

Research Article

Comparing Quantitative and Qualitative Methods for Detecting the *In Vitro* Activity of Colistin against Different Gram-Negative Bacilli

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Abstract

The surge in the prevalence of Multidrug-Resistant (MDR) Gram-negative bacterial infections with limited treatment led to colistin reusing to treat MDR infections. This study aimed to determine economical, simple, and reliable colistin susceptibility testing methods as an alternative to the microdilution technique. We compared seven colistin susceptibility testing methods, including quantitative and qualitative, namely: Disk diffusion, E-test, ComASPTM SensiTest Colistin, Colistin broth disk elution, and colistin agar test CHROMagar™ COL-APSE, and BD Phoenix ID/AST automated identification and susceptibility testing system to the gold standard Broth Microdilution (BMD). Whole-genome sequencing was performed on all isolates to determine if the genetic resistant factors affect the phenotypic profile of the colistin resistance. Our results revealed that disk diffusion is still an ineffective method for measuring colistin susceptibility in Gram-negative Bacilli with the highest major error (31.75%), the lowest Kappa 0 (0%), and categorical agreement (68.25%) values. Phoenix, and CompASPTM SensiTest colistin methods have remained superior in reproducibility, sturdiness, and simplicity of use, similar to the currently recommended broth microdilution procedure; with high sensitivity of 95.56%, and 97.73%, specificity of 95.24, and 100%, and Kappa values of 0.89 and 0.95, respectively. This study revealed that Phoenix, and ComASPTM SensiTest colistin methods are recommended for routine microbiology laboratories with a large workload.

Keywords: Colistin; Gram-negative bacilli; Resistance; Microdilution; MIC

Introduction

Colistin is a cationic polypeptide antibiotic that belongs to the family polymyxin, including polymyxin B and polymyxin E (colistin). It is the last resort for treating multidrug resistance Gram-negative Bacilli (MDR-GNB), mainly against *Escherichia coli* (*E. coli*), *Klebsiella pneumoniae* (*K. pneumoniae*), and *Pseudomonas aeruginosa* (*P. aeruginosa*) [1,2]. It is synthesized naturally by *Paenibacillus polymyxa* and is rapidly bactericidal to Gram-negative bacteria. Colistin targets lipid A of Lipopolysaccharide (LPS) in the Gram-Negative Bacilli (GNB) outer layer cell membrane. Its central role is to inhibit cell membrane function by increasing its permeability to absorb polymyxins substances. This increase in permeability leads to the destruction of inner macromolecules and cell ions, essential for cell survival [2,3]. However, several MDR-GNB bacteria developed their defense mechanisms against colistin. This resistance development could be either chromosomal or plasmid-mediated. The chromosomal resistance can occur due to mutation/insertion in LPS biosynthesis genes (*lpx*, *pmrA/B*, *mgrB*, and *phoP/Q*). On the other hand, the plasmid-mediated resistance is acquired by the horizontal gene transmission, Encoding Phosphoethanolamine Transferase (*pEtN*) enzymes [4]. As a result, the cell membrane permeability will be reduced, and polymyxin binding will be suppressed [1,5-7].

Natural resistance to colistin has been reported on several bacteria, mainly chromosomal mutations in some Gram-negative

species. However, recently plasmid-mediated colistin resistance emerged in clinically significant Gram-negative bacilli. The emergent resistance was first reported in China in November 2015 through the *mcr-1* gene [8, 9]. Since then, strains carrying the *mcr-1* gene, especially *E. coli*, have been isolated worldwide from humans [10], food-producing animals [11], and the environment [12]. Another plasmid-carried colistin-resistant gene (*mcr-2*) was detected in *E. coli* isolates from pigs in Belgium [13]. To date, ten mobile *mcr* genes have been described [14,15]. Colistin was first introduced as an antibiotic in 1952 and was used until the early 1980s to treat infections caused by GNB. Its use was discontinued because of problems with toxicity, such as nephrotoxicity and neurotoxicity [16]. The increase of multidrug resistance among clinically significant GNB renewed interest in colistin as a last resort therapeutic option for treating MDR-GNB bacteria. In clinical practice, there are some situations where clinician needs to use colistin; therefore, it is critical for clinical laboratories to determine colistin resistance to prevent prolonged administration to the patient for whom it would not be effective.

The poor diffusion and binding of colistin to plastics lead to its complicated phenotypic resistance testing [17]. In addition, some studies noted discrepancies in phenotypic colistin resistance results obtained by different methods. Besides, no reference method has been demarcated to compare colistin susceptibility testing results [4]. As such, both the Clinical and Laboratory Standards Institute (CLSI) and the European Committee on Antimicrobial Susceptibility

Testing (EUCAST) recommend only Broth Microdilution (BMD) [17]. However, the method is impractical for clinical laboratories due to the large sample load in routine microbiology laboratories [18]. Thus, an alternative, more practical method is needed. Several commercial methods are available for colistin Minimum Inhibitory Concentration (MIC) measurement, but only a few studies describe their efficacy compared to each other. In this context, we conducted this study to evaluate and compare the different available quantitative and qualitative colistin susceptibility testing methods on some GNB to find the feasible colistin test. These methods include disk diffusion (Kirby- Bauer test), gradient diffusion method (E- test), CHROMagar™ COL-APSE, ComASPTM colistin broth micro-dilution, colistin broth disk elution, agar dilution, BD Phoenix ID/AST automated identification, and susceptibility testing system.

Additionally, we performed Whole-Genome Sequencing (WGS) to identify the genetic determinants of colistin resistance and find if it affects the phenotypic testing method. The study outcomes will provide a framework for clinical laboratories to select the appropriate method to test colistin resistance.

Materials and Methods

Bacterial strains

Two sets of previously collected bacterial isolates (n=63) were employed in this study. The first set comprises isolated human pathogens collected from Hamad Medical Corporation (HMC), Doha, Qatar, during routine diagnostic testing (n=37). The second set was obtained from poultry fecal samples (n=26). Non-duplicate bacterial strains were characterized and identified as GNB *via* Biomic V 3 (Giles Scientific, USA) or Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry (MAL-DI-TOF MS) (Bruker Daltonik GmbH, Leipzig, Germany). These isolates include 43 strains of *E. coli*, 18 strains of *K. Pneumoniae*, and two strains of *P. aeruginosa*. Clinical Isolates were selected because of their resistance to third-generation cephalosporins and/or colistin. *E. coli* isolates from poultry fecal samples were obtained from farms using colistin as a prophylaxis and growth promoter. The reference strains *E. coli* ATCC[®] 25922, *K. pneumoniae* ATCC[®] 700603, and *P. aeruginosa* ATCC[®] 27853 were used as quality control organisms in all experiments. This study was approved by Qatar University Institutional Biosafety Committee (QU-IBC) number QU-IBC-2020/030.

Phenotypic methods to detect colistin resistance

The subsequent techniques were applied to all 63 isolates to determine their susceptibility to colistin, and readings were interpreted according to the CLSI [19], NCCLS [20], and Gales et al. [21] (Supplementary Table S1).

Disk diffusion method (Kirby-Bauer test)

Bacterial isolates were cultured on nutrient agar plates for 18-24hrs at 37°C. After incubation, 2 to 3 pure colonies were suspended in a phosphate buffer solution (Atom Scientific, UK) to achieve an inoculum equivalent to 0.5 McFarland Standard as measured by DensiCHEK PLUS (bioMérieux, France). The suspension was then swabbed onto a Mueller-Hinton (MH) agar plate (Himedia, Mumbai, India) and allowed to dry completely. Next, antibiotic disks impregnated with 10µg colistin (Liofilchem[®], Roseto Degli Abruzzi, Italy) were applied to the agar surface and incubated at 37°C for

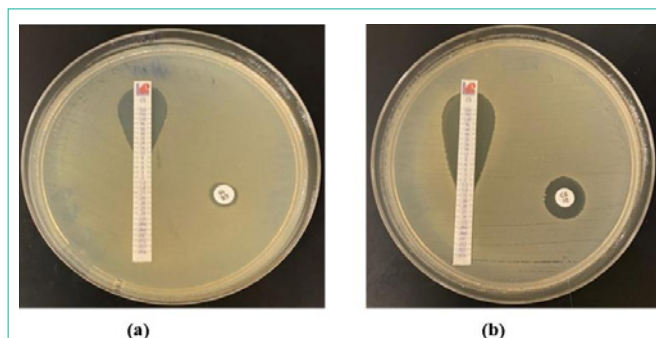


Figure 1: Colistin susceptibility testing by the disk diffusion and E-test methods (Liofilchem[®]). (a) The isolate's disk diffusion test showed an inhibition zone of 8 mm around the colistin disk (right) and MIC of 12µg/mL (left). (b) disk diffusion test of the isolate showing inhibition zone of 12 mm around the colistin disk (right), inhibition zone with MIC of 1.5 µg/mL (left).

24hrs. The zone of inhibition was measured in mm (Figure 1). These values were interpreted according to the CLSI [19].

Gradient diffusion method (E-Test)

The isolates were processed similarly to the disk diffusion. The E-test strip (Liofilchem, Zona Ind. le, Italy), containing a gradient of antibiotic concentrations, was placed onto the prepared MH agar plate. The next day, the MIC (the point where the edge of the ellipse intersects the strip) in µg/mL was recorded (Figure 1), and susceptibility was interpreted according to CLSI [19].

CHROMagar™ COL-APSE (selective colistin bacterial culture medium)

The CHROMagar™ COL-APSE (CHROMagar, Paris, France) medium was briefly prepared in-house according to the manufacturer's recommendation. The medium was challenged with 63 different GNB recovered from human and animal samples. The media's ability to detect colistin Gram-negative resistant isolates was assessed by direct streaking of a single colony, taken from overnight nutrient agar culture onto the prepared CHROMagar plate. Then the plate was incubated at 37°C for 18-24 hrs. The growth of typical single colonies appearance, *E. coli* (pink), *Pseudomonas* (natural pigmentation cream to green), *coliforms* (metallic blue) was considered as colistin-resistant (Figure 2). If the growth was inhibited, the isolate was considered sensitive.

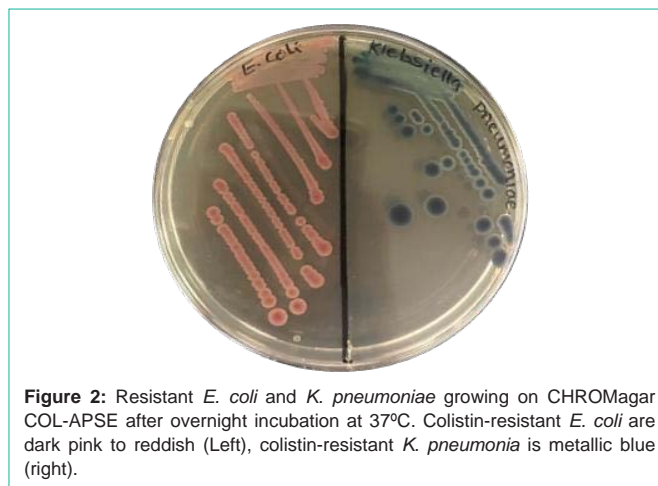


Figure 2: Resistant *E. coli* and *K. pneumoniae* growing on CHROMagar COL-APSE after overnight incubation at 37°C. Colistin-resistant *E. coli* are dark pink to reddish (Left), colistin-resistant *K. pneumoniae* is metallic blue (right).

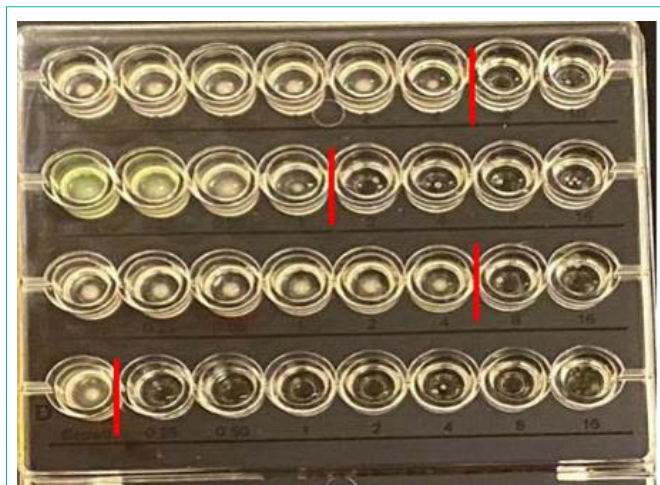


Figure 3: Colistin susceptibility testing by ComASP. Concentrations in the ComASPTM colistin plate range from 0.25µg/mL to 16µg/mL. The well on the right of the red line indicates the value of the MIC for the isolate.

ComASPTM SensiTest Colistin

ComASPTM Colistin (Liofilchem, Italy) was used as a simple version of the BMD and following the manufacturer's recommendations. Succinctly, suspensions equivalent to the 0.5 McFarland of the isolates were prepared. Further dilutions were obtained by diluting 50µL of the bacterial suspensions in 950µL of saline. Then 100µL of the diluted suspension was dispensed in the panel's wells. The panel was covered with a lid and incubated in ambient air at 37°C for 16-20 hrs. The first clear well was considered the MIC of the examined sample. Concentrations in the ComASPTM colistin plate range from 0.25µg/mL to 16µg/mL as depicted in Figure 3.

Colistin broth disk elution

This method was performed as described by CLSI [19]. Three to five colonies were picked, and turbidity was adjusted to the equivalent of a 0.5 McFarland standard, then further diluted to 1:10 in PBS. Four tubes of MH Broth (Liofilchem, Roseto degli Abruzzi, Italy) were labelled 0 (for control), 1, 2 and 4µg/mL for each isolate, corresponding to the concentration of colistin disks (Liofilchem, Roseto degli Abruzzi, Italy) that contains. One colistin disk was added to the tube labeled (1µg/mL), two colistin disks to tube labeled (2µg/mL), and four colistin disks to tube labeled (4µg/mL). The prepared inoculum (50µL) was added to all tubes to attain a final concentration of approximately 7.5×10^5 CFU/mL. The tubes were incubated in a shaking incubator (Innova'44, incubator shaker series, Eppendorf, Germany) at 33-35°C in ambient air for 16-20 hrs. MIC was read as the lowest agar concentration that completely inhibits the growth of the tested isolate (Figure 4).

Colistin agar test (CAT)

Colistin stock antibiotic solution was made by adding 15.7mg of colistin sulfate salt powder (Potency 19000 U/mg) to 1mL of distilled water to obtain a final concentration of 10mg/mL, according to the following equation: $\text{Weight (mg)} = \text{Volume (mL)} \times \text{colistin concentration (}\mu\text{g/mL)} / \text{potency (}\mu\text{g/mg)}$ [22]. Colistin agar plates with different colistin concentrations were prepared as follows: the autoclaved MH agar was allowed to cool to 45-50°C in a water

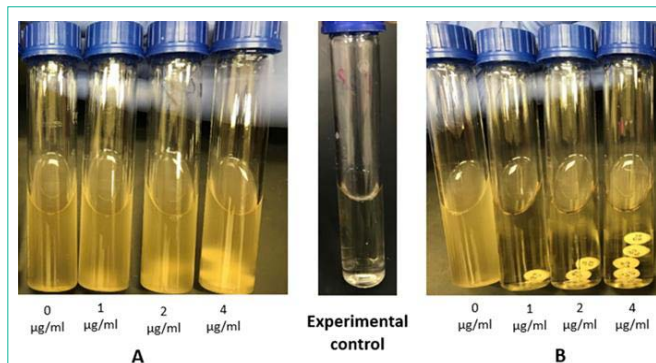


Figure 4: Colistin Broth-Disk Elution (CBDE). (A) Resistance, with a colistin MIC of >4µg/mL (B) Sensitive, with colistin MIC of 1µg/mL.



Figure 5: Colistin agar dilution test indicates the MIC for five isolates and a susceptible Control (C) in four different concentrations. A) 0µg/mL, B) 1µg/mL, C) 2µg/mL and D) 4µg/mL. Inhibition of the growth indicating the MIC of the isolates. The plates results: C = susceptible control, 1 = resistance, 2 = resistance, 3 = sensitive, "4 = sensitive," 5 = resistance.

bath before aseptically adding colistin (15µL, 30µL, and 60µL to 150mL warm MH agar to achieve concentrations of 1, 2, and 4µg/mL, respectively. Subsequently, the prepared MH agar was poured into plates labeled with appropriate concentrations. After agar solidification, each colistin agar plate of specific concentration was divided into six parts labeled with a marker to test up to 6 isolates per plate. Then 10µL of ten-fold dilution of McFarland inoculum, which was prepared as mentioned above, were streaked onto the appropriate portion of each colistin agar plate and incubated at 33-35°C in ambient air for 16-20 hrs. MIC was read as the lowest agar concentration that completely inhibits the tested isolate's growth (Figure 5).

Micro-broth dilution (BMD)

This method involves using small volumes of MH broth distributed in sterile, round-bottom 96-plastic well plates described by [22,23]. Briefly, two-fold colistin serial dilutions (50µL each), starting with the lowest concentration of 0.125µg/mL to second-highest concentration up to 250µg/mL, were added to the wells in each row. Thus, one plate contains seven samples, including *E. coli* ATCC 25922 as a quality control organism, while the last 8th row contains each tested isolate's growth control (Figure 6). 50µL of the standardized inoculum (prepared as described above) was added to each well within 15 minutes of its preparation. The inoculated 96 well plates were incubated at 35±2°C for 16-18 hrs. MIC was recorded as the lowest colistin concentration that completely inhibits bacteria growth as detected by the unaided eye.

BD Phoenix ID/AST automated identification and susceptibility testing system

BD Phoenix™ M50 (New Jersey, US) was used to determine

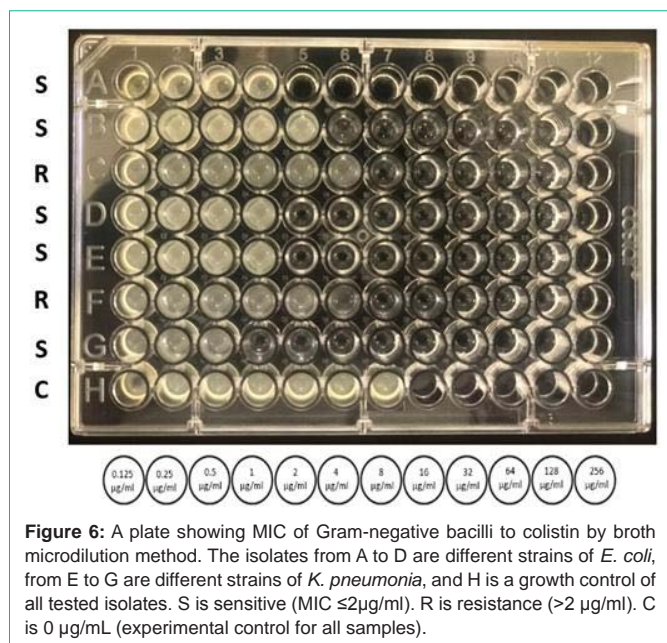


Figure 6: A plate showing MIC of Gram-negative bacilli to colistin by broth microdilution method. The isolates from A to D are different strains of *E. coli*, from E to G are different strains of *K. pneumoniae*, and H is a growth control of all tested isolates. S is sensitive (MIC $\leq 2\mu\text{g/ml}$). R is resistance ($>2\mu\text{g/ml}$). C is 0 $\mu\text{g/ml}$ (experimental control for all samples).

Antibiotic Susceptibility Testing (AST) following the manufacturer’s recommendations. Briefly, few overnight bacterial colonies were suspended in the provided BD phoenix broth to achieve an inoculum equivalent to 0.5 McFarland Standard. Next 25 μl of this prepared inoculum solution was transferred to a tube containing Phoenix AST broth and a drop of the AST indicator solution. After sample preparation was completed, 4.5 mL was added to the BD phoenix Gram-negative panel. This panel has a wide range of two-fold doubling dilution of several antibiotics (including colistin) and one growth control well. The result was read within 16-18 hrs. The Phoenix system measures the MIC by microdilution and utilizes a redox indicator to detect organism growth in the presence of an antimicrobial agent. Organism identification is used in the interpretation of the MIC values of each antimicrobial agent.

DNA extraction, sequencing, and bioinformatics

According to the manufacturer’s protocol, genetic DNA was extracted from all isolates using a QIAamp[®] UCP Pathogen mini kit (Qiagen, Düsseldorf, Germany). Each isolate’s genomic DNA was purified and quantified using a Qubit dsDNA high sensitivity assay (Thermo Fisher, Waltham, United States). Then DNA was sent to BGI Genomics Co., Ltd., China, for whole-genome sequencing. The received raw sequence reads were subjected to a quality check and analyzed using CLC genomics workbench v20.0.4 (<https://digitalinsights.qiagen.com>). Briefly, the reads were quality assessed, trimmed, followed by de novo assembly. Colistin resistance genes were identified using ResFinder v 4.1 [24] and the comprehensive

Table 1: Colistin MIC distributions by broth microdilution for the 63 Gram-negative Bacilli isolates.

Organism	Number of isolates	Colistin MIC ($\mu\text{g/ml}$)									
		0.25	0.5	1	2	4	8	16	32	64	128
<i>E. coli</i>	43			5	10	3	15	9	1		
<i>K. pneumoniae</i>	18			1	4	1	1	1	3	4	3
<i>P. aeruginosa</i>	2					1	1				
Total	63	0	0	6	14	5	17	10	4	4	3

antibiotic resistance database CARD [25].

Data analysis

The MIC and breakpoint results obtained from the examined methods were interpreted according to CLSI [19], NCCLS [20], Gales, et al. [21] guidelines and then compared to those obtained by the BMD reference method. A Very Major Error (VME) is defined as the error indicating a false susceptible result in the tested method and resistance in the BMD method’s reference method. In contrast, a Major Error (ME) denoted a false resistant result in the tested method and susceptible in the reference method. Categorical Agreement (CA) percentage is the percentage of isolates with the same results in both reference and test methods. Sensitivity, which measures the correctly identified positive portion, is characterized as true positive. On the other hand, specificity measures the correctly identified negative portion, characterized as true negative [26]. Cohen’s Kappa (CK) statistics measure inter-rater agreement for categorical items to check the test reliability. According to CK interpretation [27], the agreement level is none if Kappa value ranges between 0-0.20, minimal; if Kappa value (0.21-0.39), weak; if Kappa value (0.40-0.59), moderate; if Kappa value (0.60-0.79), strong; if Kappa value (0.80-0.9), and almost perfect if the Kappa value is above 0.90. The Positive Predictive Value (PPV) is the proportion of bacterial strains giving positive resistant test results, which are true resistant. On the other hand, the Negative Predictive Value (NPV) is the proportion of bacterial strains giving negative resistant test results, which are true sensitive as measured by BMD [26]. The sensitivity, specificity, PPV, NPV, CA, ME, VME, and CK of each test were calculated using the Equations in Supplementary Table S2, based on the BMD reference method.

Results

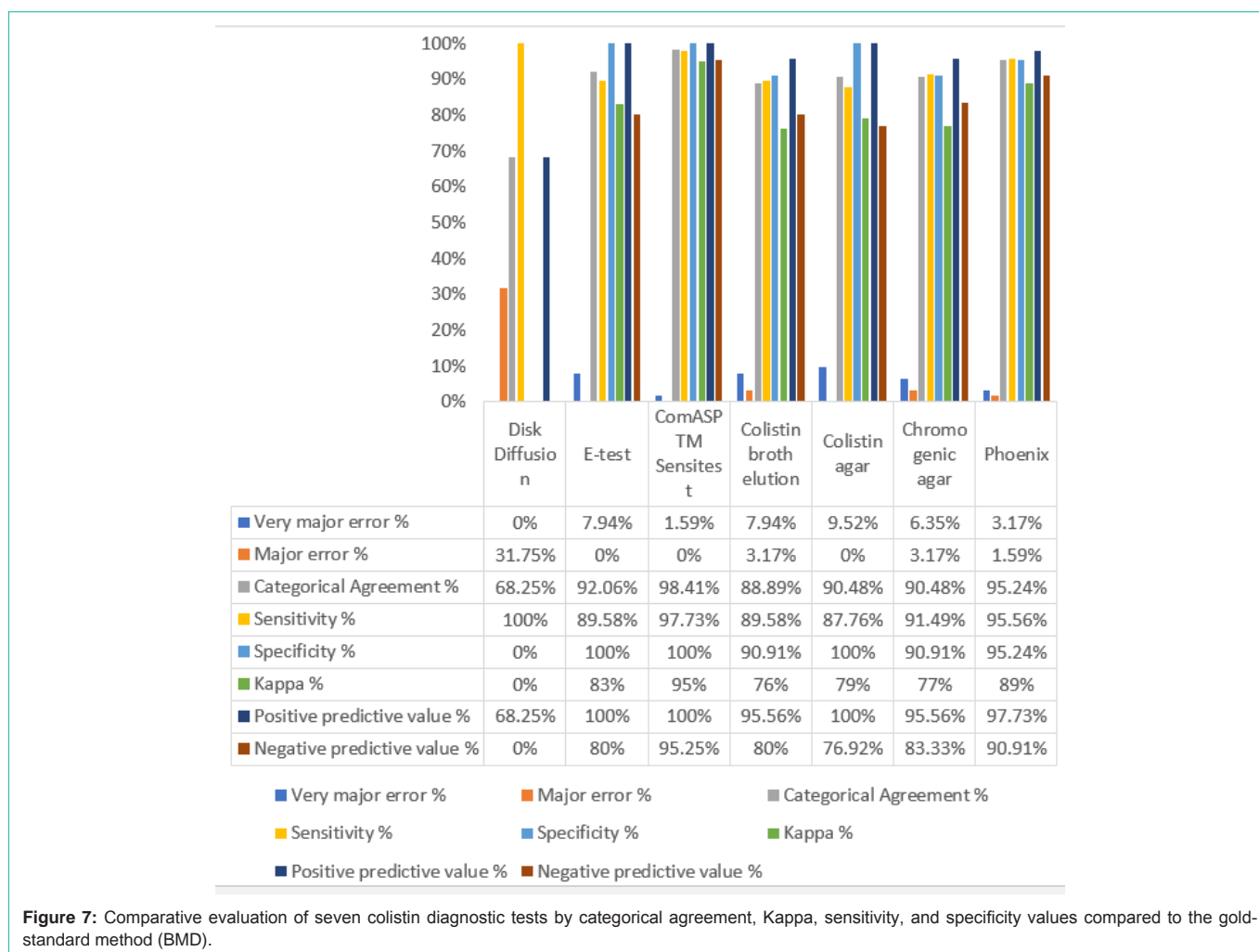
Overall, 43 (68%) of the studied Gram-negative isolates were resistant to colistin by gold standard Broth Microdilution (BMD) with resistance detected in 13 (72%) of *K. pneumoniae*, 2 (100%) *P. aeruginosa*, and 28 (65%) *E. coli* isolates (Table 1). Quality control findings were in line with published standards [19].

Comparison of Colistin disk diffusion to the reference BMD method

In all studied isolates, 95.24% (n=60) of the isolates were identified as resistant with disk diffusion compared to 68.25% (n=43) using the reference method BMD (Table 2, Supplementary Figure 1S(A)). The Very Major Error (VME) was not detected 0% (n=0), while 20 isolates showed 31.75% Major Error (ME) (Figure 7). Based on the isolates tested, the disk diffusion had a sensitivity of 100% and specificity of 0% for detecting colistin resistance with a Positive Predictive Value (PPV) of 68.25% (n=43) and a Negative Predictive Value (NPV) 0% (n=0). There was a 63.49% categorical agreement between the disk diffusion test and the BMD method. At the same time, the Kappa

Table 2: BMD, Disk, E-test, ComASPTM Sensitest Colistin, Colistin Broth Disk Elution, Colistin Agar, and Phoenix susceptibility results.

Test	Number/ percentage of resistance isolates (R)	Number/ percentage of susceptible isolates (S)
BMD	43 (68.25 %)	20 (31.75%)
Disk	60 (95.24%)	3 (4.76%)
E-test	38 (60.32%)	25 (39.68%)
ComASPTM Sensitest Colistin	42 (66.67%)	21 (33.33%)
Colistin Broth Disk Elution	40 (63.49%)	23 (36.51%)
Colistin Agar	37 (58.73%)	26 (41.27%)
Phoenix	42 (66.67%)	21 (33.33%)



value was 0 (0%) (minimal agreement).

Comparison of E-test to the reference BMD method

There was a 92.06% categorical agreement between the E-test and the gold standard method BMD. E-test showed 60.3% (n=38) colistin-resistant with MICs of >4 µg/mL, whereas 39.68% (n=25) were colistin susceptible with MIC of ≤2 µg/mL. These results were compared to the reference method BMD (Table 2, Supplementary Figure 1S(B)). The test had 7.94% (n=5) VME and no ME 0% (Figure 7). The E-test had a sensitivity of 89.58% and a specificity of 100% in detecting colistin resistance, a positive predictive value of 100%, and

a negative predictive value of 80%. The level of agreement by Kappa statistics was 0.83 (83%) (strong agreement).

Comparison of ComASPTM sensitest colistin to reference BMD method

66.67% (n=42) from isolates were colistin-resistant, and 33.33% (n=21) were colistin susceptible using ComASPTM Sensitest colistin resistance detection method (Table 2, Supplementary Figure 1S(c)). The VME was 1.59% (n=1) while ME was 0% (Figure 7). The ComASPTM Sensitest Colistin had a sensitivity of 97.73% and specificity of 100% in detecting bacterial resistance to colistin with

Table 3: Chromogenic agar susceptibility results.

Bacteria	Resistance (Growth) No. /percentage	Susceptible (No growth) No. /percentage
<i>E. coli</i>	26 (41.27%)	17 (26.98%)
<i>K. pneumoniae</i>	13 (20.63%)	5 (7.94%)
<i>P. aeruginosa</i>	2 (3.17%)	0 (0%)

PPV of 100% and NPV of 95.24%. There was a CA 98.4% (n=62) between BMD and ComASPTM Sensitest colistin. (n=62). The Kappa value was 0.95 (95%), which shows an almost perfect agreement between the two methods.

Comparison of colistin broth disk elution to the reference BMD method

A total of 40 (63.49%) of the studied isolates were colistin-resistant, and 23 (36.51%) were colistin susceptible, as shown in Table 2 and Supplementary Figure 1S(D). There was CA between Colistin Broth Disk Elution and BMD of 88.89% (n=56) (Figure 7). Five (7.94%) isolates were VME and 2 (3.17%) ME. The Colistin Broth Disk Elution test had a sensitivity of 89.58% and specificity of 90.91%. PPV of isolates was 95.56%, and NPV was 80%. This method had a Kappa value of 0.76 (76%), which shows a moderate level of agreement.

Comparison of Agar Dilution to reference BMD method

Overall, 58.7% (n=37) were colistin-resistant with MICs of >2.0 µg/mL. In comparison, 41.27% (n=26) were susceptible, as presented in (Table 2) and (Supplementary Figure 1S(E)). 9.52% (n= 6) of the isolates showed VME, and no ME was detected (Figure 7). The categorical agreement was 90.48%, and kappa statistics was 0.79 (79%). This test had a sensitivity of 87.76% and a specificity of 100%, with a 100% Positive Predictive Value (PPV) and 76.92% Negative Predictive Value (NPV).

Comparison of chromogenic agar to the reference BMD method

Chromogenic agar enhanced the growth of 41(65.08%) of isolates; *E. coli* = 26, *K. pneumoniae* = 13 and *P. aeruginosa* = 2. While it suppressed the growth of 22 (34.92%), *E. coli* = 17, and *K. pneumoniae* (Table 3, Supplementary Figure 1S(F)). The method had a sensitivity of 91.49% and a specificity of 90.91% (Figure 7). The CA between chromo-genic agar and BMD method was 90.48%. There were 4 (6.35%) VME and 2 (3.17%) ME. PPV was 95.56%, and NPV was 83.33%. Kappa value for chromogenic agar method showed a moderate level of agreement (0.77, 77%).

Comparison of Phoenix to reference BMD method

A total of 42 (66.67%) isolates were colistin-resistant, while 21 (33.33%) were colistin susceptible by Phoenix (Table 2, Supplementary Figure 1S(G)). There was a CA of 95.24% (n=60) between Phoenix

and BMD (Figure 7). The VME was 3.17% (n=2), and the ME was 1.59% (n=1). Phoenix method had 95.56% sensitivity and 95.24% specificity, with PPV of 97.73% and NPV of 90.91%. The Kappa value was 0.89 (89%), indicating a strong level of agreement.

Genetic determinant of colistin resistance

As depicted in Table 4, a total of 43 isolates of *E. coli* were divided into four types based on genetic determinants of colistin resistance. Of these, 26 were plasmid resistance (harbor *mcr1-1*); 2 were chromosomal modifications; one isolate harbor *mcr1-1* plus chromosomal modification, and 14 were sensitive, not presenting any colistin-resistant genetic determinant. Whereas *K. pneumoniae* has 11 isolates that contain chromosomal colistin resistance, 2 isolates harbored *mcr1-1* plus chromosomal modifications, and 5 were sensitive. The resistance of the two isolates of *P. aeruginosa* was due to the chromosomal modifications.

Discussion

The use of colistin to treat infections caused by multidrug-resistant Gram-negative Bacilli, including *E. coli*, *K. pneumoniae*, and *P. aeruginosa*, is being increasingly described in many countries [28]. Additionally, antimicrobial susceptibility by phenotypic methods, including the colistin method, is currently the cornerstone of Antimicrobial Resistance (AMR) surveillance [17]. Nevertheless, susceptibility testing for colistin remains challenging mainly due to its intrinsic properties. Importantly, there is no agreement between colistin susceptibility testing methodologies [29,30]. The technique most frequently used in routine laboratories is disc diffusion, but it has been shown to be inefficient [17,29,31]. This study tested colistin resistance using seven methods and compared them to BMD as a gold standard, which is the most reliable method [17,19]. However, this method consumes time, effort, and money in routine medical laboratories [18]. Furthermore, the inclusion of representative strains, both susceptible and resistant, is critical in avoiding bias against a positive or negative outcome. Therefore, the strain panel used in this study was selected to ensure that both resistant and sensitive strains are included.

The most widely used method in routine laboratories, disk diffusion, is inefficient; due to weak colistin diffusion in Mueller Hinton agar [32]. Based on our findings, the disk diffusion method using a 10 µg/mL colistin disk was not a reliable method for colistin susceptibility testing with specificity 0% and Kappa value of 0 (0%) (Figure 7). These findings agree with several studies [31,33,34] reporting an inaccurate disk method for determining colistin susceptibility. In addition, an unacceptably high rate of very major errors ranging from 5% to 11% was recorded [31,33,34]. In our study, we did not report any very major errors (0%); however, we observed a high percentage of major errors (31.75%) with a low categorical

Table 4: Genetic determinant of colistin resistance among 63 Gram-negative Bacilli isolates.

Organism	Number of isolates	Colistin resistance type			
		<i>mcr1-1</i>	GMR	<i>mcr1-1</i> & GMR	Not present
<i>E. coli</i>	43	26	2	1	14
<i>K. pneumoniae</i>	18	0	11	2	5
<i>P. aeruginosa</i>	2	0	2	0	0
Total	63	26	15	3	19

agreement of 68.25% to the reference BMD method that supports the inaccuracy described by the previous studies.

Lo-Ten-Foe et al. [33]; Tan and Ng. [35]; Behera et al. [36] reported that E-test is a fast and reliable alternative method for determining colistin susceptibility. Our comparison portrayed a strong correlation (Kappa, 83% and Categorical agreement 92%) between broth microdilution and E-test. No false resistant (ME) results by E-test were detected, whereas a 7.94% false susceptible (VME) was exhibited (Figure 7). The E-test has been shown to have high concordance (>96%) in correlation with broth microdilution in previous studies [33,36]. In convergence with our findings, these studies reported one to four percentages of false susceptibility (VME) but no false resistant results (ME).

ComASPTM Sensitest Colistin is a modified BMD method but more straightforward with a shorter turnaround time, as different colistin concentrations are already prepared and exist in the test panels. The outcomes of this method were the most perfect compared to all other tested methods. Our data demonstrated a categorical agreement of 98.4% compared to BMD. Only one isolate was colistin-resistant by BMD and susceptible by ComASPTM Sensitest, resulting in a VME value of 1.59%, and no ME was documented (Figure 7). The method has a high specificity of 100%, whereas sensitivity was 97.73%. A PPV and NPV of 100% and 95.24% were recorded, respectively. The significantly high Kappa value (95%) of ComASPTM Sensitest proposed that this method can be an alternative to BMD. Thus, ComASPTM Sensitest can be accomplished faster with less effort and required accuracy in large laboratories with a high workload. Our findings were concordant with Osei Sekyere, et al. [37]; Carretto et al. [1].

Colistin broth disk elution endorsed by [19] has low cost and needs less effort. According to Simner et al. [38], colistin broth disk elution is a reproducible and accurate method and can alter BMD. Nevertheless, our test results revealed that the Categorical agreement between colistin broth disk elution and BMD is 88.89%. Five isolates with VME and 2 with ME. The method had a moderate agreement (Kappa value of (76%) compared to BMD. The dissimilarity in results could be ascribed to the use of disks produced by different manufacturers.

Colistin agar test was recently introduced to be superior to broth dilution methods in reproducibility and robustness [39] for determining colistin MICs. CLSI [19] lately recommended CAT, an adaptation of the agar dilution process, as a test for colistin resistance in *Enterobacteriales* and *P. aeruginosa*. No ME was observed with the agar dilution method; however, 9.52% VME was detected mainly among *E. coli* isolates. Gonzales Escalante et al. [39] suggested adding an extra plate of colistin supplemented with EDTA, a well-known *mcr1-1* inhibitor, to the CAT method; this was recently implemented for the precise detection of *mcr1-1* producers. Numerous studies have demonstrated excellent agreement between colistin agar dilution and BMD [40,41]. Our study depicted a solid categorical agreement (90.48%) between the agar dilution method and the gold standard BMD (Figure 7).

In this study, we also used CHROMagar™ COL-APSE chromogenic agar to detect colistin resistance. Our numbers show specificity and sensitivity of 91.49% and 90.91%, respectively, whereas

a recent study by Sekyere et al. [37] recorded a sensitivity and specificity of 82.05% and 66.67%. On the other hand, Abdul Momin et al. [4] reported specificity and sensitivity of 100% in their study. Sekyere et al. [37] evaluated the media's performance using different amounts of cultured bacteria; this might be a constraint in our study that needs further investigation. It is remarkable to note that this media suppresses the growth of susceptible strains, but some grew on it, as demonstrated by our study and the previous studies. The CA was 90.48%, where six isolates had different results by the chromogenic agar and BMD. Four isolates showed VME, and two displayed ME. The Kappa value has a moderate level of agreement with 0.77 (77%). Therefore, it reduces the chance of choosing chromogenic agar as an alternative to BMD.

BD Phoenix™ M50. This method showed a CA of 95.24%, VME of 3.17%, and ME of 1.59%. Thus, the sensitivity and the specificity had almost the same values of 95.56% and 95.24%. In addition, the method had a Kappa value of 0.89 (89%), indicating a solid agreement level. In contrast, previous studies showed that Phoenix is a deprived method and has unacceptable and inaccurate results with low specificity and sensitivity [42,43]. Furthermore, a study done by Jayol, et al. [44] mentioned that the Phoenix system could detect plasmid-mediated colistin-resistance bacteria. Conversely, our study results indicated that Phoenix could detect both plasmid and gene-mediated colistin resistance, regardless of the resistance level.

To date, ten mobile *mcr* genes have been described [9,14,15]. In our study, we did Whole-Genome Sequencing (WGS) for 63 isolates. A high resistance rate to colistin antibiotic was mainly detected among *E. coli* isolates that harbor *mcr1-1* type of genetic determinant of colistin resistance. While most of *K. pneumoniae* isolates had chromosomal colistin resistance mutation, as demonstrated by Table 4. Genotypic methods, in particular, are unlikely to detect all of the chromosomal mutations responsible for the majority of phenotypic colistin resistance in clinical screening settings [45]. Therefore, a negative PCR molecular test result cannot be used to predict colistin susceptibility because the test cannot rule out the existence of chromosomal mechanisms of resistance or even the presence of a novel *mcr* gene. To avoid this limitation, *mcr* genes and identified chromosomal mutations that cause colistin resistance will be possible with WGS. Although strains with novel resistance mechanisms will reduce the sensitivity and negative predictive value, WGS is the most detailed approach for detecting all currently known putative colistin resistance mechanisms. As new resistance mechanisms are described, it will also allow retrospective analysis of sequencing results. The agreement between phenotypic and genotypic test results will increase our understanding of colistin resistance mechanisms.

Conclusion

In conclusion, as we continue to spectate an increasing emergence of colistin resistance in Gram-negative multidrug-resistant bacteria, reliable and rapid colistin susceptibility testing is anticipated. This study found that disk diffusion is still an ineffective method for measuring colistin susceptibility in Gram-negative Bacilli. On the other hand, Phoenix and CompASPT SensiTest colistin methods have remained superior in reproducibility, sturdiness, and simplicity of use with a performance similar to the current recommended BMD procedure. These findings may rationalize and recommend validating

these approaches using more samples and including other Gram-negative bacilli to develop a globally accepted consistent method for colistin susceptibility testing. If standardized, these methods can be executed in laboratories with a high workload and colistin resistance surveillance programs. However, our study did not ensure any relation between the type of colistin resistance genetic determinant (chromosomal/plasmid-mediated) and the performance of the specific colistin susceptibility test.

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Author Contributions

Conceptualization, N.E. methodology, N.S., H.A., N.M., M.R., K.A., K.A.; software, N.E, N.S and H.A.; validation, N.S. and H.A.; formal analysis, N.E.; investigation, N.E and S.Z. ; resources, N.E.; data curation, N.E and S.Z.; writing-original draft preparation, N.S and H. A.; writing-review and editing, N.E. and S. Z; visualization, N.E and S. Z.; supervision, N.E.; project administration, N.E.; funding acquisition, N.E. All authors have read and agreed to the published version of the manuscript”.

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