rats supplemented with oxygen under different conditions of hyperbarism. *Exp Eye Res*. 1989; 49: 789-797.
120. Ricci B, Minicucci G, Manfredi A, Santo A. Oxygen-induced retinopathy in the newborn rat: effects of hyperbarism and topical administration of timolol maleate. *Graefes Arch Clin Exp Ophthalmol*. 1995; 233: 226-230.
121. Jacobs SE, Berg M, Hunt R, Tarnow-Mordi WO, Inder TE, Davis PG. Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database of Sys Rev*. 2013.
122. Mc Cormick JG, Houle TT, Saltzman HA, Whaley RC, Roy RC. Treatment of acute stroke with hyperbaric oxygen: time window for efficacy. *Undersea Hyperb Med*. 2011; 38: 321-334.
123. Liu Z, Xiong T, Meads C. Clinical effectiveness of treatment with hyperbaric oxygen for neonatal hypoxic-ischemic encephalopathy: systematic review of Chinese literature. *BMJ*. 2006; 333: 374-382.
124. Zhou BY, Lu GJ, Huang YQ, Ye ZZ, Han YK. Efficacy of hyperbaric oxygen therapy under different pressures on neonatal hypoxic-ischemic encephalopathy. *Zhongguo Dang Dai Er Ke Za Zhi*. 2008; 10:133-135.
125. Asif MJ, Exline MC. Utilization of hyperbaric oxygen therapy and induced hypothermia after hydrogen sulfide exposure. *Respir Care*. 2012; 57: 307-310.