

Mini Review

Arguments for Caveolin-1 Knockout Mice as an Alzheimer's Disease Model

Dudau M¹, Frumosu M¹, Codrici E², Tanase C^{2,3} and Ana-Maria E^{1,2*}

¹"Carol Davila" University of Medicine and Pharmacy, Romania

²"Victor Babeş" National Institute of Pathology, Romania

³Department of Medicine, "Titu Maiorescu" University, Romania

*Corresponding author: Ana-Maria E, Carol Davila "University of Medicine and Pharmacy, 8 Eroilor Sanitari, 050474 Bucharest 5, Romania

Received: March 18, 2016; Accepted: June 02, 2016;

Published: June 04, 2016

Abstract

Neurodegenerative diseases, such as Alzheimer's Disease, are now acknowledged to be multifactorial pathologies, explaining the failure to come up with specific targeted treatments. New pathogenic links should be addressed in order to integrate the molecular interactions into a comprehensive mechanism. Caveolin-1 is a scaffolding membrane protein of caveolae – membrane microdomains involved in cell signaling, senescence and cell growth. Increased caveolin-1 expression was related to inducement of senescence and was also reported in the aged brain. However, cav-1 KO mice express AD- like alterations in hippocampus and neurologic abnormalities and incongruent reports in AD patients or AD models in this review we present the involvement of caveolin-1 in aging, with emphasis on aging brain and propose several mechanisms of interaction between caveolin-1 and amyloid precursor protein, the main pathogenic link of Alzheimer's disease. Also, in view of reports of synaptic plasticity deficits upon Cav-1 KO and its involvement in post-injury reactive neuronal plasticity, we propose Cav-1 to be neuroprotective and increased as a compensatory mechanism, rather than a direct measurement of aging process.

Keywords: Amyloid precursor protein; Caveolin-1; Alzheimer's disease; Cav-1 knockout mouse; Aging brain

Introduction

"Neurodegeneration" is a term which can be translated into "loss of neurons" accompanied by clinical features related to cognition and affect. However, high variability in terms of histopathologic changes found in patients with the same clinical diagnosis led to a thorough investigation of molecular causes. The limit between "normal" aging and neurodegeneration was even more so difficult to delineate from the perspective of "cognition reserve" – the ability of some subjects to counteract neuronal or synaptic loss by "using pre-existing cognitive processing approaches or by enlisting compensatory approaches" [1]. The most frequent causes of neurodegeneration are vascular dementia, Alzheimer's and Parkinson's diseases, each with its own molecular hallmarks. Alzheimer's disease diagnostic criteria include extraneuronal amyloid plaques and intraneuronal hyperphosphorylated tau tangles, but other molecular factors may contribute [2]. Parkinson's disease is characterized by progressive loss of dopaminergic neurons, with yet unknown, but apparently multifactorial molecular basis [3]. Vascular dementia is also a multifactorial entity [4]. However, molecular research results reported a considerable overlapping between these entities, with two important consequences: 1) the need for a more accurate diagnosis, which led to search of new biomarkers; 2) new disease models emerged. Animal models still hold an important role for mechanism elucidation, as they allow assessment of apparently non-related effects of the molecular defect imposed on the animal. Classical

neurodegeneration mouse models include, among others, transgenic animals for mutated amyloid precursor protein, mutated tau protein, mutated presenilin enzyme, injection of neurotoxins in the corpus striatum, ligation of carotid arteries. Caveolin-1

is a membrane protein responsible for scaffolding a membrane microdomain called "caveola" (pl. caveolae). Caveolae are considered signaling nodes and caveolin-1 over expression was related to cellular aging [5]. Caveolin-1 knockout mouse is a well established model for endothelial dysfunction [6]. In this review we will present this animal as a potential AD- like disease model, with arguments related to stem cells, amyloid precursor protein and metabolic and signaling particularities in neurons.

The concept of brain aging

"Brain aging" concept referred at the beginning to a progressively deteriorating performance. Studies on aged animals [7] from more than three decades ago reported neuronal loss with aging, along with a decreased volume of gray matter. Later reports challenged previous data, showing preserved neuronal number, despite cortex thinning in human brain [8], and were soon followed by confirming studies on animals [9-12]. Modern imaging methods, from computer tomography analysis in the early 80's [13] to MRI and fluorodeoxyglucose PET analyses [14,15], demonstrated that brain atrophy does occur with age, in healthy, non-demented elderly. The modifications affect both grey and white matter, but the loss is rather functional than cellular, more like defective circuitry, rather than neuronal loss. Cell preservation in aging was reported even in areas susceptible to dementia, such as frontal and medial temporal cortex, in which thinning is not always indicative of disease. Rather, instead of neuronal loss, a 3D

neuronal network loosening would account for frontal and temporal neocortical thinning [16]. A decrease in dendritic branching in animal [17] and human prefrontal cortex [18-20] supports this hypothesis. Surprisingly, hippocampal neurons, related to

neurogenesis and learning, do not alter their dendritic length, nor reduce their spine density in aged humans [21] or rats [22]. MRI data showed that white matter reduction is also constant in the aged human brain, possibly as an indicator of a defective myelination, strongly correlated hypertension and stroke [23]. However, is still under debate whether the presence of white matter lacunae yielded is significantly related to cognitive impairment [24]. Aging is characterized by a reduced neurogenesis, due to altered neurotrophin signaling.

Activation of senescence programmers within the niche, imbalanced growth factor signaling. Although brain resides in a protected environment, isolate by the blood-brain barrier, there are evidence of blood-born aging factors to cross the blood brain barrier to negatively influence the neurogenesis [25]. Brain aging is also a “decrease in homeostatic reserve” [26] which affects, at different rates, different cell types that share a homeostatic balance. Cellular abilities to limit and buffer Reactive Oxygen Species (ROS), to sustain a protective response to cytotoxic stimuli, or to limit vicious circles such as inflammatory environments are diminished. DNA damage (some ROS-related), mitochondrial aging and decreased ATP reserves [27] and affected cellular calcium removal systems [28] add to neuronal vulnerability. Thus, understanding the aging process of nervous tissue is a more challenging task due to a more complex regulation, signaling and intercellular interactions [4].

Caveolin-1 in aging brain

Caveolins 1, 2 and 3, the scaffolding proteins of caveolae, have been related to cellular senescence for more than ten years [29], changes in their expression were interpreted either as determinants, either as effectors of the aging process. Main function of caveolin-1 has been as proposed to be of signaling node, therefore cav-1, its family members (caveolins 2 and 3) and associated proteins (cavins) would select which signals are to be transmitted into cells. New data revealed that cav-1 is involved in the regulation of many cellular processes relevant to cell biology such as growth, migration, control of mitochondrial antioxidant levels and senescence [30]. Senescent cells express increased levels of caveolins [31] and *in vitro* over expression of cav-1 induced an early senescence in different cell types [32-34]. Contradictory, cav-1 KO animal models showed reduced lifespan [35], paradox that was attributed to cav-1 function as tumor suppressor [36,37]. Cho et al. proposed the “Gate theory of aging”, when “gatekeeper molecules at the membrane level would play the prime role in determining the senescent phenotype”; caveolae and caveolins are suitable for this role due to their regulation of cell signaling, calcium storage and quantization of cross-talk between signaling cascades [29]. Plasma membrane composition also changes with age, including the cholesterol composition. Such changes could influence the expression and distribution of caveolins in caveolae. Different tissues age differently in terms of caveolins expression: cardiac muscle shows increased cav-1 in the fractions of membrane forming caveolae [38], unlike smooth muscle, which does not change levels of cav-1 and cavin-1 [39]; aged endothelial cells increase their levels of cav-1 [40]. Brain and nervous tissue have their own particularities in term of aging and caveolin content. Aged mice increase their cav-1 expression in the hippocampus, similar to cav-1 expression in hippocampal tissue in patients with Alzheimer’s disease [41]. Cerebellum does not change expression of cav-1 with

age or pathology. Down regulation of hippocampal caveolin-1 was related to reduce synaptic plasticity in aging and its increased expression could be interpreted as a compensatory mechanism [42]. Cav-1 is expressed in neurons, mostly in pyramidal neurons of the frontal motor cortex, but also in parietal cortices, CA1 layer, stratum oriens and stratum radiatum of hippocampus [43]. The protein is also present in glial cells, although no caveolae have been identified in either cell type. Over expressing cav-1 in neurons led to a decrease in primary neurite outgrowth and branching, but an increase in neurite density [44]. In glutamatergic neurons, cav-1 interacts with glutamate receptors. Treatment with glutamate, kainate and AMPA increased the expression of caveolin-1, suggesting that “activation of ionotropic receptors regulates neuronal expression of caveolin” [45].

Caveolin-1 knockout mouse model

After identification of caveolin-1 as the prime component of [46], a knockout mouse model was generated for the study of the protein function *in vivo*. Surprisingly, cav-1KO mice were viable but showed evidence of hyperproliferative and vascular abnormalities, consistent with the wide distribution of caveolae in endothelial cells throughout the body. First roles attributed to cav-1 were stabilization of caveolin-2 to caveolae, mediator for caveolar endocytosis of specific ligands, negative regulator of cell proliferation and eNOS activity inhibition in endothelial cells [47].

Although viable, aged cav-1-deficient mice display significantly lower body weights and were resistant to diet- induced obesity, as compared with wild-type controls mice, even on a high fat diet. Serum profiles of these animals showed normal insulin, glucose, and cholesterol levels, but severely elevated triglyceride and free fatty acid levels, especially in the post-prandial state [6]. They have, however drastically reduced insulin receptor protein levels (>90%), without any changes in insulin receptor mRNA levels [48]. This mouse model is also characterized by alterations in other signaling pathways than nitric oxide and insulin, such as Extracellular-Signal Regulated Kinase (ERK), calcium signaling [45], modified balance of pro-and anti-inflammatory cytokines [49]. Cav-1 KO mice have reduced brain weight and develop a number of neurological phenotypes, with motor and behavioral abnormalities, including muscle weakness, clasping, reduced activity, abnormal spinning and gait abnormalities, without neuronal loss [50]. This finding could be related to previously report synaptic loss, sharing the same mechanisms at neuronal plate. Also, as a membrane protein, cav-1 loss could count for less myelination, which is basically a glial cell membrane enwrapping around the axons.

Cav-1 and neurogenesis

An interesting approach regarding cav-1 involvement in nervous tissue homeostasis was reported starting from Neuronal Precursor Cells (NPC) from dentate gyrus and subventricular zone. Cav-1 KO mice showed increased number of newly formed neuroblasts than wild type of matching age, while *in vitro* knockdown of Cav-1 promoted oligodendroglial differentiation of NPCs via β -catenin expression [51]. In turn, cav-1 promoted differentiation of NPCs towards astroglial line [52]. Another cav-1 related way to modulate NPC proliferation was recently reported by Samarasinghe et al. A non-transcriptional glucocorticoid signaling pathway that operates via lipid-raft associated glucocorticoid receptors requires cav-1 to

alter NPCs proliferative capacity [53]. Cav-1 mediated signaling via GR could impact on development of NPCs by “regulating the degradation of cell cycle regulators or migration of differentiated cells derived from NPSCs to their final position in the cortex” [53].

Caveolin-1 in neurodegeneration and Alzheimer’s disease

Although most data reported until several years back associated increased cav-1 with ageing, more and more results stated a neuroprotective role of cav-1. Starting with the cav-1 KO phenotype, to reports of synaptic plasticity deficits upon cav-1 KO [42] and involvement in post-injury reactive neuronal plasticity [44], cav-1 seems to be neuroprotective. From this perspective, the increase of cav-1 in brain of aged animals or brains of AD patients could be interpreted as a compensatory mechanism and not a direct measurement of aging process. Caveolin-1 and amyloid precursor protein – putative mechanisms of cooperation APP is a transmembrane protein, which can be metabolized in two ways: a physiologic one, generating soluble neurotrophic fragments and a pathologic one, generating insoluble amyloid beta peptides (A β) that aggregate in the extracellular matrix, forming senile plaques in the AD brain. Membrane regions of high cholesterol content, such as caveolae, favor the pathologic pathway and generation of A β . It has been demonstrated that APP localizes preferentially in cholesterol rich- membrane microdomains [43]. Cav-1 KO mouse model exhibits AD characteristics, such as elevated A β deposition in the hippocampus, cerebrovascular changes and increased astrogliosis, with early onset [54].

From literature data, several *mechanisms* can be put forth, through which amyloid precursor protein expression or processing can be modified by cav-1:

1- Cav-1 presents a Cav Scaffolding Domain (CSD), which can interact with other membrane proteins, including APP, thus facilitating its proteolysis [41].

2- APP is preferentially expressed in cholesterol –rich domains, such as caveolae [43] and its amyloidogenic processing is favored by hypercholesterolemia [55]. Disruption of caveolae by cav-1 KO may redirect APP traffic towards lipid rafts (also cholesterol enriched) leading to increased extracellular A β peptides deposition. Furthermore, both enzymes involved in amyloidogenic processing are located in lipid

rafts: β secretase compartmentalizes in non-caveolar lipid rafts [56] and γ secretase in detergent-resistant membranes [57].

3- Over-expressed cav-1 in β -secretase expressing cells resulted in decreased A β production, suggesting a protective role by cav-1 [54].

4- Caveolae and cav-1 act as signaling nodes, regulating activation of various kinases. In turn, APP has eight phosphorylation sites in the Cterminal domain, potentially affected by the modification of cav-1 expression. Phospho-Thr668 is essential for its binding to Fe65 and its nuclear translocation possibly followed by induction of glycogen synthase kinase 3 β and tau phosphorylation [58].

5- Loss of cav-1 may alter phosphorylation of other proteins involved in APP trafficking: Mint1/X11 α is one of four neuronal trafficking adaptors that interact with APP. Src-related tyrosine phosphorylation of Mint1 regulates the destination of APP, restricting its distribution to distal neurites [59].

Conclusion

Neurodegeneration is a multifactorial process, not necessarily related to a pathological process. Conversely, pathological alterations may be present, without any clinical signs. Although there are numerous and well characterized animal models to study the most frequent neurodegenerative diseases, other models may unravel new pathogenic links. Caveolin-1 knockout mouse is emerging as a novel, non-mutational model, which, due to its endothelial dysfunctions, could be related to vascular dementia. However, recent data regarding amyloid precursor protein processing in these mice, may argue also in favor of Alzheimer disease model, providing a multifactorial dementia model.

Acknowledgment

This work was supported by a grant of the Romanian National Authority for Scientific Research and Innovation CNCS- UEFISCDI, project number PN-II-RU-TE 2014-4-1534.

References

1. Stern Y. Cognitive reserve in ageing and Alzheimer’s disease. *Lancet Neurol.* 2012; 11: 1006-1012.
2. KA J. Alzheimer’s Disease: Current Clinical and Neuropathologic Diagnostic Criteria. *Austin Alzheimer’s and Parkinson disease.* 2014; 1.
3. Robinson PA. Understanding the molecular basis of Parkinson’s disease, identification of biomarkers and routes to therapy. *Expert Rev Proteomics.* 2010; 7: 565-578.
4. Enciu AM, Constantinescu SN, Popescu LM, MureÅanu DF, Popescu BO. Neurobiology of vascular dementia. *J Aging Res.* 2011; 2011: 401604.
5. Caravia L, Dudau M, Gherghiceanu M, Tanase C, Enciu AM. Could caveolae be acting as warnings of mitochondrial ageing? *Mech Ageing Dev.* 2015; 146-148: 81-7.
6. Razani B, Combs TP, Wang XB, Frank PG, Park DS, Russell RG, et al. Caveolin-1-deficient mice are lean, resistant to diet-induced obesity, and show hypertriglyceridemia with adipocyte abnormalities. *The Journal of biological chemistry.* 2002; 277: 8635-8647.
7. Sturrock RR. A quantitative lifespan study of changes in cell number, cell division and cell death in various regions of the mouse forebrain. *Neuropathol Appl Neurobiol.* 1979; 5: 433-456.
8. Terry RD, DeTeresa R, Hansen LA. Neocortical cell counts in normal human adult aging. *Ann Neurol.* 1987; 21: 530-539.
9. Vincent SL, Peters A, Tigges J. Effects of aging on the neurons within area 17 of rhesus monkey cerebral cortex. *Anat Rec.* 1989; 223: 329-341.
10. Merrill DA, Roberts JA, Tuszyński MH. Conservation of neuron number and size in entorhinal cortex layers II, III, and V/VI of aged primates. *J Comp Neurol.* 2000; 422: 396-401.
11. Peters A, Leahu D, Moss MB, McNally KJ. The effects of aging on area 46 of the frontal cortex of the rhesus monkey. *Cereb Cortex.* 1994; 4: 621-635.
12. Gazzaley AH, Thakker MM, Hof PR, Morrison JH. Preserved number of entorhinal cortex layer II neurons in aged macaque monkeys. *Neurobiol Aging.* 1997; 18: 549-553.
13. Soininen H, Puranen M, Riekkinen PJ. Computed tomography findings in senile dementia and normal aging. *J Neurol Neurosurg Psychiatry.* 1982; 45: 50-54.
14. Scahill RI, Frost C, Jenkins R, Whitwell JL, Rossor MN, Fox NC. A longitudinal study of brain volume changes in normal aging using serial registered magnetic resonance imaging. *Arch Neurol.* 2003; 60: 989-994.
15. Mungas D, Harvey D, Reed BR, Jagust WJ, DeCarli C, Beckett L, et al. Longitudinal volumetric MRI change and rate of cognitive decline. *Neurology.* 2005; 65: 565-571.

16. Freeman SH, Kandel R, Cruz L, Rozkalne A, Newell K, Frosch MP, et al. Preservation of neuronal number despite age-related cortical brain atrophy in elderly subjects without Alzheimer disease. *J Neuropathol Exp Neurol.* 2008; 67: 1205-1212.
17. Grill JD, Riddle DR. Age-related and laminar-specific dendritic changes in the medial frontal cortex of the rat. *Brain Res.* 2002; 937: 8-21.
18. de Brabander JM, Kramers RJ, Uylings HB. Layer-specific dendritic regression of pyramidal cells with ageing in the human prefrontal cortex. *Eur J Neurosci.* 1998; 10: 1261-1269.
19. Uylings HB, de Brabander JM. Neuronal changes in normal human aging and Alzheimer's disease. *Brain Cogn.* 2002; 49: 268-276.
20. Dickstein DL, Kabaso D, Rocher AB, Luebke JI, Wearne SL, Hof PR. Changes in the structural complexity of the aged brain. *Aging Cell.* 2007; 6: 275-284.
21. Burke SN, Barnes CA. Neural plasticity in the ageing brain. *Nat Rev Neurosci.* 2006; 7: 30-40.
22. Rapp PR, Gallagher M. Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. *Proc Natl Acad Sci USA.* 1996; 93: 9926-9930.
23. Kennedy KM, Raz N. Pattern of normal age-related regional differences in white matter microstructure is modified by vascular risk. *Brain Res.* 2009; 1297: 41-56.
24. Kramer JH, Mungas D, Reed BR, Wetzel ME, Burnett MM, Miller BL, et al. Longitudinal MRI and cognitive change in healthy elderly. *Neuropsychology.* 2007; 21: 412-418.
25. Enciu AM. Stem cells in neurodegenerative diseases. Tanase C, Neagu M, editors. In: *Stem Cells between Regeneration and Tumorigenesis*: Bentham Publishing.
26. Toescu EC. Normal brain ageing: models and mechanisms. *Philos Trans R Soc Lond B Biol Sci.* 2005; 360: 2347-2354.
27. Brunk UT, Terman A. The mitochondrial-lysosomal axis theory of aging: accumulation of damaged mitochondria as a result of imperfect autophagocytosis. *Eur J Biochem.* 2002; 269: 1996-2002.
28. Toescu EC, Verkhratsky A. Parameters of calcium homeostasis in normal neuronal ageing. *J Anat.* 2000; 197 Pt 4: 563-569.
29. Cho KA, Park SC. Caveolin-1 as a prime modulator of aging: a new modality for phenotypic restoration? *Mech Ageing Dev.* 2005; 126: 105-110.
30. Baker N, Tuan RS. The less-often-traveled surface of stem cells: caveolin-1 and caveolae in stem cells, tissue repair and regeneration. *Stem Cell Res Ther.* 2013; 4: 90.
31. Wheaton K, Sampsel K, Boisvert FM, Davy A, Robbins S, Riabowol K. Loss of functional caveolae during senescence of human fibroblasts. *J Cell Physiol.* 2001; 187: 226-235.
32. Volonte D, Zhang K, Lisanti MP, Galbiati F. Expression of caveolin-1 induces premature cellular senescence in primary cultures of murine fibroblasts. *Molecular biology of the cell.* 2002; 13: 2502-2517.
33. Cho KA, Ryu SJ, Oh YS, Park JH, Lee JW, Kim HP, et al. Morphological adjustment of senescent cells by modulating caveolin-1 status. *J Biol Chem.* 2004; 279: 42270-42278.
34. Dai SM, Shan ZZ, Nakamura H, Masuko-Hongo K, Kato T, Nishioka K, et al. Catabolic stress induces features of chondrocyte senescence through overexpression of caveolin-1: possible involvement of caveolin 1-induced down-regulation of articular chondrocytes in the pathogenesis of osteoarthritis. *Arthritis Rheum.* 2006; 54: 818-831.
35. Park DS, Cohen AW, Frank PG, Razani B, Lee H, Williams TM, et al. Caveolin-1 null (-/-) mice show dramatic reductions in life span. *Biochemistry.* 2003; 42: 15124-15131.
36. Williams TM, Cheung MW, Park DS, Razani B, Cohen AW, Muller WJ, et al. Loss of caveolin-1 gene expression accelerates the development of dysplastic mammary lesions in tumor-prone transgenic mice. *Molecular biology of the cell.* 2003; 14: 1027-1042.
37. Capozza F, Williams TM, Schubert W, McClain S, Bouzahzah B, Sotgia F, et al. Absence of caveolin-1 sensitizes mouse skin to carcinogen-induced epidermal hyperplasia and tumor formation. *The American journal of pathology.* 2003; 162: 2029-2039.
38. Ratajczak P, Damy T, Heymes C, Oliviero P, Marotte F, Robidel E, et al. Caveolin-1 and -3 dissociations from caveolae to cytosol in the heart during aging and after myocardial infarction in rat. *Cardiovasc Res.* 2003; 57: 358-369.
39. Lowalekar SK, Cristofaro V, Radisavljevic ZM, Yalla SV, Sullivan MP. Loss of bladder smooth muscle caveolae in the aging bladder. *NeuroUrol Urodyn.* 2012; 31: 586-592.
40. Yoon HJ, Cho SW, Ahn BW, Yang SY. Alterations in the activity and expression of endothelial NO synthase in aged human endothelial cells. *Mech Ageing Dev.* 2010; 131: 119-123.
41. Gaudreault SB, Dea D, Poirier J. Increased caveolin-1 expression in Alzheimer's disease brain. *Neurobiol Aging.* 2004; 25: 753-759.
42. Liu Y, Liang Z, Liu J, Zou W, Li X, Wang Y, et al. Downregulation of caveolin-1 contributes to the synaptic plasticity deficit in the hippocampus of aged rats. *Neural Regen Res.* 2013; 8: 2725-2733.
43. Kang MJ, Chung YH, Hwang CI, Murata M, Fujimoto T, Mook-Jung IH, et al. Caveolin-1 upregulation in senescent neurons alters amyloid precursor protein processing. *Experimental & molecular medicine.* 2006; 38: 126-133.
44. Gaudreault SB, Blain JF, Gratton JP, Poirier J. A role for caveolin-1 in post-injury reactive neuronal plasticity. *Journal of neurochemistry.* 2005; 92: 831-839.
45. Head BP, Insel PA. Do caveolins regulate cells by actions outside of caveolae? *Trends Cell Biol.* 2007; 17: 51-57.
46. Rothberg KG, Heuser JE, Donzell WC, Ying YS, Glenney JR, Anderson RG. Caveolin, a protein component of caveolae membrane coats. *Cell.* 1992; 68: 673-682.
47. Razani B, Engelman JA, Wang XB, Schubert W, Zhang XL, Marks CB, et al. Caveolin-1 null mice are viable but show evidence of hyperproliferative and vascular abnormalities. *J Biol Chem.* 2001; 276: 38121-38138.
48. Cohen AW, Razani B, Wang XB, Combs TP, Williams TM, Scherer PE, et al. Caveolin-1-deficient mice show insulin resistance and defective insulin receptor protein expression in adipose tissue. *American journal of physiology Cell physiology.* 2003; 285: C222-235.
49. Codrici EPID, Mihai S, Enciu AM, Albuiescu R, Tanase C, Hinescu ME. Cytokines, chemokines and growth factors profile in caveolin-1 transgenic mice. 40th Congress of the Federation-of-European-Biochemical-Societies (FEBS) - The Biochemical Basis of Life; Berlin, GERMANY: FEBS JOURNAL. 2015; 186.
50. Trushina E, Du Charne J, Parisi J, McMurray CT. Neurological abnormalities in caveolin-1 knock out mice. *Behav Brain Res.* 2006; 172: 24-32.
51. Head BP, Peart JN, Panneerselvam M, Yokoyama T, Pearn ML, Niesman IR, et al. Loss of caveolin-1 accelerates neurodegeneration and aging. *PLoS One.* 2010; 5: e15697.
52. Refolo LM, Malester B, LaFrancois J, Bryant-Thomas T, Wang R, Tint GS, et al. Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol Dis.* 2000; 7: 321-331.
53. Dislich B, Lichtenthaler SF. The Membrane-Bound Aspartyl Protease BACE1: Molecular and Functional Properties in Alzheimer's Disease and Beyond. *Front Physiol.* 2012; 3: 8.
54. Hur JY, Welander H, Behbahani H, Aoki M, Frånberg J, Winblad B, et al. Active gamma-secretase is localized to detergent-resistant membranes in human brain. *FEBS J.* 2008; 275: 1174-1187.
55. Chang KA, Kim HS, Ha TY, Ha JW, Shin KY, Jeong YH, et al. Phosphorylation of Amyloid Precursor Protein (APP) at Thr668 regulates the nuclear translocation of the APP intracellular domain and induces neurodegeneration. *Molecular and cellular biology.* 2006; 26: 4327-4338.
56. Dunning CJ, Black HL, Andrews KL, Davenport EC, Conboy M, Chawla S, et

- al. Multisite tyrosine phosphorylation of the N-terminus of Mint1/X11alpha by Src kinase regulates the trafficking of amyloid precursor protein. *Journal of neurochemistry*. 2016; 137: 518-527.
57. Li Y, Luo J, Lau WM, Zheng G, Fu S, Wang TT, et al. Caveolin-1 plays a crucial role in inhibiting neuronal differentiation of neural stem/progenitor cells via VEGF signaling-dependent pathway. *PLoS one*. 2011; 6: e22901.
58. Li Y, Lau WM, So KF, Tong Y, Shen J. Caveolin-1 promote astroglial differentiation of neural stem/progenitor cells through modulating Notch1/NICD and Hes1 expressions. *Biochemical and biophysical research communications*. 2011; 407: 517-524.
59. Samarasinghe RA, Di Maio R, Volonte D, Galbiati F, Lewis M, Romero G, et al. Nongenomic glucocorticoid receptor action regulates gap junction intercellular communication and neural progenitor cell proliferation. *Proceedings of the National Academy of Sciences*. 2011; 108: 16657-16662.