

Review Article

Different Co-Culture Systems Have a Useful Impact on Preimplantation Embryo Development

Samy R¹ and Ghanem N^{1,2*}¹Animal Production Department, Faculty of Agriculture, Cairo University, Giza, Egypt²Cairo University Research Park, Faculty of Agriculture, Cairo University, Egypt

*Corresponding author: Nasser Ghanem, Animal Production Department, Faculty of Agriculture, Cairo University Research Park, Cairo University, Egypt

Received: March 01, 2021; Accepted: April 03, 2021;

Published: April 10, 2021

Abstract

The production of *in vitro* produced embryos of good morphological quality and viability is a prerequisite for successful assisted reproduction biotechnologies in animal breeding and human. The co-culturing system has been applied to improve preimplantation development that could subsequently resulted in successful pregnancy. There are different types of reproductive and non-reproductive cells that have been used during preimplantation development. The most well-known reproductive cells are those recovered from ovaries (cumulus and granulosa cells), oviduct and endometrium cells. While, in last decade stem cells such as mesenchymal stem cells and murine embryonic fibroblasts that originated from different tissues have been used to support early embryonic development. The positive effect co-culturing system was suggested to be due to direct mechanical cell-to-cell contact that occurred between the dividing embryos embryo and helper cells in addition to secretions of various bioactive biological components like growth factors and scavenging the deleterious byproducts that resulted from embryo metabolism. In current review, we will highlight the effects of different culture systems on embryo development and their suggested mechanisms to exert the beneficial impacts.

Keywords: Oocytes; Embryos; Co-culture; Mode of action

Co-culture Systems Importance

Co-culture system has a supportive effect on embryo normal development by strengthen the embryo ability to overcome any damage occurred in early stage according to several mechanisms include: 1) Detoxifying the culture medium from heavy metal ions by chelation; 2) Secretion of nutrients, substrates and embryotropic factors like amino acids and growth factors into the medium to induce embryonic genome activation and improve the cell structure [1]; 3) Stabilization of the pH and gases concentrations (CO₂ and O₂) and decreasing the oxygen tension [2].

Co-culture Systems

Different co-culture systems support early embryo development according to the types of the cells such as cumulus cells, granulosa cells, oviduct cells, uterine cell monolayers, liver cell monolayers, chicken skin cell monolayers mouse testicular cell monolayers and monkey kidney cells [3,4].

Pre-antral follicles growth and survival rates increase when co-cultured with mesenchymal cells because its secretions providing some important factors as extracellular matrix proteins (1), high molecular mass proteins (6), basement membrane components [5], Transforming Growth Factor- (TGF-) [6], Fibroblast Growth Factor-7 (FGF-7) and Hepatocyte Growth Factor (HGF) [7]. Moreover, HGF and KGF and Kit Ligand protein (KL) expression indirectly regulate the roles of gonadotropins on follicular development [8]. Fibroblast Growth Factor-7 (FGF-7) suppresses granulosa cells apoptosis and stimulates the growth of cultured rat pre-antral follicles [9].

A specific type of cells known as VERO are kind of epithelial cells

derived from the kidney of African green monkey At (*Cerpopithecus aethiops*) and the high oxygen concentration is suitable for VERO culture [10]. High oxygen level (20%), the VERO cells increase the bovine blastocyst rate, total cell number and the ability of blastocyst to be tolerant with cryopreservation [11-13].

Cumulus and Granulosa Cells

Cumulus and granulosa cells have supportive effects through cell to cell communication and selective transport for important nutrients from the culture medium to the developed oocytes and embryos. Moreover, they have the ability to synthesis steroid hormones [14] such as activin [15], inhibin [16], thecal differentiation factor [17], fibronectin [18] may cause enhancement in pre-antral follicles growth and survivability.

Granulosa cells secrete many beneficial proteins like Kit Ligand protein, which regulates thecal cells function, proliferation and growth [19].

Human oocytes co-cultured with cumulus cells during *in vitro* maturation caused a significant increase in 8-cell stage embryos after 72 hour compared with oocytes matured in control medium (Mansour et al. [14]). In addition co-culture system has a beneficial impact on human embryos development rates, quality and pregnancy rates which increased by the co-culturing embryos produced by IVF with various cellular monolayers [14,20-26].

Porcine pre-antral follicles co-cultured with cumulus cells from antral follicles with diameter more than 3mm resulted in improvement of growth rates [27]. Porcine cumulus and mural granulosa cells play important role in cumulus cells expansion by producing Cumulus Expansion Enabling Factor (CEEF) *in vitro* [28].

Oviductal and Uterine Cells

Fertilization and early embryonic development till blastocyst stage occur in the oviduct, which provides the suitable environment for gametes survival, early pregnancy success and can alter the embryo gene expression, epigenetic modification and metabolism [29]. However, the crucial secretions of oviduct such as glycoproteins, amino acids, lactate, and growth factors like Insulin-Like Growth Factor (IGF), Interleukin (IL)-1 and Platelet-Derived Growth Factor (PDGF) [2,30], some oviduct secretions are identified as chemokines, cytokines, growth factors and apoptosis regulators [31]. Therefore, to increase the *in vitro* Fertilization (IVF) rate, IVF protocols were developed using co-culture systems with oviduct epithelial cells in sheep [32], cattle [33] and mice [29].

Pre-antral follicles co-cultured with Oviductal Epithelial Cells (OEC) showed a decrease in pre-antral follicles growth and survivability rates due to OEC secretion in oviduct is necessary for embryo development and blastocyst formation [34]. Co-culture embryos with oviduct monolayer resulted in an increase in blastocyst rates at oxygen concentration (20%) compared to oxygen level (5%) [35]. Several studies reported that the effect of co-culture involved also the cells, the morphological changes occur when the cells co-cultured as a monolayer [36,37] which impact on gene expression [38] so as an expected result the effect of co-culture on embryo development may be altered during the co-culture period.

Mode of Action and Molecular Impact of Co-culture

Despite the production of several commercial defined embryo culture media, co-culture systems with different types of somatic cells is still used for preimplantation development in many mammalian species. Taken into consideration that there is interest in application of co-culture either autologous or heterologous systems, the actual mechanism explaining their effects is not yet well established. However, there is suggested mode of actions that are most probable justify action of co-culture; there is no evidence yet to identify the correct mechanism of the co-culture effect. In fact, the mode of action of co-culture systems has been explained largely by two mechanisms [2,39].

The first mode is dependent on the scavenging ability of co-culture system in removing the deleterious components that resulted from embryo metabolism and ameliorating effect of oxidative stress that resulted from *in vitro* the culture conditions (medium, composition, pH, and osmolality and oxygen tension). The second possible mode of co-culture action is dependent on bioactive secretions of helper cells into culture medium or what is called embryo trophic factors. In experiment done in bovine embryos [40-46] produced *in vitro* that were co-cultured with adipose tissue-derived from bovine mesenchymal cells which resulted in increasing blastocyst formation rate and quality as assessed through increased number of total embryo cells and upregulation of metabolism related transcript known as G6PDH and gene regulating embryo differentiating (POU5F1).

Conclusion

It could be implicated that co-culture of mammalian embryos with helper cells could be more beneficial than the traditional cells

free system. However, the co-culture system and its conditions should be modified and optimized for each species and type of cells. The co-culture micro environmental conditions should be monitored in order to understand the modulatory potential actions of this system and could be applied in standard way.

References

- Gandolfi F, Moor RM. Stimulation of early embryonic development in the sheep by co-culture with oviduct epithelial cells. *Journal of Reproduction and Fertility*. 1987; 81: 23-28.
- Bongso A, Fong CY, Ng SC, Ratnam S. The search for improved *in vitro* systems should not be ignored embryo co-culture may be one of them. *Hum Reprod*. 1993; 8: 1155-1162.
- Goto K, Kajihara Y, Kosaka S, Koba M, Nakanishi Y, Ogawa K. Pregnancies after co-culture of cumulus cells with bovine embryos derived from *in vitro* fertilization of *in vitro* matured follicular oocytes. *J Reprod Fertil*. 1988; 83: 753-758.
- Bavister BD. Culture of preimplantation embryos: facts and artifacts *Hum. Reprod Update*. 1995; 1: 91-148.
- Gosden RG, Boland NI, Spears N, Murry AA, Chapman M, Wade JC, et al. The biology and technology of follicular oocyte development *in vitro*. *Reprod Med Rev*. 1993; 2: 129-152.
- Skinner MK, Coffey RJ. Regulation of ovarian cell growth through the local production of transforming growth factor alpha by theca cells. *Endocrinol*. 1988; 123: 2632-2638.
- Parrot JA, Skinner MK. Thecal cell, granulosa cell interaction involves a positive feedback loop among keratinocyte growth factor and kit ligand during ovarian follicular development. *Endocrinol*. 1998; 139: 2240-2245.
- Parrot JA, Vigne JL, Chu BZ, Skinner MK. Mesenchymal-epithelial interactions in the ovarian follicle involve keratinocyte and hepatocyte growth factor production by thecal cells and their action on granulosa cells. *Comparative Study Endocrinology*. 1994; 135: 569-575.
- McGee EA, Chun SY, Lai S, He Y, Hsueh AJ. Keratinocyte growth factor promotes the survival, growth and differentiation of preantral follicles. *Fertility and Sterility*. 1999; 71: 732-738.
- Ammerman NC, Beier-Sexton M, Azad AF. Growth and maintenance of Vero cell lines. *Current Protocols in Microbiology*. 2008; 11: A.4E.1-A.4E.7.
- Ouhibi N, Menezes Y, Benet G, Nicollet B. Culture of epithelial cells derived from the oviduct of different species. *Human Reproduction*. 1989; 4: 229-235.
- Menck MC, Guyader-Joly C, Peynot N, Le Bourhis D, Lobo RB, Renard JP. Beneficial effects of Vero cells for developing IVF bovine eggs in two different coculture systems. *Reproduction Nutrition Development*. 1997; 37: 141-150.
- Carnegie JA, Morgan JJ, McDiarmid N, Durnford R. Influence of protein supplements on the secretion of leukaemia inhibitory factor by mitomycin-pretreated Vero cells: possible application to the *in vitro* production of bovine blastocysts with high cryotolerance. *Journal of Reproduction and Fertility*. 1999; 117: 41-48.
- Mansour RT, Aboulghar MA, Serour IG, Abbas AM. Coculture of human pronucleate oocytes with their cumulus cells. *Hum Reprod*. 1994; 9: 1727-1729.
- Li R, Phillips DM, Mather JP. Activin promotes ovarian follicle development *in vitro*. *Endocrinol*. 1995; 136: 849-856.
- Campbell BK, Scaramuzzi RJ, Webb R. Control of antral follicle development and selection in sheep and cattle. *J Reprod Fertility Suppl*. 1995; 49: 325-350.
- Magarelli PC, Zachow RJ, Magoffin DA. Development and hormonal regulation of rat theca cell differentiation factor secretion in ovarian follicles. *Biol Reprod*. 1996; 55: 416-420.
- Carnegie JA. Secretion of fibronectin by granulosa cells occurs primarily during early follicular development. *J Reprod Fert*. 1990; 89: 579-589.
- Parrot JA, Skinner MK. Direct actions of KL on theca cell growth and

- differentiation during follicle development. *Endocrinol.* 1997; 138: 3819-3827.
20. Wiemer KE, Cohen J, Amborski GF, Wright G, Wiker S, Munyakazi L, et al. *In vitro* development and implantation of human embryos following culture on fetal bovine uterine fibroblast cells. *Hum. Reprod.* 1989; 4: 595-600.
 21. Menezo Y, Guerin JF, Czyba JC. Improvement of human early embryo development *in vitro* by co-culture on monolayers of Vero cells. *Biol Reprod.* 1990; 42: 301-306.
 22. Bongso A, Ng SC, Fong CY, Ramam S. Co-cultures a new lead in embryo quality improvement for assisted reproduction. *Feml Stenl.* 1991; 56: 179-191.
 23. Menezo Y, Hazout A, Dumont M, Herbaut N, Nicollet B. Co-culture of embryos on Vero cells and transfer of blastocysts in humans *Hum. Reprod.* 1992; 7: 101-106.
 24. Gregory L, Booth AD, Wells C, Walker SM. A study of the cumulus-corona cell complex *in-vitro* fertilization and embryo transfer, a prognostic indicator of the failure of implantation. *Hum. Reprod.* 1994; 9: 1308-1317.
 25. Tucker MJ, Ingargiola PE, Massey JB, Morton PC, Wiemer KE, Wiker SR, et al. Assisted hatching with or without bovine oviductal epithelial cell co-culture for poor prognosis *in-vitro* fertilization patients. *Hum. Reprod.* 1994; 9: 1528-1531.
 26. Freeman MR, Whitworth CM, Hill GA. Granulosa cell co-culture enhances human embryo development and pregnancy rate following *in-vitro* fertilization *Hum. Reprod.* 1995; 10: 408-414.
 27. Wu MF, Huang WT, Tsay CHF, Liu BT, Chiou CM, Yen SC, et al. The stage-dependent inhibitory effect of porcine follicular cells on the development of preantral follicles. *Anim Reprod Sci.* 2002; 73: 73-88.
 28. Prochazka R, Naggova EE, Brem G, Schellander K, Motlik J. Secretion of Cumulus Expansion Enabling Factor (CEEF) in porcine follicles. *Mol.Reprod. Develop.* 1998; 49: 141-149.
 29. Watkins AJ, Papenbrock T, Fleming TP. The preimplantation embryo: handle with care. *Seminars in Reproductive Medicine.* 2008; 26: 175-185.
 30. Moreau GM, Arslan A, Douglas DA. Development of immortalized endometrial epithelial and stromal cell lines from the mink (*Mustela vison*) uterus and their effects on the survival *in vitro* of mink blastocysts in obligate diapause. *Biol Reprod.* 1995; 53: 511-518.
 31. Kolle S, Reese S, Kummer W. New aspects of gamete transport, fertilization, and embryonic development in the oviduct gained by means of live cell imaging. *Theriogenology.* 2010; 73: 786-795.
 32. Gandolfi E, Tiziana A, Brevini L, Brown CR, Moor RM. Characterization of proteins secreted by sheep oviduct epithelial cells and their function in embryonic development. *Development.* 1989; 106: 303-312.
 33. Eyestone WH, First NL. Co-culture of early cattle embryos to the blastocyst stage with oviducal tissue or in conditioned medium. *Journal of Reproduction and Fertility.* 1989; 85: 715-720.
 34. White KL, Hebnke K, Rickords LF, Southerm LL, Thompson DL, Wood TC. Early embryonic development *in vitro* by co-culture with oviductal epithelial cells in pigs. *Biol Reprod.* 1989; 41: 425-430.
 35. Clemente M, de la Fuente J, Lonergan P, Gutierrez-Adan A, Rizos D. Effect of oxygen tension on embryo development and gene transcription of bovine blastocysts produced *in vitro* by co-culture with oviduct epithelial cells. *Biology of Reproduction.* 2008; 78: 133.
 36. Rief S, Sinowatz F, Stojkovic M, Einspanier R, Wolf E, Prella K. Effects of a novel co-culture system on development, metabolism and gene expression of bovine embryos produced *in vitro*. *Reproduction.* 2002; 124: 543-556.
 37. Tahir MZ, George F, Donnay I. Comparison of different membrane supports for monolayer culture of bovine oviduct epithelial cells. *BMC Proceedings.* 2011; 5: P117.
 38. Schmaltz-Panneau B, Locatelli Y, Uzbekova S, Perreau C, Mermillod P. Bovine oviduct epithelial cells dedifferentiate partly in culture while maintaining their ability to improve early embryo development rate and quality. *Reproduction in Domestic Animals.* 2015; 50: 719-729.
 39. Joo BS, Kim MK, Na YJ, Moon HS, Lee KS, Kim HD. The mechanism of action of coculture on embryo development in the mouse model: direct embryo-to-cell contact and the removal of deleterious components. *Fertil Steril.* 2001; 75: 193-199.
 40. Miranda MS, Nascimento HS, Costa MP, Costa NN, Brito KN, Lopes CT, et al. Increasing of blastocyst rate and gene expression in co-culture of bovine embryos with adult adipose tissue-derived mesenchymal stem cells. *Assist Reprod Genet.* 2016; 33: 1395-1403.
 41. Ascari IJ, Martins SC, Camargo LSA, Mendez-Otero R, Jasmin. Development of bovine embryos *in vitro* in coculture with murine mesenchymal stem cells and embryonic fibroblasts. *Mol Biol Rep.* 2018; 45: 1827-1837.
 42. Carnegie JA, Durnford R, Algire J, Morgan J. Evaluation of mitomycin-treated vero cells as a co-culture system for IVM/IVF-derived bovine embryos. *Theriogenology.* 1997; 48: 377-389.
 43. Itoh T, Hoshi H. Efficient isolation and long term viability of bovine small preantral follicles *in vitro*. *Soc in vitro Biol J.* 2000; 36: 235-240.
 44. Packer AL, Hsu TC, Besmer P, Bachvarova RF. The ligand of the c-kit receptor promotes oocyte growth. *Developmental Biol.* 1994; 161: 194-205.
 45. Satoh T, Kobayashi K, Yamashita S, Kikuchi M, Sendai Y, Hoshi H. Tissue inhibitor of metalloproteinase (TIMP-1) produced by granulosa and oviduct cells enhances *in vitro* development of bovine embryo. *Biol. Reprod.* 1994; 50: 835-844.
 46. Vigne JL, Halburnt LL, KM Skinner. Characterization of bovine ovarian surface epithelium and stromal cells: Identification of secreted proteins. *Biol. Reprod.* 1994; 51: 1213-1221.