

## Research Article

# A Major Recessive Gene Associated with Anthracnose (*Colletotrichum capsici*) Resistance in Chilli Pepper

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Chilli anthracnose caused by *Colletotrichum* sp., is an important disease in Bangladesh and many other Asian countries. Recently the local Bangladeshi genotype 'Comilla-2' was found resistant to *C. capsici*. Inheritance of resistant to *C. capsici* was analyzed in segregating populations derived from a cross of 'BARI chilli-1' x 'Comilla-2'. BARI chilli-1 as susceptible and Comilla-2 as resistant parent was used in the study. Detached matured green fruits were inoculated using the microinjection method. Disease response was evaluated using disease incidence and overall lesion diameter at 8 days after inoculation. The disease reaction of F<sub>1</sub> plants in case of disease incidence and overall lesion diameter were clearly skewed to the susceptible parent, with average values of 46.70% and 13.2 mm, respectively. The distribution of disease incidence in the F<sub>2</sub> population was skewed toward the susceptible parent and the distribution of overall lesion diameter in the F<sub>2</sub> population showed a similar trend. Based on the scale of resistance and susceptibility, less than 25.0% disease incidence or less than 9.0 mm overall lesion diameter were evaluated as resistance. In the cross 'BARI chilli-1' x 'Comilla-2', the segregation ratios of resistance and susceptibility scored by disease incidence and overall lesion diameter in the F<sub>2</sub>, BC<sub>r</sub> and BC<sub>s</sub> populations and chi-squared test significantly fitted one recessive gene model i.e. 1:3 Mendelian model. The result indicates that the resistance of 'Comilla-2' to *C. capsici* is controlled by a single recessive gene.

**Keywords:** Chilli (*Capsicum annum*); anthracnose (*Colletotrichum capsici*); Inheritance.**Introduction**

Chilli (*Capsicum annum* L.) is the most popular species grown worldwide. Anthracnose disease of chilli caused by several *Colletotrichum* spp. is responsible for serious yield loss and affects crop quality in tropical and subtropical regions. The crop is severely infected by anthracnose which causes yield losses up to 50% [1]. *Colletotrichum* species reduces marketable yield of chilli fruits up to 80% [2]. The damage caused by the pathogen is very serious in many Asian countries including Bangladesh. *Colletotrichum capsici* is mainly responsible for chilli fruit anthracnose in Bangladesh. In other countries like Thailand and Indonesia, the primary causal agents of anthracnose are *C. gloeosporioides* and *C. capsici* [3]. *Colletotrichum gloeosporioides* attacks chilli fruits at both the green and red stages, while *C. capsici* mainly attacks the fruits at the red stage [4]. Many fungicides are used to control the disease in developing countries like Bangladesh, but these are not only increase production cost but also cause consumer antipathy. The most economic and environmentally friendly method to control the disease is the use of resistant varieties. Currently, no resistant chilli cultivar is available in Bangladesh against anthracnose. Anthracnose resistant varieties in this species are rarely available [5]. Only one variety of *C. annum*, which was a local Korean variety 'Daepoong-cho' was recently reported to be resistant to *Colletotrichum capsici* [6]. However, study of inheritance of resistant for the development of anthracnose resistant chilli variety is essential.

Several sources of resistance to *C. capsici* have been reported [7,8,3,9,1] and using this genetic resources, researchers have studied the inheritance of anthracnose resistance. The inheritance patterns vary depending on the source of resistance and the *Colletotrichum* isolate. For instance, resistance to *C. dematium* was inherited partially dominantly as reported by [8]. The authors also found that resistance to *C. gloeosporioides* was inherited as over-dominant or partially dominant in F<sub>1</sub> plants [4]. In contrast, some reports have demonstrated that resistance to anthracnose is inherited recessively. For example, [7] found that resistance to *C. capsici* was inherited recessively with epistatic effects and the resistance of *C. chinense* Jacq. PBC932 to *C. capsici* was observed to be inherited through a single recessive gene [1]. The AVRDC has evaluated the resistance to anthracnose of many pepper accessions and has detected several resistance resources [10].

Reported that field resistance against anthracnose is present in Comilla-2 local *Capsicum annum* genotype in Bangladesh [11]. On the other hand, this genotype was also found resistant in the earlier screening experiment. Therefore, the present study was carried out to determine the inheritance of resistance to anthracnose (*C. capsici*) in *Capsicum annum* (Comilla-2).

**Materials and Methods****Plant materials**

Two parents 'Comilla-2' (selected as resistant chilli germplasm) and 'BARI chilli-1-1'(susceptible) were used for inheritance study

of anthracnose resistance to *C. capsici*. F<sub>1</sub> plants were obtained by crossing 'BARI chilli-1' as female and 'Comilla-2' as male parents. F<sub>2</sub> population was obtained by self-pollination of F<sub>1</sub> plants. The backcross populations BC<sub>R</sub> and BC<sub>S</sub> were produced by backcrossing F<sub>1</sub> plants to 'Comilla-2' and 'BARI chilli-1' respectively.

In the cross 'BARI -1' and 'Comilla-2' for the inheritance study, populations consisting of resistant parent (five plants), susceptible parents (five plants), F<sub>1</sub> (five plants), BC<sub>R</sub> (36 plants), BC<sub>S</sub> (10 plants) and F<sub>2</sub> (84 plants) were grown in pot Plant Pathology Department of BSMRAU.

**Fungal isolate**

*Colletotricum capsici* was isolated from infected chilli fruits of BSMRAU experimental farm following standard procedures [12,13]. The isolate was purified following single spore isolation method and identified following a standard key [14,15]. The isolate was maintained on Potato Dextrose Agar (PDA) medium at 25°C in an incubator. After 8 days of incubation, the plates were flooded with distilled water. Conidia were collected by scraping the

culture surface with sterilized glass slides. Density of spore suspension was adjusted to 5x10<sup>5</sup> conidia/ml using a hemacytometer.

**Inoculation**

Healthy fruits of chilli were harvested from individual plants at the matured green and ripen stage. The fruits were washed with distilled water to remove associated microbes from the fruit surface. The fruits punctured with multi pointed needles and inoculated with 10µl of the spore suspension with a micropipette. Each fruit was swabbed with 10µl prepared conidial suspension and raped with poly bags to maintain humidity. Inoculation was conducted in glass petridish with three to five replications. The inoculated fruits were incubated at room temperature for 8 days.

**Disease evaluation**

Severity of disease was evaluated and expressed in percentage of infected sites and overall lesion diameter at 8 days after inoculation (DAI), as described previously [9]. Chi-square goodness of-fit test were used for statistical analysis.

**Results and Discussion**

**Inheritance of resistance to *Colletotrichum capsici* in 'Comilla-2'**

The values of disease incidence and overall lesion diameter of parents and F<sub>1</sub> plants are presented in Table 1. The symptoms of anthracnose started to develop at 2 DAI in susceptible parent 'BARI chilli-1' and there was no further change in disease incidence after 8 DAI. Therefore, the time of disease evaluation for the inheritance study was set at 8 DAI.

The resistant parent 'Comilla-2' and the susceptible parent 'BARI chilli-1' showed the significant differences in disease incidence and overall lesion diameter. The disease incidence and overall lesion diameter were 15.20% and 3.42 mm in resistant parent ('Comilla-2') and 67.10% and 18.74 mm in susceptible parent ('BARI chilli-1'). The disease reactions of F<sub>1</sub> plants in case of disease incidence and overall lesion diameter were clearly skewed to the susceptible parent, with average values of 46.70% and 13.2 mm, respectively. The result

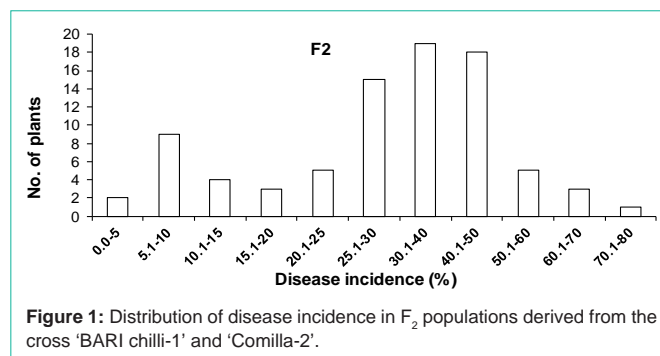


Figure 1: Distribution of disease incidence in F<sub>2</sub> populations derived from the cross 'BARI chilli-1' and 'Comilla-2'.

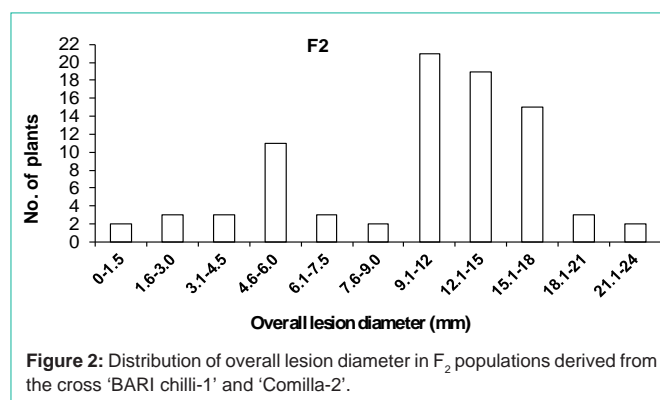


Figure 2: Distribution of overall lesion diameter in F<sub>2</sub> populations derived from the cross 'BARI chilli-1' and 'Comilla-2'.

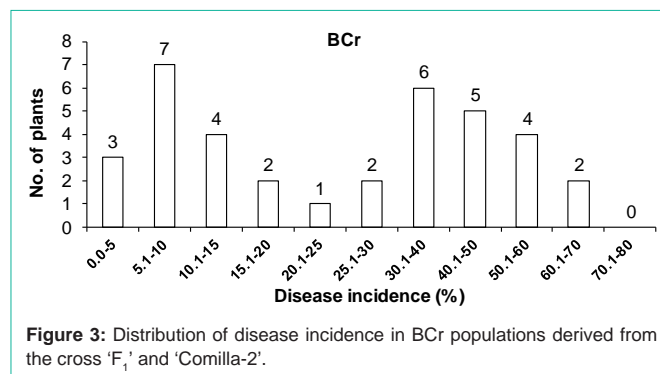
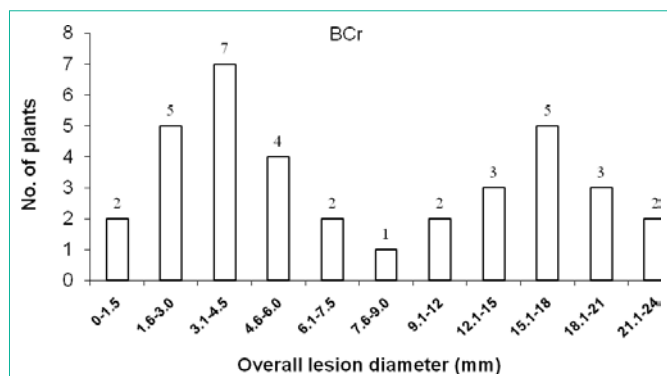


Figure 3: Distribution of disease incidence in BC<sub>r</sub> populations derived from the cross 'F<sub>1</sub>' and 'Comilla-2'.

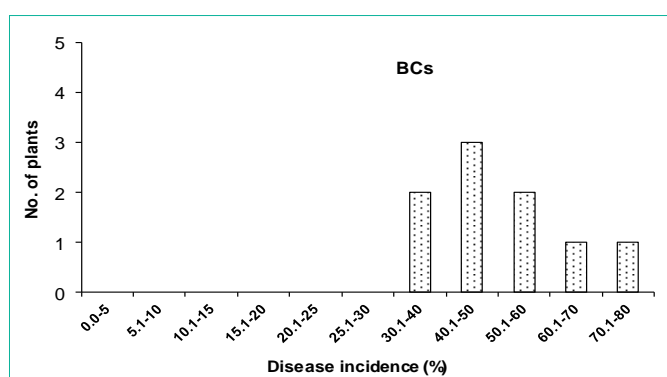
indicates that the resistant parent 'Comilla-2' bearing a recessive gene that conferring the resistant to anthracnose.

The distribution of disease incidence in the F<sub>2</sub> population was skewed toward the susceptible parent (Figure 1) and the distribution of overall lesion diameter in the F<sub>2</sub> population showed a similar trend (Figure 2). To determined the criteria of resistant and susceptibility, we scored the disease reactions of segregating populations using disease indices and overall lesion diameter. As a result, the distribution of disease incidence and overall lesion diameter in F<sub>2</sub> and BC<sub>R</sub> populations was divided using the scale of 25.0% and 9.0 mm respectively (Figure 1,2). Based on the scale of resistance and susceptibility, less than 25.0% disease incidence or less than 9.0 mm overall lesion diameter were evaluated as resistance.

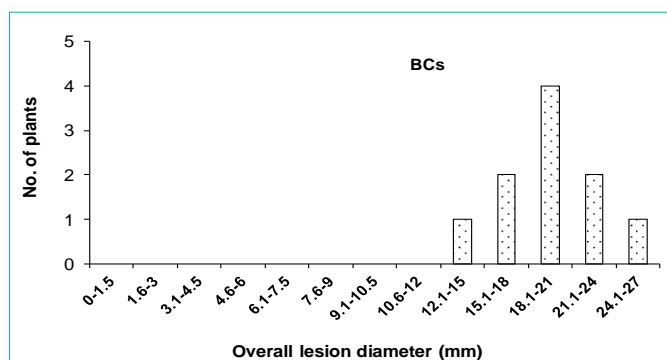
In the cross 'BARI chilli-1' x 'Comilla-2', the segregation of resistance and susceptibility scored by disease incidence in the F<sub>2</sub> population was 18 to 66 (Table 2). The chi-squared and P values in the



**Figure 4:** Distribution of overall lesion diameter in BCr populations derived from the cross 'F<sub>1</sub>' and 'Comilla-2'.



**Figure 5:** Distribution of disease incidence in BCs (Back cross with susceptible parent) populations derived from the cross 'F<sub>1</sub>' and 'BARI chilli-1'. Distribution was skewed to susceptible parent.



**Figure 6:** Distribution of overall lesion diameter in BCs (Back cross with susceptible parent) populations derived from the cross 'F<sub>1</sub>' and 'BARI chilli-1'. Distribution was skewed to susceptible parent.

F<sub>2</sub> population were 0.571 and 0.50-0.30, respectively, which fitted one recessive gene model i.e. 1:3 Mendelian model. The segregation ratios in the BCr and BCs populations in case of resistant and susceptibility was 16:20 and 0:10, respectively (Table 2 and Figure 3 & 4), which fitted expected segregation ratios, 1:1 and 0:1, respectively (Table 2). The segregation of resistance and susceptibility scored by overall lesion diameter in the F<sub>2</sub> population was 23 to 61 (Table 3). The chi-squared and P values in the F<sub>2</sub> population were 0.254 and 0.70-0.50, respectively, which fitted one recessive gene model i.e. 1:3 Mendelian model. Segregation ratios in BCr and BCs populations were 21:15 and

**Table 1:** Disease incidence and overall lesion diameter of 'Comilla-2', 'BARI chilli-1' and their F<sub>1</sub> progenies 8 days after inoculation with *C. capsici*.

Population	Disease incidence	Overall lesion diameter (mm)
Comilla-2	15.20 ± 1.96	3.42 ± 0.19
BARI chilli-1	67.10 1.67	18.74 ± 0.94
F <sub>1</sub> (BARI-1 x Comilla-2)	46.70 ± 6.87	13.2 ± 3.12

<sup>a</sup>Mean ± standard deviation

**Table 2:** Segregation ratio of resistance and susceptibility scored by disease incidence in segregating populations derived from the cross 'Comilla-2', and 'BARI chilli-1'.

Population	Expected ratio (R : S)	Observed frequency		X <sup>2</sup>	Pprobability
		R	S		
Disease incidence <sup>a</sup>					
Comilla-2		5	0		
BARI chilli-1		0	5		
F <sub>1</sub> (Bari-1 x Comilla-2)	1 : 3	0	5	0.571	0.50 - 0.30
F <sub>2</sub>	1 : 1	18	66	0.444	0.70 - 0.50
BCr	0 : 1	16	20		
BCs		0	10		

<sup>a</sup>Less than 25% disease incidence was evaluated as resistance (R).

<sup>b</sup>Less than 9.0 mm overall lesion diameter was evaluated as resistance BC<sub>R</sub>-F<sub>1</sub> back crossing with resistant parent ('Comilla-2') BC<sub>S</sub>-F<sub>1</sub> back crossing with susceptible (S) parent ('BARI chilli-1')

**Table 3:** Segregation ratio of resistance and susceptibility scored by overall lesion diameter in segregating populations derived from the cross 'Comilla-2' and 'BARI chilli-1'.

Population	Expected ratio (R:S)	Observed frequency		X <sup>2</sup>	Pprobability
		R	S		
Overall lesion diameter <sup>b</sup>					
Comilla-2		5	0		
BARI chilli-1		0	5		
F <sub>1</sub> (Bari-1 x Comilla-2)		0	5		
F <sub>2</sub>	01:03	23	61	0.254	0.70-0.50
BCr	01:01	21	15	1	0.50-0.30
BCs	00:01	0	10		

<sup>a</sup>Less than 25% disease incidence was evaluated as resistance.

<sup>b</sup>Less than 9.0 mm overall lesion diameter was evaluated as resistance.

0:10 respectively (Table 3 and Figure 5, 6). This result was also fitted to the one recessive gene model.

The distribution of segregating population and results of chi-squared tests in the cross 'BARI chilli-1' x 'Comilla-2' indicate that the resistance of 'Comilla-2' to *C. capsici* is controlled by a single recessive gene. The continuous distribution of disease incidence was displayed in all of the segregating populations, which indicates that minor genes may have affected the resistance. Also, in the crosses, F<sub>1</sub> plants showed somewhat different responses depending on the susceptible parent.

In several reports on the inheritance of resistance to *C. acutatum* or *C. gloeosporioides* in chili peppers resistance was inherited dominantly [4,5,9]. [1] showed that the resistance of *C. chinense* Jacq. 'PBC 932' to *C. capsici* inherited through a single recessive

gene. [16] reported that the resistance to *C. acutatum* was inherited through a single recessive gene. [6] suggested that the resistance of chilli germplasm 'Daepoong-cho' to *C. capsici* is controlled by a single recessive gene. Therefore, the results of the present investigation are in conformity with the findings of previous reports.

It was reported that for pepper breeding perspective, dominant resistance is more useful than recessive resistance because it will be manifested in F1 hybrids even if only one parent has the allele additionally. In case of dominant resistance, hybrid production is easier as compared to resistance conferring with recessive gene. Producing F1 varieties using recessive resistance sources requires much time and effort. However, recessive resistance is more durable than dominant resistance. This information can benefit chilli pepper breeding programs in the production of anthracnose-resistant varieties.

## Competing Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be interpreted as a potential conflict of interest.

## Author's Contribution

Md. Siddiqur Rahman was involved in the execution of the research work; collection, analysis and interpretation of the data; manuscript writing, etc. Prof. Abdul Mannan Akanda was assist during work and editing of the manuscript.

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