

Research Article

Effects of a Quaternary Lidocaine Derivative, QX-314, on Slow Ventral Root Potentials in Neonatal Rats

Hayakawa C¹, Inagaki K¹, Minoura Y² and Onimaru H^{3*}

¹Department of Orthopedic Surgery, Showa University School of Medicine, Hatanodai, Shinagawa-ku, Tokyo, Japan

²Departments of Medicine Division of Cardiology, Showa University School of Medicine, Hatanodai, Shinagawa-ku, Tokyo, Japan

³Department of Physiology, Showa University School of Medicine, Hatanodai, Shinagawa-ku, Tokyo, Japan

*Corresponding author: Onimaru H, Department of Physiology, Showa University School of Medicine, Hatanodai, Shinagawa-ku, Tokyo, Japan

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Abstract

The use of QX-314 (a quaternary derivative of lidocaine) in the clinical setting has been proposed to achieve the selective inhibition of nociceptors that express transient receptor potential vanilloid 1 (TRPV1) channels with fewer motor deficits. It has been established that slow ventral root potential induced by ipsilateral dorsal root stimulation in the isolated (typically lumbar) spinal cord of newborn rats reflects the nociceptive response. This *in vitro* experimental model is useful for assessing the actions of analgesics. However, there is no report regarding the effects of QX-314 and the combined application of capsaicin in the *in vitro* experimental model. Therefore, we examined the effects of extracellularly applied QX-314 and capsaicin on the spinal reflex response in spinal cord preparations from newborn rats. We found that co-application of QX-314 (1mM) and capsaicin (10 μ M) caused significant attenuation of the reflex response that continued after washout. We suggest that QX-314 passed into neurons in the reflex arc through TRPV1 channels activated by capsaicin. Our report further provides the basic neuronal mechanisms to support the clinical use of QX-314 that seems to be potentially expected for analgesic treatment.

Keywords: QX-314; Nociceptive response; Newborn rat; *in vitro*

Abbreviations

ACSF: Artificial Cerebrospinal Fluid; DRG: Dorsal Root Ganglion; QX-314; Lidocaine N-Ethyl Bromide; TRPV1: Transient Receptor Potential Vanilloid 1

Introduction

A quaternary derivative of lidocaine, QX-314, is thought to be membrane impermeable due to its positive charge and to produce local anesthetic effects when it is intra cellularly administered to neurons [1,2]. Binshtok et al [3] reported that the extracellular application of QX-314 in neurons of Dorsal Root Ganglion (DRG) could block nociceptors when it was applied together with capsaicin (a transient receptor potential vanilloid 1 [TRPV1] agonist) because QX-314 could permeate through the TRPV1 channels that were opened by capsaicin [4-6]. Thus, the clinical use of QX-314 has been proposed to achieve the selective inhibition of nociceptors that express TRPV1 channels with fewer motor deficits [3]. It has been established that the slow ventral root potential induced by ipsilateral dorsal root stimulation in the isolated (typically lumbar) spinal cord of newborn rats reflects the nociceptive reflex. This *in vitro* experimental model is useful for assessing the actions of analgesics [7-9]. Dexmedetomidine and morphine inhibited slow ventral root potentials following ipsilateral dorsal root stimulation in newborn rat spinal cord preparation *in vitro* [10]. We recently reported that lidocaine depressed nociception-related responses at lower concentrations than those that induced respiratory depression [11]. However, there is no report regarding the effects of QX-314 and combined application of capsaicin on nociception-related responses in the above *in vitro* experimental model. Therefore, we examined the effects of extracellularly applied QX-314 and capsaicin on the spinal reflex response in spinal cord preparations from newborn rats.

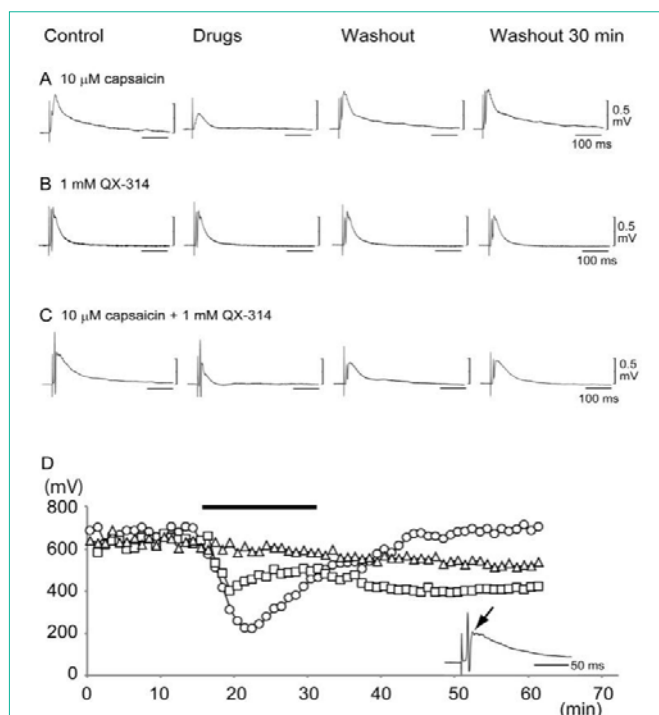


Figure 1: Effects of QX-314 on spinal reflex responses. A, Application of 10 μ M capsaicin. B, Application of 1mM QX-314. C, Co-application of 10 μ M capsaicin and 1 mM QX-314. Control, before drug application; Drug, 4-6 min after drug application; Washout, at 15min after drug application; Washout 30min, after 30min washout of drug. Each recording shows an average from three responses. D, Time course of changes of the amplitude of the slow ventral root potential in the reflex response. Inset, Example of a fast-sweep representation of the reflex response in which the arrow denotes the peak of the slow ventral root potential whose amplitude is plotted.

Materials and Methods

Preparation and solutions

The experimental protocols were approved by the Animal Research Committee of Showa University, which operates in accordance with Law No.105 (of the Japanese Government) for the care and use of laboratory animals. Spinal cord preparations were isolated from Wistar rats (postnatal day 0-3) under deep isoflurane anesthesia. The preparations then were cut transversely between the L1 and L5 nerve roots. Preparations were superfused continuously at 2.5-3 ml/min in a 2ml chamber with artificial cerebrospinal fluid (ACSF) [12] composed of (in mM) 124 NaCl, 5.0 KCl, 1.2 KH₂PO₄, 2.4 CaCl₂, 1.3 MgCl₂, 26 NaHCO₃, and 30 glucose and equilibrated with 95% O₂ and 5% CO₂ at a pH of 7.4. The preparations were maintained at a temperature of 25-26 °C. Lidocaine N-Ethyl Bromide (QX-314) was purchased from Sigma-Aldrich (Tokyo, Japan) and was stocked as a 100mM solution in purified water. Drugs were dissolved with the above-described ACSF and bathapplied.

To evaluate the effects of QX-314 on putative nociceptive responses, the ipsilateral dorsal root of L3 was stimulated using a glass suction electrode, and the induced reflex response was recorded from the L3 ventral root through a 0.5Hz high-pass filter. The dorsal root was stimulated every 60s with a 5-20 V, 200µs square pulse. We measured the peak amplitude of the slow ventral root potentials [10,13]. The preparations were superfused with ACSF for at least 15-30 min until the spinal reflex response became stable. Then, capsaicin and/or QX-314 were bath applied for 15min and washed out.

Data analysis

All data analyses were performed using the Lab Chart 7 Pro software program (AD Instruments, Castle Hill, Australia). The data are presented as the mean and standard deviation for all preparations. The significance of the values was analyzed by one-way ANOVA followed by a Tukey-Kramer multiple comparisons test (Graph Pad In Stat; Graph Pad Software Inc., La Jolla, CA, USA) at a confidence level of $P < 0.05$.

Results and Discussion

Figure 1 shows typical examples of the time course of changes in reflex responses after the application of 10µM capsaicin, 1mM QX-314 or 10µM capsaicin plus 1mM QX-314. Sole application of capsaicin induced a transient decrease of the reflex response followed by recovery even during capsaicin treatment (Figure 1A, open circles in D). Sole application of QX-314 tended to result in a slow and slight (but not significant) decrease of the reflex response (Figure 1B, open triangles). Co-application of capsaicin and QX-314 induced a rapid decrease of the reflex response without recovery (Figure 1C, open squares in D). The group data are summarized in Figure 2. Although a tendency for the reflex response to decrease was observed with the co-application of capsaicin and QX-314 (10 or 100 µM), a significant change was confirmed only in the case of 1mM QX-314 (Figure 1 and Figure 2, Figure 2C). Previous studies have reported that extracellularly applied QX-314 passes into cells through capsaicin-activated TRPV1 channels [3-6]. Our present results were consistent with these previous observations. We have suggested that TRPV1 channels are expressed in cells of the spinal cord [14]. It was thought that QX-314 passed into neurons in the reflex arc through

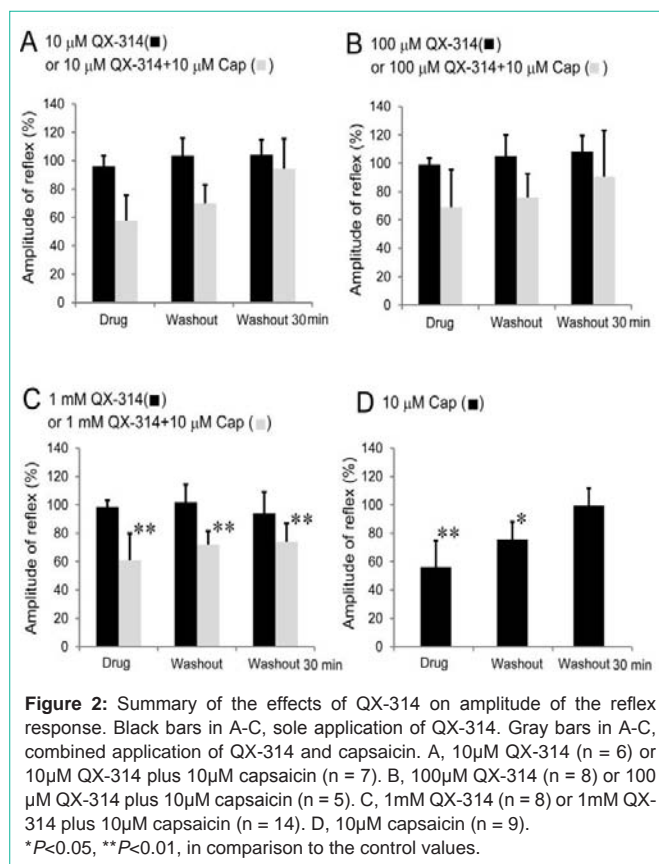


Figure 2: Summary of the effects of QX-314 on amplitude of the reflex response. Black bars in A-C, sole application of QX-314. Gray bars in A-C, combined application of QX-314 and capsaicin. A, 10µM QX-314 (n = 6) or 10µM QX-314 plus 10µM capsaicin (n = 7). B, 100µM QX-314 (n = 8) or 100 µM QX-314 plus 10µM capsaicin (n = 5). C, 1mM QX-314 (n = 8) or 1mM QX-314 plus 10µM capsaicin (n = 14). D, 10µM capsaicin (n = 9). * $P < 0.05$, ** $P < 0.01$, in comparison to the control values.

TRPV1 channels activated by capsaicin. Rivera-Acevedo et al [15] reported that QX-314 exerts biphasic effects on the TRPV1 channels, inhibiting the capsaicin-evoked TRPV1 current at lower (micro molar) concentrations and activating TRPV1 channels at higher (millimolar) concentrations. In the experiments on DRG neurons by Binstok et al [3], 5mM QX-314 was used. Although DRG neurons were not included in the reflex arc of our experiments, our results indicated that 1mM QX-314 was necessary to induce significant effects.

Several reports have shown that the sole application of extracellular QX-314 could affect the action potentials [2,16] and that it produced long-lasting local anesthesia with a slow onset in animal models *in vivo* [17]. We previously reported that the extracellular application of QX-314 induced inhibitory effects on respiratory activity in the brainstem-spinal cord preparation isolated from newborn rats, and the combined application of capsaicin (10 or 100 µM) with QX-314 (100µM) showed no additional effect [18]. Because depressive effects of neuronal activity in the central nervous system could be induced by micro molar concentrations of QX-314 [18,19], such side effects should be considered for the use of QX-314 in clinical cases. Moreover, co-application of capsaicin with QX-314 is not practical in clinical use. Thus, to achieve the selective inhibition of nociceptors that express TRPV1 channels with fewer motor deficits [3], co-application of QX-314 and lidocaine (or bupivacaine) instead of capsaicin has been suggested to be clinically more useful [3,20].

Conclusion

Our report further provides the basic neuronal mechanisms

to support the clinical use of QX-314 that seems to be potentially expected for analgesic treatment, although consideration of the side effects would be necessary.

Acknowledgements

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